Biochemical Studies on the Cardioprotective Effect of Squalene against Isoprenaline-induced Myocardial Infarction in Rats

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This study was designed to examine the cardioprotective effect of squalene against isoprenaline-induced myocardial infarction in male albino rats. Levels of diagnostic markers [troponin T, homocysteine], lipoproteins [apolipoprotein A1, apolipoprotein B, lipoprotein (a)] (in plasma), total cholesterol, lipid peroxides (in plasma and heart tissue), and endogenous non-enzymatic antioxidants [vitamin C and vitamin E] (in heart tissue) were determined. Supplementation of squalene at 2% level along with feed for 45 days significantly prevented the isoprenaline-induced elevation in the diagnostic markers in plasma of experimental group of rats. It exerted an antilipidemic action against isoprenaline-induced hypercholesterolemia by maintaining the levels of cholesterol and lipoprotein components at near normalcy. Squalene supplementation exhibited an antioxidant effect against isoprenaline-induced myocardial infarction by blocking the induction of lipid peroxidation. A tendency to prevent the isoprenaline induced reduction in the non-enzymatic antioxidants such as vitamin E and C was also observed. The results of the present study indicate that the cardioprotective effect of squalene might be ascribable to its antilipidemic, antioxidant and membrane stabilizing properties.

Key words: Squalene, myocardial infarction, cholesterol, lipid peroxidation

Cardiovascular disease is a major public health concern and leading cause of death all over the world. According to WHO reports, about 16.7 million people around the globe die of myocardial infarction each year. This is about one-third of all deaths globally (WHO, 2004). It is predicted that heart disease and stroke will become the leading cause of both death and disability worldwide by the year 2020, with the number of fatalities projected to increase to more than 20 million a year and to more than 24 million a year by 2030. It is well evident that developing countries like India are struggling to manage the impact of infectious diseases simultaneously with the growing

burden on society and health systems caused by non-communicable diseases such as myocardial infarction. Moreover, it is very much painful and serious concern to realize that myocardial infarction in India occurs 10 to 15 years earlier than in the west. It is also important to note that an increasing number of young Indians are falling prey to myocardial infarction and there are an estimated 45 million patients of coronary heart disease in India (Yusuf et al. 2001). As myocardial injury is irreversible in nature, most of the modern drugs available are effective in the prevention of spreading or dispersal of necrotic damage to the adjacent cells. So there is a need for search of new

cardioprotective agents, which could limit the myocardial injury by strengthening the heart muscle.

Squalene, an isoprenoid molecule present in deep-sea shark liver oil in higher quantities, has been reported to possess antioxidant (Ko et al. 2002) and membrane stabilizing properties (Ivashkevich et al. 1981). It is an intermediate metabolite in cholesterol metabolism and is secreted in human sebum, where it protects the skin from ultraviolet (UV) radiation (Kohno et al. 1995; Storm et al. 1993). Several experimental investigations demonstrated the detoxifying activities of the squalene against diverse chemicals such as hexachlorobiphenyl, hexachlorobenzene, arsenic, theophylline, phenobarbital and strychnine (Fan et al. 1996; Kamimura et al. 1992; Richter & Schafer, 1982). It has also been reported to possess anticarcinogenic activity against several carcinogens, including azoxymethane-induced colon cancer (Rao et 1998) nicotine-derived al. and nitrosaminoketone-(NMK) induced lung carcinogenisis (Smith et al. 1998). Its antiageing property has already been well established (Passi et al. 2002). Though the beneficial properties of squalene are promising and well studied, the cardioprotective effects of squalene have not yet been explored.

Myocardial infarction induced by isoprenaline [L- β - (3,4-dihydroxyphenyl)-aisopropylaminoethanol hydrochloride], a β adrenergic agonist has been reported to show many metabolic and morphologic aberrations in the heart tissue of the experimental animals similar to those observed in human myocardial infarction (Nirmala & Puvanakrishnan, 1996). It induces myocardial necrosis by a multiple step mechanism (Ravichandran *et al.* 1990). The primary disturbance of isoprenaline-induced myocardial infarction has been reported to enhance adenylate cyclase activity resulting in increased cAMP formation, which in turn would lead to increased lipid accumulation in the myocardium (Subhash *et al.* 1978). The administration of isoprenaline produces necrotic lesions and increased lipid peroxidation in the myocardium, which plays a crucial role in the pathogenesis of isoprenaline-induced myocardial infarction (Noronha-Dutra *et al.* 1984; Singal *et al.* 1982; Singal *et al.* 1983).

In the present study, an attempt has been made to assess the cardioprotective action of squalene in isoprenaline-induced myocardial infarction in rats by virtue of its hypolipidemic, antiperoxidative and membrane-stabilizing properties.

Materials & Methods

Isoprenaline, tetraethoxy propane and cholesterol were obtained from M/s. Sigma Chemical Company, St. Louis. MO, USA. Squalene (Specific gravity: 0.853; Refractive index: 1.493; Saponification Value: 30; Iodine value: 344; Boiling point: 240-245°C) was prepared from the shark liver oil of *Centrophorus* sp. caught in the Andaman waters (Farvin *et al.* 2004). All the other chemicals used were of analytical grade.

Male Wistar strain albino rats, weighing 120-150g were selected for the study. The animals were housed individually in polypropylene cages under hygienic and standard environmental conditions (28±2°C, humidity 60-70%, 12 h light/dark cycle). The animals were allowed a standard diet [M/s Sai Feeds, Bangalore, India] and water *ad libitum*. The experiment was carried out as per the guidelines of Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), New Delhi, India and approved by the Institute Animal Ethics Committee (IAEC).

Seven days after acclimatization, the animals were divided into four groups of 6

rats each. Group I and Group III animals were fed on standard diet with added coconut oil at 2% level for 45 days and Group II and Group IV animals were fed on standard diet with added squalene at 2% level for the same period. After 45 days feeding, the Group III and Group IV animals were intraperitoneally (i.p.) injected with isoprenaline [11mg (dissolved in physiological saline)/ 100g body weight/ day for 2 days] for the induction of myocardial infarction. Control animals (Group I and Group II) were i.p. injected with physiological saline alone for 2 days.

At the end of the experimental period, i.e., 24 h after last injection of isoprenaline, the experimental animals were sacrificed, blood was collected using sodium citrate as anticoagulant and the plasma separated was used for assay of various biochemical parameters. The heart tissue was excised immediately and washed with chilled isotonic saline. Accurately weighed heart tissue was homogenized in ice-cold 0.1M Tris-HCl buffer, pH 7.2 and centrifuged. The supernatant was used for further biochemical analyses.

Troponin T was estimated in plasma samples by electrochemiluminescence immunoassay "ECLIA" on Roche Elecsys 1010/ 2010 and Modular Analytics E170 (Elecsys module) immunoassay analyzer. Homocysteine (Hcy) concentration in plasma was assayed by Microtiter plate assay package (Diazyme Laboratories). Apolipoprotein A1, apolipoprotein B, lipoprotein (a) were estimated plasma samples in using immunoturbidimetric test kit from DiaSys Diagnostic Systems GmbH, Germany. Total cholesterol content in plasma and heart tissue was estimated by the method of Parekh & Jung (1970) after extracting total lipids by the method of Folch et al. (1957). Lipid peroxide content in plasma and heart

tissue was determined by the thiobarbituric acid (TBA) reaction as described by Okhawa *et al.* (1979). Endogenous antioxidants such as ascorbic acid (vitamin C) and α tochopherol (vitamin E) were determined in the heart tissue by the methods of Roe & Kuether (1943) and Baker *et al.* (1980) respectively.

Results were expressed as mean ± SD. Multiple comparisons of the significant ANOVA were performed by Tukey's multiple comparison test. A *p*-value <0.05 was considered as statistically significant. All data were analyzed with the aid of statistical package program SPSS 10.0 for Windows.

Results & Discussion

Troponins are regulatory proteins essential for contraction and relaxation processes in myocardium. In the present study, a significant (p<0.001) increase was observed in the level of troponin T in plasma of Group III isoprenaline administered rats as compared to that of Group I control animals (Fig 1). This is in accordance with earlier reported studies (Acikel *et al.* 2004). Since cardiac troponins are not normally found in blood, an increased level of troponin T in serum indicates myocyte injury and necrosis.

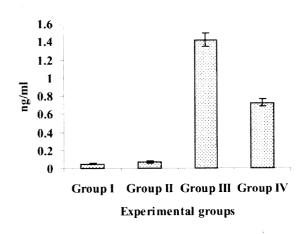


Fig 1. Level of Troponin T in plasma of normal and experimental groups of rats

Myocyte injury results in damage to contractile proteins and is a key mechanism responsible for the release of the structurally bound cardiac troponin T (Sarko & Pollack, 2002), and once outside the myocyte, these macromolecules are cleared from the interstitium by cardiac lymphatics. Eventually when the capacity of lymphatics to clear the macromolecules is exceeded, the markers become detectable in the peripheral circulation and are released with a stoichiometric relationship proportional to the amount of myocardial injury (Goldman et al. 2001). Recent years have witnessed an increased use of myocardial troponins as markers of myocardial injury. O" Brien et al. (1997) have shown that troponin T is a powerful biomarker in laboratory animals for sensitive and specific detection of cardiac injury.

Prior administration of squalene significantly (p<0.001) reduced the isoprenalineinduced release of troponin T from myocardium into the blood stream, thereby demonstrating its protective action on the cell membrane. It probably did so by maintaining the delicate balance of tonicity in cells in the myocardium. The presence of squalene in cell membrane is playing a major role in cell volume regulation by modulating the elasticity of plasma membrane. Cell volume

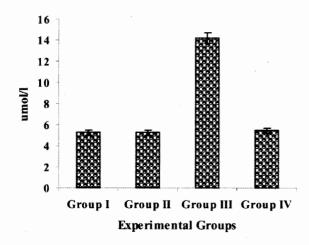


Fig 2. Level of homocysteine in plasma of normal and experimental groups of rats

affects the most basic processes of cell function, and as such it exerts an important role in the onset, severity, and outcome of myocardial infarction. Earlier reported studies indicate that squalene can overt severe osmolar changes associated with possible cell death (Ivashkevich *et al.* 1981).

In the present study, a significant (p<0.001) elevation in the level of homocysteine was noted in the plasma of Group III rats as compared to that of Group I control animals (Fig 2). This is in accordance with an earlier reported study (Hagar, 2002). Homocysteine is a thiol containing potentially cytotoxic 4-carbon α -amino acid formed during methionine metabolism. Recent prospective studies show that even mild hyperhomocysteinemia is associated with an increased risk of cardiovascular diseases independently of classical risk factors (Senaratne et al. 2000). Though the exact pathophysiological mechanism of homocysteine is still unclear, several experimental evidences are being reported in support of the involvement of homocysteine in the induction of myocardial infarction.

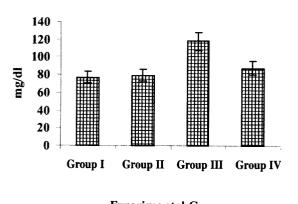
Homocysteine has been reported to induce atherosclerosis either by impairing coronary microvascular dilator function (Tawakol et al. 2002), or by stimulating smooth muscle proliferation (Tang et al. 1998), platelet activation, thrombogenesis (Rodgers and Kane, 1986), and endothelial dysfunction (Tsai et al. 1994). Both in vivo and *in vitro* studies suggest that homocysteine is a potent inducer of inflammatory processes in endothelial cells at the level of gene expression (Roth et al. 2001; Shai et al. 2004). Elevated level of homocysteine has also been reported to be associated with increased interleukin production in monocytes (Van Aken et al. 2000) and up regulation of vascular cell adhesion molecules (Silverman et al. 2002).

In this study, it is observed that the prior administration of squalene significantly (p<0.001) reduced the level of homocysteine in plasma of Group IV rats as compared to that of Group III myocardial infarction induced rats (Fig 2). It probably did so by inhibiting the production of monocyte/macrophage-derived interleukins, which triggers firm adhesion of rolling monocytes to vascular endothelium, a necessary prelude to the initiation of atherosclerosis (Gerszten et al. 1999). The HMG-CoA reductase inhibitors like lipophilic cerivastatin and fluvastatin have been reported to reduce the cardiovascular risk and vulnerability of atherosclerotic plaque through non-lipid mechanisms such as inhibition of interleukin expression (Ito et al. 2002). Since squalene is more lipophilic than statins, it is more permeable to vascular smooth muscle cells. Hence, it is possible that similar to prime HMG-CoA reductase inhibitor, squalene may also inhibit both homocysteine and interleukin production.

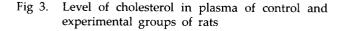
The present study reveals that animals treated with isoprenaline showed a significant (p<0.001) increase in the levels of total cholesterol in plasma and heart tissue as compared to that of normal control animals

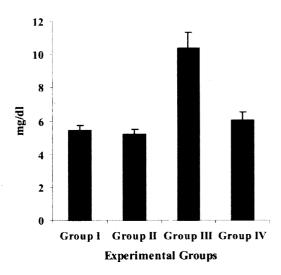
(Fig. 3 & 4), indicating the isoprenalineinduced hypercholesterolemic condition. Hypercholesterolemia has long been recognized as one of the major reversible risk factors for coronary heart disease. High level of circulating cholesterol and its accumulation in the heart tissue are well associated with cardiovascular damage (Joan et al. 1984; Buring et al. 1992). Our finding is in agreement with an earlier reported study (Ithayarasi & Devi, 1997b), which indicates that high levels of cholesterol in serum and heart of isoprenaline-treated rats is mainly responsible for altered cardiovascular functions. Also the levels of apolipoprotein B and lipoprotein (a) was found to be increased drastically with a concomitant decline in the apolipoprotein A1 in Group III isoprenalineadministered rats compared to group I control rats (Fig. 5 & 6). According to earlier reports apolipoprotein B and lipoprotein (a) were thought to be better predictors of acute myocardial infarction than total cholesterol and LDL-cholesterol (Sharrett et al. 2001; Graziani, et al. 1998).

Apolipoprotein A1 is primarily found in high-density lipoprotein (HDL) particle and serves the function of preventing the



Experimental Groups





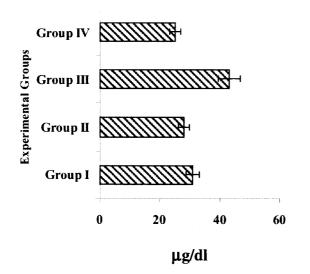


Fig 5. Level of lipoprotein (a) in plasma of control and experimental groups of rats

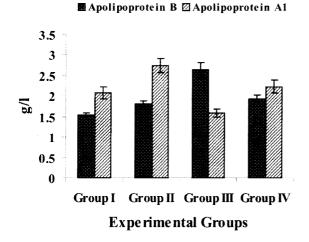


Fig 6. Level of apolipoprotein A1 and apolipoprotein B in plasma of control and experimental groups of rats

deposition of cholesterol-loaded macrophages on the arterial wall as foam cells, which is the prominent early feature of atherosclerotic lesion formation ultimately resulting in atherosclerosis. Over 90% of low density lipoprotein (LDL) particle is composed of apolipoprotein B. It can be used for assessing the cholesterol depositing capacity of the blood (Sehayek *et al.* 1994). Lipoprotein (a) is predominantly a genetic trait bound to both HDL and LDL and it is considered as a marker of atherosclerosisis (Simon *et al.* 1993) and has many properties in common with low-density lipoprotein (LDL). Isoprenaline mainly increases the low-density lipoproteins (LDL) cholesterol level in the blood, which in turn and leads to the build up of harmful deposits in the arteries, and thus favors induction of myocardial infarction (Goldstein and Brown, 1984). Increased cholesterol levels in the heart during myocardial infarction could be due to increased uptake of LDL from the blood by the tissues (Mathew *et al.* 1981). The abnormal cholesterol deposition is favored by the dangerous tendency of cholesterol to passive exchange between the plasma lipoproteins and the cell membranes (Brown and Goldstein, 1986).

In the present study, supplementation with squalene significantly (p<0.001) prevented the isoprenaline-induced elevation in the levels of total cholesterol, apolipoprotein B and lipoprotein (a) in Group IV rats as compared to that of Group III rats (Fig 5 & 6). Also the level of apolipoprotein A1 maintained at near normalcy. It probably did so by its hypocholesterolemic property.

Earlier Qureshi et al. (1996) have shown that squalene intake is effective in lowering the plasma total cholesterol, VLDL cholesterol and LDL cholesterol levels in experimental animals. In the present study also a slight reduction in the level of total cholesterol was observed in Group II squalene fed establishing animals, normal the anticholesterolemic property of squalene. The increase noticed in the apolipoprotein A1 also exhibits the beneficial action of squalene in preventing the cardiovascular complications. Dietary squalene was shown inhibit the activity of 3-hydroxy to methylglutaryl coenzyme A (HMG CoA) reductase, an endoplasmic reticulum (ER) protein, which catalyzes the rate-determining reductive deacylation of HMG CoA to mevlonate in cholesterol biosynthesis (Standberg et al. 1989). The

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hypocholesterolemic effect of squalene observed in the present study is probably related to its ability to modulate cholesterol metabolism either by regulating HMG CoA reductase activity through a feed back inhibition mechanism or by increasing cholesterol esterification process in the liver, as reported earlier by Khor & Chieng (1997). Further more oral supplementation of squalene has been previously reported to upregulate the fecal excretion of cholesterol and its non-polar derivatives as bile acid conjugates in experimental animals (Standberg *et al.* 1990; Nakamura *et al.* 1997).

Lipid peroxidation in vivo has been identified as one of the basic deteriorative reaction in cellular mechanisms of the myocardial ischemia (Singal et al. 1983). Injection of isoprenaline induced a significant (p<0.001) increase in the level of lipid peroxidation (Fig. 7) in the heart tissue of Group III isoprenaline-administered rats as compared to Group I rats. This result agrees with the findings of Nirmala & Puvanakrishnan, (1996b), which indicates that lipid peroxidation is also a key factor involved in the pathogenesis of isoprenalineinduced myocardial infarction. The enhanced lipid peroxidation in heart tissue of

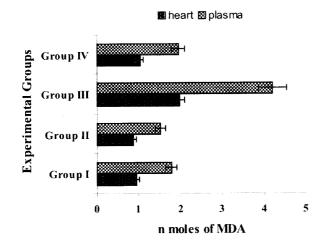


Fig 7. Levels of lipid peroxides in heart tissue and plasma of control and experimental groups of rats

isoprenaline-administered rats might have been due to the reduction in tissue antioxidant defense status, as reported earlier (Sushmakumari *et al.* 1989; Padma & Devi, 2002).

Lipid peroxidation of membranes is regulated by the availability of substrate in the form of PUFA, the availability of inducers such as free radicals and excited state molecules to initiate propagation, the antioxidant defense status of environment and the physical status of the membrane lipids (Anandan et al. 1998). Isoprenaline has also been proposed to be acting as a cardiotoxic agent due to its ability to destruct myocardial cells, possibly by a free radical mechanism (Singal et al. 1982). The reaction of isoprenaline-induced hydroxyl radicals (OH) with polyunsaturated lipids present in the myocardial cell membranes ultimately leads to the formation of lipid radicals and eventually to short-chain aldehydes and hydroxy alkenals. These events can be followed by the formation of conjugated dienes, malondialdehyde, and alkanes.

In the present study, the animals supplemented with squalene showed a significant (p<0.001) decrease in the level of lipid peroxidation in the plasma and heart tissue, thus indicating the antioxidant nature of squalene in experimentally induced oxidative stress condition. The antioxidant effect is probably due to the presence of isoprenoid unit in the structure of squalene. The unpaired electron present in the hydroxyl radical (OH) generated during isoprenaline-induced myocardial infarction might have been trapped for dismutation by its free radical scavenging isoprenoid unit.

A significant (p<0.001) decline was observed in the content of non-enzymic antioxidants, ascorbic acid and α -tocopherol, in the heart tissue of Group III isoprenalineadministered rats as compared to Group I control rats (Fig 8). This is in line with an earlier reported study (Senthil et al. 2004). Vitamin C is the primary antioxidant in plasma and cells and is the first antioxidant to be depleted under conditions of oxidative stress. It prevents oxidative modification of both the cytosolic and membrane components of the cells. Not only can the vitamin C directly quench superoxide radicals in aqueous milieu before they can attack lipids, but it can also reduce formation of tocopheroxyl radicals and so inhibits free radicals formation in the lipid phase (Frei et al. 1989). Vitamin E is a prominent membrane constituent of cardiac muscle, which halts lipid peroxidation by acting as a peroxyl radical trapping, chain-breaking antioxidant (Hensley et al. 2004). It is the only endogenous lipid soluble vitamin stabilizing the lipid bilayer of cell membranes, where it interacts with phospholipases to reduce membrane rearrangements (Schafer et al. 2002). It plays a major role in maintaining the structural integrity of cell membranes by limiting lipid peroxidation by reative oxygen species (Azzi et al. 2003). As

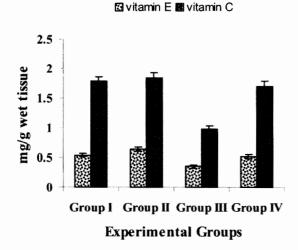


Fig 8. Levels of ascorbate and α-tocopherol in heart tissue of control and experimental groups of rats

long as the concentrations of redox cycling antioxidants, ascorbate and glutathione are maintained in the myocardium, distal antioxidant system would not be consumed. The reduction noted in the levels of these antioxidant vitamins in isoprenaline-induced myocardial infarction condition might be due to increased utilization of these vitamins for the removal of free radicals produced during isoprenaline induced oxidative stress. The generation of free radicals in isoprenaline-induced myocardial infarction might have exceeded the ability of these free radical scavenging non-enzymic antioxidants to dismute the radicals, resulting in membrane damage and reduction of scavengers.

It is worth noting that, the prior administration of squalene at 2% level along with feed significantly (p<0.001) reduced the isoprenaline-induced decline in the levels of these vitamins in Group IV animals as compared to those of Group III isoprenalineinjected rats. It probably did so by sharing the responsibility of these antioxidant vitamins in counteraction of free radicals generated during isoprenaline-induced oxidative stress.

 $LOO' + SQ - \rightarrow LOOH + SQ'$

Where LOO denotes a lipid peroxy radical, SQ denotes squalene.

SQ consists of six 2-methyl-2-pentene units and the electron donating property of the methyl group at the 2-position is likely to play an important role in the quenching activity. In fact, the replacement of hydrogen by methyl group in ethylene results in the reduction of the ionization potential. In SQ, the electron donating property of methyl groups bonded to quaternary carbons will be essential to the small ionization potential. Further more, the methyl groups also supply hydrogen for the ene reaction (Gilbert &

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Baggott, 1991), which may facilitate the quenching of singlet oxygen. Molecules containing a structure such as C-C (CH3) =-CH-C are expected to have large quenching activity.

In conclusion, the results of the present study indicate that the cardioprotective effect of squalene against isoprenalineinduced myocardial infarction may probably be related to its hypercholesterolemic property, to the counteraction of free radicals by its antioxidant nature, to the strengthening of myocardial membrane by its membrane stabilizing action, or to its ability to maintain near to the normal level of endogenous antioxidants like vitamin E, and vitamin C, which protects myocardial membrane against oxidative damage by decreasing lipid peroxidation.

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References

- Acikel, M., Buyukokuroglu, M.E., Erdogan, F., Aksoy, H., Bozkurt, E. and Senocak H. (2005). Protective effects of dantrolene against myocardial injury induced by isoprenaline in rats: biochemical and histological findings. *Int J Cardiol.* 98, 389-394
- Anandan, R., Devi, K.P., Devaki., T. and Govindaraju, P. (1998). Preventive effects of *Picrorhiza kurroa* on D-galatosamine– induced hepatitis in rats. J. Clin. Biochem. Nutr. 25, 87-95
- Azzi, A., Gysin, R., Kempna, P., Ricciarelli, R., Villacorta, L., Visarius, T. and Zingg, J. M. (2003). The role of alpha-tocopherol in preventing disease: from epidemiology to molecular events. *Mol Aspects Med.* 24, 325-336

- Baker, H., Frank, O., De Angelis, B. and Feingold, S. (1980). Plasma tochopherol in man at various times after ingesting free or acetylated tochopherol. *Nutr Rep Int.* 21, 531-536
- Brown, M.S. and Goldstein, J.L. (1986). A receptor mediated pathway for cholesterol homeostasis. *Science*. **232**, 34-47
- Buring, J.E., O'Connor, C.T. and Goldhaber, S.Z. (1992). Decreased HDL2 and HDL3 cholesterol, Apo. AI and Apo. A II and increased risk of myocardial infarction. *Circulation.* 85, 22-29
- Fan, S., Ho, I., Yeoh, F.L., Lin, C., Lee, T. (1996). Squalene inhibits sodium arsenite-induced sister chrmatid exchanges and micronuclei in Chinese hamster overy-K cells. *Mutat Res.* 368, 165-169
- Farvin, K.H.S., Anandan, R., Kumar, S.H.S., Shiny, K.S., Sankar, T.V., Thankappan, T.K. (2004). Effect of squalene on tissue defence system in isoprenaline-induced myocardial infarction in rats. *Pharmacol Res.* 50, 231-236
- Folch, J., Lees, M. and Stanely, G.H.S. (1957). A simple method for isolation and purification of total lipids from animal tissues. J. Biol. Chem. 226, 497-509
- Frei, B., England, L. and Ames, B.N. (1989) Ascorbate is an outstanding atioxidant in human blood plasma. *Proc Natl Acad Sci USA*. 88, 6377-6381
- Gerszten, R.E., Garcia-Zepeda, E.A., Lim, Y.C., Yoshida, M., Ding, H. A., Gimbrone, M., Luster, A.D., Luscinskas, F.W. and Rosenzweig, A. (1999). MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature.* 398, 718-723
- Gilbert, A. and Baggott, J. (1991). Essentials of Molecular Photochemistry, Blackwell, Oxford. pp 512-516
- Goldman, B., Chrisyenson, R.H., Hamm, C.W., Meinertz, T. and Ohman, E.M. (2001). Implications of troponin testing

in clinical medicine. Curr Control Trials Cardiovasc Med. 2, 75-84

- Goldstein, J.L. and Brown, M.S. (1984). Progress in understanding the LDL receptor and HMG CoA reductase to membrane protein that regulate the plasma cholesterol. *J. Lipid Res.* **25**, 1450-1460
- Graziani, M.S., Zanolla, L., Righetti, G., Marchetti, C., Mocarelli, P. and Marcovina, S.M. (1998). Plasma apolipoproteins A-I and B in survivors of myocardial infarction and in a control group. *Clinical Chemistry*. 44, 134-140
- Hagar, H.H. (2002). Folic acid and vitamin B(12) supplementation attenuates isoprenaline-induced myocardial infarction in experimental hyperhomocysteinemic rats. *Pharmacol. Res.* **46**, 213-219
- Hensley, K., Benaksas, E.J., Bolli, R., Comp, P., Grammas, P., Hamdheydari, L., Mou, S., Pye, Q.N., Stoddard, M.F., Wallis, G., Williamson, K.S., West, M., Wechter, W.J. and Floyd, R.A. (2004). New perspectives on vitamin E: gamma-tocopherol and carboxy elthyl hydroxy chroman metabolites in biology and medicine. *Free Radic Biol Med.* 36, 1-15
- Ithayarasi, A.P. and Devi, C.S. (1997). Effect of a-tocopherol on isoprenaline induced changes in lipids and lipoprotein profile in rats. *Indian J. Pharmacol.* **29**, 399-404
- Ito, T., Ikeda, U., Yamamoto, K., Shimada, K. (2002). Regulation of interleukin-8 expression by HMG-CoA reductase inhibitors in human vascular smooth muscle cells. *Atherosclerosis.* 165, 51-55
- Ivashkevich, S.P., Apukhovskaia, L.I. & Vendt, V.P. (1981). Effects of sterols having different chemical structure and squalene on osmotic resistance of erythrocytes. *Biokhimiia.* 46, 1420-1425
- Joan, F., Peter, Z., and Philip, D. (1984). Cholesterol, In. Clinical Chemistry on

Diagnosis and Treatments, (Amold, E. and Lyold, L., Eds), PG publishing, New Delhi, 230-256

- Kamimura, H., Koga, N., Ogari K. and Yoshimura, H. (1992). Enhanced elimation of theophylline, phenobarbital and strychnine from the bodies of rats and mice by squalene treatment. J Pharmacobio Dyn. 15, 215-221
- Khor, H.T. and Chieng, D.Y. (1997). Effect of Squalene, tocotrienols and a-tocopherol supplementations in the diet on serum and liver lipids in hamster. *Nutrition Research* 17, 475-483
- Ko, T.F., Weng, T.M. and Chiou R.Y. (2002). Squalene content and anti oxidant activity of *Terminalia catappa* leaves and seeds. *J. Agric. Food. Chem.* **50**, 5343-5348
- Kohno, Y., Egawa, Y. & Itoh, S. (1995). Kinetic study of quenching reaction of singlet oxygen and scavenging reaction of freeradical by squalene in n-butanol. *Biochem Biophys Acta*. **1257**, 52-56
- Mathew, S., Menon, P.V.G., and Kurup, P.A. (1981). Changes in myocardial and aortic lipids, lipolytic activity and fecal excretion of sterols and bile acids in isoprenaline-induced myocardial infarction in rats. *Indian. J. Biochem. Biophys.*, 18, 131-134.
- Nakamura, Y., Tonogai, Y., Tsumura, Y., Shibata, T. and Uchiyama, M. (1997).
 Effect of dietary squalene on the fecal steroid excretions and the lipid levels of serum and the liver in the rat. *Nutrition Research.* 17, 243-257
- Nirmala, C. and Puvanakrishnan, R. (1996). Protective role of curcumin against isoprenaline induced myocardial infarction in rats. *Mol. Cell Biochem.* **159**, 85-93.
- Noronha-Dutra, A.A., Steen, E.M., and Woolf, N. (1984). The correlation between catecholamine and lipid peroxidation induced in heart cells. *Basic Res. Cardiol.* 80, 133-136

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- O' Brien, P.J., Dameron, G.W., Beck, M.L. (1997). Cardiac troponin T is a sensitive, specific marker of cardiac injury in laboratory animals. *Lab Anim Sci.* 47, 486-495
- Ohkawa, H., Onishi, N., and Yagi, K. (1979). Assay for lipid peroxides in animal tissue by thiobarbituric acid reaction. *Anal. Biochem.* **95**, 351-358
- Padma, V.V. and Devi, C.S. (2002). Effect of fish oil on mitochondrial respiration in isoprenaline induced myocardial infarction in rats. *Indian J Exp Biol.* 40(3), 268-272
- Parekh, A.C. and Jung, D.H. (1970). Cholestrol determination with ferric acetate- uranil acetate and sulphuric acid – ferrus sulphate reagents. *Anal. Chem.* 42, 1423-1427
- Passi, S., De Pita, O., Puddu, P. and Littarru, G.P. (2002). Lipophilic antioxidants in human sebum and aging. *Free Radic. Res.* 36, 471-477
- Qureshi, A.A, Lehmann, J.W., and Peterson, D.M. (1996). Amaranth and its oil inhibit cholesterol biosynthesis in six week old female chickens. J. Nutrition. **126**, 1972-1978
- Rao, C.V., Newmark, H.L. and Reddy, B.S. (1998). Chemopreventive effect of squalene on colon cancer. *Carcinogenesis*. 19, 287-290
- Ravichandran, L.V., Puvanakrishnan, R. and Joseph, K.T. (1990). Alterations in the heart lysosomal stability in isoprenaline induced myocardial infarction in rats. *Biochem Int.* **22**, 387-396
- Richter, E. and Schafer, S.G. (1982). Effect of squalene on hexachlorobenzene (HCB) concentrations in tissue of mice. J Environ Sci Health. 17, 195-203
- Rodgers, G.M. and Kane, W.H. (1986). Activation of endogenous factor V by a

homocysteine-induced vascular endothelial cell activator. *J Clin Invest.* 77(6), 1909-1916

- Roe, H.G. and Kuether, C.A. (1943). Detection of ascorbic acid in whole blood and urine through the 2, 4-dinitrophenylhydrazine derivative of dehydroascorbic acid. J Biol Chem. 147, 399-407
- Roth, J., Goebeler, M., Ludwig, S. (2001). Homocysteine inhibits tumor necrosis factor-induced activation of endothelium via modulation of nuclear factorkappa b activity. *Biochim Biophys Acta*. 1540, 154–165
- Sarko, J., and Pollack, C.V. (2002). Cardiac Troponins. *The journal of emergency medicine.* 23, 57-65
- Schafer, F.Q., Wang, H.P., Kelley, E.E., Cueno, K.L., Martin, S.M. and Buettner, G.R. (2002). Comparing α-carotene, vitamin E and nitric oxide as membrane antioxidants. *Biol Chem.* 383(3-4), 671-681
- Sehayek, E. and Eisenberg, S. (1994). The role of native apolipoprotein containing lipoproteins in atherosclerosis: cellular mechanisms. *Curr Opin Lipidol.* 5, 350–353
- Senaratne, M.P., Griffiths, J. and Nagendran, J. (2000). Elevation of plasma homocysteine levels associated with acute myocardial infarction. *Clin Invest Med.* 23, 220–226
- Senthil, S., Veerapan, R.M., Ramakrishna Rao, M. and Pugalendi, K.V. (2004). Oxidative stress and antioxidants with cardiogenic shock complicating acute myocardial infarction. *Clinica Chimica Acta.* 348, 131-137
- Shai, I., Stampfer, M.J., Mab, J., Manson, J.E., Hankinson, S.E., Cannuscio, C., Selhub, J., Curhanc, G. and Rimma, E.B. (2004). Homocysteine as a risk factor for coronary heart diseases and its association with inflammatory biomarkers, lipids

and dietary factors. *Atherosclerosis*. 177, 375–381

- Sharrett, A.R., Ballantyne, C.M., Coady, S.A., Heiss, G., Sorlie, P.D., Catellier, D. and Patsch, W. (2001). Coronary Heart Disease Prediction From Lipoprotein Cholesterol Levels, Triglycerides, Lipoprotein(a), Apolipoproteins A-I and B, and HDL Density Subfractions. *Circulation.* 104, 1108-1113
- Silverman, M.D., Tumuluri, R.J., Davis, M. (2002). Homocysteine upregulates vascular cell adhesion molecule-1 expression in cultured human aortic endothelial cells and enhances monocyte adhesion. Arterioscler Thromb Vasc Biol. 22, 587–592
- Simon, D.I., Ezratty, A.M. and Loscalzo, J. (1993). Lipoprotein(a) and atherothrombosis. *Curr Opin Cardiol.* 8, 814–820
- Singal, P.K., Kapur, N., Dhillon, K.S., Beamish, R.E. and Dhalla, N.S. (1982). Role of free radicals in catecholamine induced cardiomyopathy. *Can. J. Physiol. Pharmacol.* 60, 1390-1397
- Singal, P.K., Beamish, R.E. and Dhalla, N.S. (1983). Potential oxidative pathways of catecholamines in the formation of lipid peroxides and genesis of heart disease. *Adv Exp Med Biol.* 161, 391-401
- Smith, T.J., Yang, G.Y., Seril, D.N., Liao, J., Kim, S. (1998). Inhibition of 4-(methyl nitrosamino) –1- (3pyridyl) –1- butanoneinduced lung tumerogenisis by dietary olive oil and squalene. *Carcinogenesis*. 19, 703-706
- Stansberg, T.E., Tilvis, R.S. and Miettinen, T.A. (1989). Effects of cholestyramine and squalene feeding on hepatic and serum plant sterol in the rat. *Lipids.* 24, 705-708
- Storm, H.M., Oh, S.Y., Kimler, B.F. and Norton, S. (1993). Radioprotection of mice by dietary squalene. *Lipids.* 28, 555-559

- Subhash, D., Narinder, K.K. and Nityanand, S. (1978). Effect of isoprenaline on lipid profile and cardiac enzymes in rats. *Indian J. Exp. Biol.* 16, 376-378
- Sushmakumari, S., Jayadeep, A., Suresh kumar, J.S. and Menon, P.V.G. (1989). Effect of carnitine on malanaldehyde, taurine and glutathione levels in heart of rats subjected to myocardial stress by isoprenaline. *Indian J. Exp. Biol.* 27, 134-137
- Tang, L., Mamotte, C.D., Van Bockxmeer, F.M. and Taylor, R.R. (1998). The effect of homocysteine on DNA synthesis in cultured human vascular smooth muscle. *Atherosclerosis*. 136(1), 169-173
- Tawakol, A., Forgione, M.A., Stuehlinger, M., Alpert, N.M., Cooke, J.P., Loscalzo, J., Fischman, A.J., Creager, M.A., Gewirtz, H., Tawakol, A., Forgione, M.A., Stuehlinger, M., Alpert, N.M., Cooke, J.P., Loscalzo, J., Fischman, A.J., Creager, M.A. and Gewirtz, H. (2002). Homocysteine impairs coronary microvascular dilator function in humans. J Am Coll Cardiol. 40(6), 1051-1058
- Tsai, J.C., Perrella, M.A., Yoshizumi, M., Hsieh, C. M., Haber, E., Schlegel, R. and Lee, M.E. (1994). Promotion of vascular smooth muscle cell growth by homocysteine: a link to atherosclerosis. *Proc Natl Acad Sci U S A.* 91(14), 6369-6373
- Van Aken, B.E., Jansen, J., Van Deventer, S.J. (2000). Elevated levels of homocysteine increase IL-6 production in monocytic Mono Mac 6 cells. *Blood Coagul Fibrin*olysis. **11**, 159–164
- WHO (2004). Atlas of Heart Disease and Stroke.
- Yusuf, S., Reddy, S., Ounpuu, S. (2001). Global burden of cardiovascular diseases: part I: general considerations, the epidemiologic transition, risk factors, and impact of urbanization. *Circulation* 104, 2746– 2753