

Effect of Intercropping on Inoculum Potential of Arbuscular Mycorrhizal Fungi in Agroforestry Systems

Anil Kumar, S. Hashmi, A. Shukla and O.P. Chaturvedi

National Research Centre for Agroforestry, Jhasni 284, India

Abstract: To access the effect of intercropping on inoculum potential of arbuscular mycorrhizal fungi in agroforestry systems, root and/or soil samples collected from *Albizia procera* (safed siris)-based, *Embllica officinalis* (aonla)-based and *Eucalyptus teriticornis* (eucalyptus)-based agroforestry systems were analyzed. Results on safed siris based agroforestry system indicated that intercropping in its plantation increased the inoculum potential of AM fungi, under and outside tree canopy. Among tested treatments, maximum inoculum potential was recorded in crop rotation of black gram + wheat (agroforestry plot), followed by green gram + mustard (agroforestry plot) and pure plantation of safed siris. In aonla-based agroforestry system, wheat as an inter-crop increased AM activity in aonla rhizosphere during crop period. The colonization index was significantly superior in aonla plants with wheat (28.4%) than in aonla plants grown without wheat (20.2%) during crop period and it was at par in both treatments, i.e. with (22.5%, 23.7%) and without wheat (25.6%, 24.3%) during pre-crop and post-crop periods. AM species composition did not show any qualitative changes in these observations, however quantitative changes were recorded. Similarly, colonization index of eucalyptus roots was significantly more in agroforestry plots (29.7%) than in pure plantation (13.7%). The observations are important for management of indigenous populations of AM fungi in agroforestry systems.

Key words: Agroforestry systems, AM fungi, inoculum potential, intercropping.

Arbuscular mycorrhizal (AM) fungi form symbiotic association with most economically important plants. These fungi improve plant growth under low fertility conditions, confer tolerance against plant pathogens, improve water uptake by plants, contribute to the formation of soil structure and also help plants to establish in new area (Panwar and Vyas, 2002). They are obligate symbionts and are non-specific in their selection of hosts (Bonfate-Fasolo, 1987).

Agroforestry is an intensive land use management system that maximizes production, gives supplement food and fodder, improves soil, utilizes waste and

degraded lands, provides employment opportunities, minimizes adverse effect of climatic factors, aids industrial growth and improves environment. Looking to the importance of agroforestry in sustainable agriculture and role of AM fungi, present study was conducted to assess the effect of intercropping on inoculum potential of AM-fungi in agroforestry systems.

Materials and Methods

The study was conducted at National Research Centre for Agroforestry, Jhasni (25°27'-N latitude, 78°35'-E longitude and at about 271 m above msl), which lies in the hot semi-arid zone with three distinct

Table 1. Effect of intercropping on propagules of AM fungi in siris (*Albizia procera*)-based agroforestry system

Month of sampling	Sampling location	Number of infective propagules of AM fungi per 100 g soil in intercrop			Mean Pooled mean	
		Black-gram+ wheat (agroforestry plot)	Green-gram+ mustard (agroforestry plot)	Siris	S.Em±	CD (0.05%)
July 2005	Under tree canopy	416.8*	557.6	201.8	392.1	275.1
	Outside tree canopy	416.8	529.6	180.8	375.7	248.0
	Mean	416.8	543.6	191.3	383.9	
August 2005	Under tree canopy	304.8	86.2	65.2	152.1	
	Outside tree canopy	217.0	92.4	51.6	120.3	
	Mean	260.9	89.3	58.3	136.2	
	Pooled mean	338.8	316.4	124.8		
					S.Em±	CD (0.05%)
Month of sampling					22.0	63.0
Tree crop combination					27.0	77.2
Sampling location					22.0	NS
Month of sampling * tree crop combination					38.2	109.1
Month of sampling * sampling location					31.2	NS
Tree crop combination * sampling location					38.2	NS
Month of sampling * tree crop combination * sampling location					54.0	NS

*Average of four replications.

seasons. The area receives annual rainfall between 700-1150 mm mostly during monsoon period (Mid June-September). The mean maximum temperature varies from 23°C (January) to 42°C (May). Corresponding mean minimum temperature varies from 5°C (January) to 26°C (May).

Results and Discussion

According to Sieverding (1991), indications about soil inoculum of AM fungi can be obtained by using data on spore enumeration, AM fungal mycelium biomass, root infection ratings, most probable number and the mycorrhizal inoculum potential. In the present study, observations on most probable number, root infection rating and spore enumeration were utilized for the

purpose. Root and/or soil samples were collected from safed siris (*Albizia procera*)-based, aonla (*Emblia officinalis*)-based and on eucalyptus (*Eucalyptus tereticornis*)-based agroforestry systems. In safed siris field, soil samples for most probable number analysis were collected from two locations (under and 5 m outside tree canopy) and three intercrop treatments, viz., black gram + wheat (agroforestry plot), green gram + mustard (agroforestry plot) and control (pure tree plantation) during July 2005 and August 2005. The trees were planted in 1998 and intercrops were being grown regularly in the experimental plots. In aonla field, root and soil samples from rhizosphere of aonla, with and without wheat as intercrop, were

Table 2. Colonization index and spore count of AM fungi in rhizosphere of aonla variety NA-7 with and without wheat as intercrop

Date of sampling	Colonization index in aonla (%)			AM spore count per 100 g soil in rhizosphere of aonla		
	With wheat	Without wheat	Mean	With wheat	Without wheat	Mean
Pre-crop period:						
July 2003	27.8* (31.8)#	27.5 (31.6)	27.7 (31.7)	22.5*	12.5	17.5
August 2003	17.6 (24.8)	23.8 (29.2)	20.7 (27.0)	8.8	10.0	9.4
October 2003	10.8 (20.1)	12.5 (20.7)	12.2 (20.4)	37.5	38.0	37.8
Mean	18.7 (25.6)	20.9 (27.2)		22.9	20.2	
Crop period:						
December 2003	36.6 (37.2)	22.2 (28.1)	29.2 (32.7)	19.0	12.8	15.9
March 2004	40.2 (39.3)	26.9 (31.2)	33.4 (35.3)	2.5	2.5	2.5
Mean	38.4 (38.3)	24.6 (29.7)		10.8	7.6	
Post-crop period:						
June 2004	23.0 (28.6)	26.4 (30.9)	24.7 (29.8)	35.5	29.5	32.5
August 2004	31.0 (33.8)	22.4 (28.2)	26.6 (31.0)	17.8	21.0	19.4
September 2004	23.1 (28.7)	28.3 (32.1)	25.6 (30.4)	8.5	8.5	8.5
October 2004	18.4 (25.4)	20.4 (26.8)	19.4 (26.1)	12.5	15.5	14.0
Mean	23.7 (29.1)	24.3 (29.5)		18.6	18.6	
			S.Em±	CD (0.05%)	S.Em±	CD (0.05%)
Pre-crop period:						
Treatment			1.8	NS	3.7	NS
Date of sampling			2.2	6.7	4.6	13.6
Interaction			3.2	NS	6.5	NS
Crop period:						
Treatment			2.4	7.5	1.8	NS
Date of sampling			2.4	NS	1.8	5.5
Interaction			3.5	NS	2.5	NS
Post-crop period:						
Treatment			2.3	NS	2.2	NS
Date of sampling			3.2	NS	3.0	8.9
Interaction			4.5	NS	4.3	NS

* Average of four replications; # Figures in parenthesis indicate angular transformation values.

Table 3. AM species composition of aonla rhizosphere with and without wheat as an intercrop during December 2003

AM species	Spore count per 100 g soil in aonla rhizosphere		
	With wheat	Without wheat	Mean
<i>Glomus</i> sp. I	6.5*	2.8	4.7
<i>Glomus</i> sp. II	0.8	3.3	2.1
<i>Acaulospora</i> sp.	0.3	0.5	0.4
<i>Gigaspora</i> sp.	0.3	2.0	1.2
Others	4.8	1.8	3.3
Total	12.5	10.3	

* Average of four observations.

collected from July 2003 to October 2004, for analysis of root colonization index, AM spore count and AM species composition. Aonla plants were seven years old and the wheat was grown in the agroforestry plots only during rabi 2003-2004. Root samples of *Eucalyptus* from pure plantation and agroforestry plots (crop rotation: black gram + wheat) were collected during April 2005 and June 2005 for observations on colonization index. The plantation was two years old and the intercrops are being grown in agroforestry plots on regular basis.

Number of infective propagules of AM fungi in soil samples were estimated by using most probable number method of Porter (1979). Mycorrhizal colonization percentage was determined by grid line intersection after clearing and staining root samples (Phillip and Hayman, 1970). Spores were isolated from soil according to the method of Gerdemann and Nicolson (1963) and counted. The data were statistically analyzed by ANOVA and the differences among means were tested by using critical difference (CD) values at 5% level of probability.

Results on safed siris (*Albizia procera*)-based agroforestry system indicated that

intercropping in its plantation increased the inoculum potential of AM fungi under and outside tree canopy. Among treatments, maximum inoculum potential was recorded in black gram + wheat-agroforestry plot (338.8 propagules/100 g soil) followed by green gram + mustard-agroforestry plot (316.4 propagules/100 g soil), which were at par. Lowest inoculum potential was observed in pure plantation of safed siris (124.8 propagules/100 g soil). The inoculum potential was significantly more in July 2005 (383.9 propagules/100 g soil) than in August 2005 (136.2 infective propagules/100 g soil). The values were at par under (275.1 propagules/100 g soil) and outside (248.0 propagules/100 g soil) tree canopy (Table 1). In aonla plants, colonization index was significantly superior with wheat (28.4%) than in aonla plants grown without wheat (20.2%) during crop period. While during pre-crop and post-crop periods, it was at par in both treatments, i.e. with (22.5%, 23.7%) and without wheat (25.6%, 24.3%). The observed differences can be explained on the basis of better moisture availability in agroforestry plot, as the intercrop was irrigated. The observations on AM spore counts per 100 g soil from rhizosphere of aonla plants grown with wheat were at par

with respective observations on aonla plants grown without wheat, during all above mentioned crop periods (Table 2). AM species composition did not show any qualitative changes in these observations, however quantitative changes were recorded (Table 3). Thus, the results showed that wheat as an inter-crop increased AM activity in aonla rhizosphere during crop period. Similarly, colonization index of *Eucalyptus* roots was significantly more in agroforestry plots (29.7%) than in pure plantation (13.7%). Differences among these values for April 2005 (20.7%) and June 2005 (20.5%) were non-significant.

Fallowing has been reported by many workers to reduce soil population of AM fungi (Barbara and Hetrick, 1984). Therefore, more extensive green cover and biological diversity in agroforestry systems as compared to pure block plantation may enrich the AM flora. Though AM fungi are obligate symbionts, but these are non-specific in their selection of host (Bonfante and Fasolo, 1987). Thus, possibility of cross infectivity among tree and crop components of a given agroforestry system exists, which is likely to improve soil AM microflora. Reid and Bowen (1979) have reported that maximum AM colonization of *Medicago* plants occurred at field capacity and it reduced under conditions, which were either too wet or too dry. Thus, the improved moisture conditions of agroforestry systems with irrigated intercrops like wheat may increase the population of AM fungi in soil compared to dry soils of pure plantation. Sieverding (1991) has reported that small phosphate applications may generally improve AM fungal activities on infertile soils and their population benefits from N application

probably due to longer maintenance of photosynthetic leaf area. In selected experimental fields in present study, the fertilizers were applied to intercrops and not to tree. So tree component must be receiving these at sub optimal level, which might have improved the AM flora under tree canopy. Similar observations have been made by Rajshekara *et al.* (1989) in agroforestry system. They studied the effect of cropping system on AM development in red gram and sunflower and reported that colonization and sporulation of AM fungi increased with decreasing distance from the tree (*Dalbergia sissoo*, *Dendrocalamus strictus*, *Tectona grandis*, *Leucaena leucocephala* and *Casurina equisetifolia*). *L. leucocephala* supported the highest mycorrhizal development in both crops.

References

- Barbara, A. and Hetrick, D. 1984. Ecology of VA mycorrhizal fungi. In *VA Mycorrhiza* (Eds. C.L. Powell and D.J. Bagyaraj), pp. 35-55. CRC Press, Florida, USA.
- Bonfante-Fasolo, P. 1987. Vesicular arbuscular mycorrhizal fungus plant interactions at cellular level. *Symbiosis* 3: 249-254.
- Geredmann, J.W. and Nicolson, T.H. 1963. Spores of mycorrhizal *Endogone* species extracted from soil by wet sieving and decanting. *Transactions of British Mycological Society* 46: 235-244.
- Panwar, J. and Vyas, A. 2002. Arbuscular mycorrhizal association in *Tamarix aphylla* in the Indian Thar Desert. *Mycorrhiza News* 14: 14-16.
- Phillip, J.M. and Hayman, D.S. 1970. Improved procedure for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Transactions of British Mycological Society* 55: 158-161.
- Porter, W.M. 1979. The most probable number method for enumerating infective propagules of vesicular arbuscular mycorrhizal fungi in soil. *Australian Journal of the Soil Research* 17: 515-519.

- Rajshekhara, E., Sreenivassa, M.N. and Bhat, R.S. 1989. Effect of cropping system on vesicular-arbuscular mycorrhizal development in red gram and sunflower. *Karnataka Journal of Agricultural Sciences* 2: 231-233.
- Reid, C.P.P. and Bowen, G.D. 1979. Effects of soil moisture on VA mycorrhiza formation and root development in *Medicago*. In *The Soil Root Interface* (Eds. J.L. Harley and R.S. Russell), p. 211. Academic Press, London.
- Sieverding, E. 1991. *Vesicular Arbuscular Mycorrhiza Management in Tropical Agrosystems*. Deutsche Gescellschaft fur Technische Zusammenarbeit (GTZ) BmbH, Postfach, Federal Republic of Germany.