

Water Relations, Carbon Exchange and Nitrogenase in Chickpea under Soil Water Deprivation

M S Kuhad, A S Nandwal, I S Sheoran and K D Sharma

Department of Botany, Haryana Agricultural University, Hisar - 125 004 India

Chickpea widely grown in the Arid and Semi-Arid Zones of India is characterized as drought tolerant but poor yielder. This crop is usually grown on conserved moisture and experiences drought during reproductive phase, because of decreased soil moisture and rise in temperature. There is also a competition for photosynthates between nodules and reproductive parts at this stage, which results in decline in N_2 fixation (Luthra *et al.* 1983). Since N_2 fixation and CO_2 exchange are important physiological processes for plant growth and yield, information is required to elucidate the exact mechanism of these processes under water stress. Therefore, it becomes imperative to study the response of these processes in relation to plant water status and under conditions of soil water deprivations.

Seeds of chickpea (*Cicer arietinum* L.) cv H-75-35, inoculated with *Rhizobium* culture (Ca-181) were raised in pots (dia 30 cm) filled with 5 kg of dune sand. Two plants per pot were supplied with N free nutrient solution at weekly interval. At the time of pod formation, i.e. 110 days after sowing, the pots were divided into two sets. One set maintained at field capacity, i.e. 50 % of soil saturation% (SSP) and soil moisture content (SMC) of $12 \pm 0.5\%$ served as control. In the second set irrigation was withheld and the plants were sampled at two days interval till the plants showed the permanent wilting (SMC = $2.0 \pm 0.5\%$). Half of severe stressed plants were irrigated and sampled after two days to see the recovery. Soil moisture content was determined gravimetrically. Fourth leaf from top was taken for measuring the leaf water potential (ψ_w) and osmotic potential (ψ_s) with pressure chamber (Model Hr-3005, Soil Moisture Equipment Corporation, USA) and Vapour Osmometer (Model 5100 B, Wescor Inc, USA), respectively.

the ψ_w and ψ_s of nodulated roots were measured. Photosynthesis and leaf respiration of intact plants were measured as described earlier (Luthra *et al.* 1983). Nitrogenase activity of nodules was determined by acetylene reduction assay (ARA). Leaf area was recorded on leaf area meter (Li-3000, Licor, USA).

Leaf water potential and osmotic potential decreased gradually with decrease in soil moisture (Table 1). Osmotic potential showed more negative value than water potential, thereby lowering the turgor. At -2.2 MPa leaf water potential, plant showed the sign of permanent wilting. Decrease in turgor potential of leaf resulted in marked reduction in leaf area (Table 1) except at SMC of 8 %. Nodulated root maintained higher water potential and osmotic potential than leaf. At permanent wilting nodulated root had more turgor than leaf. After recovery both the components showed better turgor than control. The greater decrease in osmotic potential of nodulated root under water stress seems to be due to accumulation of organic solutes (Nandwal 1989).

Nitrogen fixation is known to be sensitive to water stress (Venkateswarlu *et al.* 1989, Nandwal *et al.* 1991). Here also, the decrease in nodulated root water potential was accompanied by reduction in nitrogenase activity. Interestingly at leaf and nodulated root water potential of -0.42 MPa and -0.35 MPa respectively, nitrogenase activity was maximum. This might be due to better availability of photosynthates (Table 1) and O_2 diffusion. With further decrease in water potential, fixation declined and no activity was detected at permanent wilting. Chopra *et al.* (1984) reported that reduction in N_2 fixation was due to decreased supply of photosynthates, whereas Sprent (1981) held the view that this decrease is mainly due to the direct

Table 1 Water potential, osmotic potential, photosynthesis, respiration and nitrogenase activity in chickpea experiencing water deprivations.

Parameters	Days after withholding irrigation					
	Control	2	4	6	8	Recovery
	SMC = 12 ±0.5% at Field capacity (FC)	SMC = 8 ±0.5% 30% Less than FC	SMC = 5.5 ±0.5% 55% Less than FC	SMC = 4 ±0.5% 70% Less than FC	SMC = 2 ±0.5% 85% Less than FC	SMC = 12 ±0.5% at Field capacity
Leaf ψ_w (-MPa)	0.12	0.42	1.15	1.60	2.20	0.09
Leaf ψ_s "	1.26	1.25	1.40	1.81	2.25	1.27
Leaf ψ_p "	1.14	0.83	0.25	0.21	0.05	1.18
Nodulated root ψ_w (-MPa)	0.08	0.35	0.70	0.98	1.70	0.08
Nodulated root ψ_s (-MPa)	0.95	0.90	1.10	1.36	2.10	1.02
Nodulated root ψ_p (-MPa)	0.87	0.55	0.40	0.38	0.40	0.94
Photosynthesis (mg CO ₂ Plant ⁻¹ h ⁻¹)	30.4	31.2(3)	19.4(-37)	6.7(-78)	0.61(-98)	21.9(-31)
Respiration (mg CO ₂ Plant ⁻¹ h ⁻¹)	8.2	8.1(-1)	6.1(-27)	5.3(-36)	3.7(-55)	7.7(-6)
Nitrogenase activity (μ mole C ₂ H ₄ g ⁻¹ h ⁻¹)	9.5	11.0(10)	3.2(-67)	0.70(-93)	0.0(-100)	4.1(-56)
Leaf area (cm ² Plant ⁻¹)	877	892(2)	778(-10)	742(-14)	560(-36)	404(-54)

Values in parentheses are percent increase or decrease over control.

effect on O₂ diffusion in nodules. Durand *et al.* (1987) showed that factors other than photosynthate supply, became limiting to ARA much before a reduction in photosynthesis was observed.

In the present study, the reduction in photosynthesis was less than N fixation and recovery was also better for photosynthesis. This suggests that it is not only the photosynthates which controls the N fixation, but some other critical factors may also be involved, in which water potential of nodulated root is one of them, as reported earlier (Nandwal *et al.* 1991). Decrease in photosynthesis at decreasing leaf water potential was due to decrease in leaf area.

References

- Chopra RK, Koundal KR & Sinha SK 1984 A simple technique of studying water deficit effects on nitrogen fixation in nodules without influencing the whole plant. *Plant Physiology* **76** 254-256
- Durand JL, Sheehy JE & Minchin FR 1987 Nitrogenase activity, photosynthesis and nodule water potential in soybean plants experiencing water deprivations. *Journal of Experimental Botany* **38** 311-321
- Luthra YP, Sheoran IS & Singh R 1983 Ontogenetic interaction between photosynthesis and symbiotic nitrogen fixation in pigeonpea. *Annals of Applied Biology* **103** 549-556
- Nandwal AS 1989 *Physiological Studies on Pigeonpea (Cajanus cajan L. Mill) Under Water Stress*. Ph.D. Thesis, Haryana Agricultural University, Hisar, India
- Nandwal AS, Bharti S, Sheoran IS & Kuhad MS 1991 Drought effects on carbon exchange and nitrogen fixation in pigeonpea (*Cajanus cajan* L.). *Journal of Plant Physiology* **138** 125-127
- Sprent JI 1981 Nitrogen fixation. In: *Physiology and Biochemistry of Drought Resistance in Plants* (Eds. LG Paleg & D. Aspinall) Academic Press, New York, 131-143
- Venkateswarlu B, Maheshwari M & Saharan N 1989 Effect of water deficit on N₂ (C₂H₄) fixation in cowpea and groundnut. *Plant and Soil* **14** 69-74