Fishery Technology 2004, Vol. 41(2) pp : 149 - 152

Research Note

Isolation and Identification of Histamine-forming Enterobacteria in Freshly Landed Tuna (Euthynnus affinis) Using a Dichotomous Scheme

S.B. Patange*, M.K. Mukundan and S. Sanjeev

Quality Assurance and Management Division Central Institute of Fisheries Technology, Matsyapuri, Cochin - 682029, India

Key words: Histamine, enterobacteria, tuna

Histamine poisoning has been attributed mainly to the accumulation of toxic levels of histamine in a spoiling fish. Histamine is reportedly produced by a wide range of microorganisms, majority of which are gram-negative rods of *Enterobacteriaceae* family (Frank *et al.*, 1985; Taylor and Sumner, 1986; Klausen and Huss, 1987; Okuzumi *et al.*, 1994).

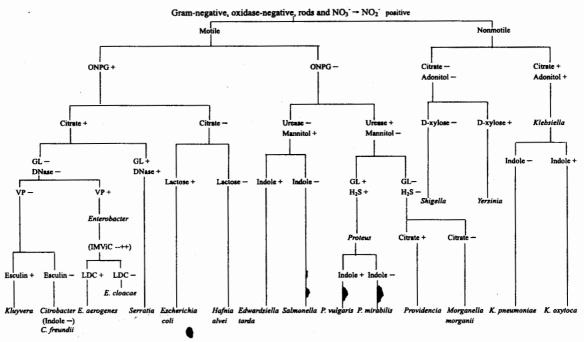
For the detection and enumeration of histamine forming bacteria (HFB), Niven's differential agar medium (Niven et.al., 1981) has been widely used (Baranowski, 1985; Chen et al., 1989; Roig-Sagues, 1997). Modification to Niven's medium has been reported for the detection of HFB in general with respect to changes in pH and histidine content (Yoshinaga and Frank, 1982; Chen et al., 1989; Mavromatis and Quantick, 2002). The positive isolates from Niven's medium are reported to be further characterized by using rapid identification kits such as API 20E test strips or PASCO Gram Negative Identification System (Frank et al., 1985; Lopez-Sabater et al., 1996). Several investigators have adopted conventional biochemical tests for characterization of these isolates (Subburaj et al., 1984; Gopakumar et al., 1988; Lakshmanan et al., 2002). Polymerase chain reaction techniques and DNA probes have

also been reported for the detection of HFB (Alves et al., 2002; Kim et al., 2003). The aim of this work was to formulate a dichotomous scheme with minimal number of conventional biochemical tests for the identification of HFB belonging to Enterobacteriaceae family detected on a differential agar medium.

The study was undertaken using fresh tuna (Euthynnus affinis) procured from Cochin fishing harbour. The aerobic plate count (APC) was estimated as per the standard method (Maturin and Peeler, 1995) using tryptone glucose agar. Violet red bile glucose agar (VRBGA) (HiMedia, Mumbai) was used for the enumeration and isolation of Enterobacteriaceae according to ICMSF (1978).

About 28 well-isolated purple colonies surrounded by a purple halo on a VRBGA plates giving 33 colonies were purified and then subjected to gram-staining, oxidase and nitrate reduction tests for provisional identification as *Enterobacteriaceae*. Motility and other conventional biochemical tests such as O/F test, indole production, MR-VP, citrate utilization, malonate utilization, phenylalanine deaminase, hydrogen sulfide on TSI, ONPG, urease and carbohydrate utilization tests were performed according to Edwards and Ewing (1972) and MacFaddin (1980).

^{*} PhD scholar; corresponding author



Abbreviations: ONPG, o-nitrophenyl-β-D-galactopyranoside; DNase, Deoxyribonuclease; GL, Gelatin Liquefactionl VP, Voges-Proskauer; IMViC, Indole-Methyl Red-Voges-Proskauer-Citrate and LDC, Lysine Decarboxylase

Fig. 1. A dichotomous key for identification of enterobacterial genera with selected species of histamine-forming bacteria

Histamine formation by the isolates was confirmed by using the medium of Yamani and Untermann (1985). The histamine-positive isolates were identified up to species level using the dichotomous key of Prescott

Table 1. Histamine-forming enterobacteria isolated from fresh tuna.

Organism	No. of isolates	HDC ^a positive isolates
Citrobacter spp.	2	1
Enterobacter aerogenes	2	2
Enterobacter spp.	2	1
Escherichia coli	3	3
Klebsiella pneumoniae	1	1
Klebsiella oxytoca	2	2
Morganella morganii	1	1
Proteus mirabilis	3	3
Proteus vulgaris	1	1
Proteus spp.	1	_
Serratia spp.	3	2
Yersinia spp.	2	_
Untypable	5	
Total	28	17

^a Histidine decarboxylase

et al. (1996) after modifications (Figure 1) with the help of conventional biochemical tests given by Kreig and Holt (1984).

The APC and enterobacterial count of the fresh tuna were observed to be 1.5 x 10⁵ cfu/g and 2.17 x 10² cfu/g respectively. The incidence of enterobacterial HFB observed in fresh tuna is detailed in Table 1. It is seen that the majority of the enterobacterial genera encountered have the ability to decarboxylate histidine to histamine. However, they have been reported to vary in the rate of histamine production (Arnold and Brown, 1978). *H. alvei*, another potent histamine-forming organism, could not be detected in the isolates.

The present study revealed that the proportion of enterobacterial HFB has accounted for less than 0.1 % of the APC that was closely related to the low enterobacterial count in fresh tuna.

The authors are thankful to the Director, Central Institute of Fisheries Technology, Cochin for the permission to publish this paper.

References

- Alves, R.T., Santos, A.T. and Martins, M.F. (2002) Detection of histamine-producing bacteria using polymerase chain reaction techniques and DNA probes. *Eur. Food Res. Technol.*, **214**, pp. 178-180.
- Arnold, S.M. and Brown, W.D. (1978) Histamine (?) toxicity from fish products. *Adv. Food Res.*, **24**, pp. 114-154.
- Baranowski, J. (1985) Assay for histidine decarboxylase activity. In: Histamine in marine products production by bacteria, measurement and prediction of formation. *FAO Fish Tech Pap.*, **252**, pp. 10-13.
- Chen, C.M., Wei, C.I., Koburger, J.A. and Marshall, M.R. (1989) Comparison of four agar media for detection of histamine-producing bacteria in tuna. *J. Food Prot.*, **52**, pp. 808-813.
- Edwards, P.R. and Ewing, W.H. (1972) In: *Identification of enterobacteriaceae*. 2nd ed. Buregess Publishing Co. Minneapolis.
- Frank, H.A., Baranowski, J.D., Chongsiriwatana, M., Brust, P.A. and Premaratne, R.J. (1985) Identification and decarboxylase activities of bacteria isolated from decomposed mahimahi (*Coryphaena hippurus*) after incubation at 0 and 32° C. *Int. J. Food Microbiol.*, **2**, pp. 331-341.
- Gopakumar, K., Surendran, P.K. and Vijayan, P.K. (1988) *Incidence of histidine decarboxylating bacteria and histamine levels in fish sold in retail markets*. Papers presented at the seventh session of the Indo-Pacific fishery commission working party on fish technology and marketing, 19-22 April, 1988. FAO Fish. Rep. No. **401** Suppl., pp. 126-132.
- ICMSF. (1978) Microorganisms in foods 1. Their significance and methods of enumeration. 2nd ed. pp 140-143. ICMSF of Interna-

- tional Association of Microbiological Societies, University of Toronto Press, Toronto.
- Kim, S.H., An, H., Field, K.G., Wei, C.I., Barros-Velazquez, J., Ben-Gigirey, B., Morrisey, M.T., Price, R.J. and Pitta, T.P. (2003). Detection of *Morganella morganii*, a prolific histamine former, by the polymerase chain reaction assay with 16s rDNA-targetted primers. *J. Food Prot.*, 66(8), pp. 1385-1392.
- Klausen, N.K. and Huss, H.H. (1987) A rapid method for detection of histamine producing bacteria. *Int. J. Food Microbiol.*, 5, pp. 137-146.
- Kreig, N.R. and Holt, J.G. (ed.) (1984) *Bergey's manual of systematic bacteriology*. Vol. 1, pp. 408-516. The Williams and Wilkins, Baltimore.
- Lakshmanan, R., Jeyashakila, R. and Jeyasekaran, G. (2002) Survival of amine-forming bacteria during the ice storage of fish and shrimp. *Food Microbiol.*, 19, pp. 617-625.
- Lopez-Sabater, E.I., Rodriguez-Jerez, J.J., Hernandez-Herrero, M. and Mora-Ventura, M.T. (1996) Incidence of histamine-forming bacteria and histamine content in scombroid fish species from retail markets in the Barcelona area. *Int. J. Food Microbiol.*, **28**, pp. 411-418.
- MacFaddin, J.F. (1980) Biochemical tests for the identification of medical bacteria. 2nd edition. The Williams and Wilkins, Baltimore.
- Maturin, L.J. and Peeler, J.T. (1995) *Aerobic plate count*. In: FDA's bacteriological analytical manual, 8th ed. AOAC International, Gaithersburg.
- Mavromatis, P. and Quantick, P.C. (2002) Modification of Niven's medium for the enumeration of histamine-forming bacteria and discussion of the parameters associated with its use. *J. Food Prot.*, **65**, pp. 546-551.
- Niven, C.F., Jeffrey, M.B. and Corlett Jr., D.A. (1981). Differential plating medium for

- quantitative detection of histamine-producing bacteria. *Appl. Environ. Microbiol.*, **41(1)**, pp. 321-322.
- Okuzumi, M., Hiraishi, A., Kobayashi, T. and Fuji, T. (1994) *Photobacterium histaminum* sp. nov., a histamine producing marine
- bacterium. *Int. J. Syst. Bacteriol.*, **44**, pp. 631-636.

 Prescott, L.M., Harley, J.P. and Klein, D.A.
- (1996) The bacteria: Gram-negative bacteria of general, medical or industrial importance. In: *Microbiology*, 3rd ed. pp. 427. Wm. C. Brown Publishers, London.
- Roig-Sagues, A.X., Lopez-Sabater, E.I., Rodriguez-Jerez, J.J., Hernandez-Herrero, M. and Mora-Ventura, M.T. (1997) Evaluation of three decarboxylating agar media to detect histamine and tyramine-producing bacteria in ripened sausages. *Lett Appl. Microbiol.*, **25**, pp. 309-312.

- Subburaj, M., Karunasagar, I and Karunasagar, I. (1984) Incidence of histidine decarboxylating bacteria in fish and market environs. *Food Microbiol.*, 1, pp. 263-267.
- Taylor, S.L. and Sumner, S.S. (1986) Determination of histamine, putrescine and cadaverine. In: *Seafood quality determination* (Kramer, D.E. and Liston, J. edn), pp. 235-245. Elsevier Science Publ., Amsterdam.
- Yamani, M.I. and Untermann, F. (1985) Development of a histidine decarboxylase medium and its application to detect other amino acid decarboxylases. *Int. J. Food Microbiol.*, **2**, pp. 273-278.
- Yoshinaga, D.H. and Frank, H.A. (1982) Histamine-producing bacteria in decomposing skipjack tuna (*Katsuwonus pelamis*). *Appl. Environ. Microbiol.*, **44**, pp. 447-452.