## Effect of Mercuric Nitrate on the Chromosomes of Channa punctatus (Bloch, 1793)

Ansy Mathew, N.P.\* and Shrinivas Jahageerdar

Central Institute of Fisheries Education Mumbai - 400 061, India

The effect of different doses (0.019, 0.038 and 0.075 ppm) of mercuric nitrate on the chromosomes of *Channa punctatus* was studied. The karyotype was done by standard colchicine, potassium chloride, acetomethanol and air-drying geimsa staining technique. It was observed that the percentage of Total Metaphase spreads with Chromosomal Aberrations (T.M.C.A.) increased with the increasing concentration levels. Mercuric nitrate induced structural chromosomal aberrations included breaks, fragments, dicentric and ring type chromosomes. The study revealed that 120 h of exposure in mercuric nitrate at concentration levels as low as 0.019 ppm induced chromosomal aberrations in fish.

Key words: Karyotype, mercuric nitrate, Channa punctatus

Studies on chromosomes are essential in fish genetics, taxonomy and environmental toxicology research programmes. The genetic variations can be detected at chromosomal and genomic level and in practice conventional karyological techniques are used as reliable tools. In the absence of direct genotoxic test the karyotyping of fish can be used as a simple test and also as starting point for detailed genotoxic studies. The sublethal mutations do not cause immediate mortality to the fish. However, they have more severe consequences for the development of the entire population. Heavy metals such as lead, mercury, zinc, copper and cadmium are known to cause damage to the genetic material in aquatic animals (Abel, 1989). The LC<sub>50</sub> for mercuric nitrate was considered as 0.15 mg/l (Sebastian and Gerlach, 1981). The indirect exposure of fishes to Phenyl Mercuric Acetate (P.M.A.) increased the frequency of chromosomal aberrations at all dose levels from 0.01 to 0.05 ppm (Krishnaja and Rege, 1982).

Manna (1986, 1989) reviewed genotoxic studies in fish and opined, tilapia to be a good cytogenetic model for testing genotoxicity. Subsequently Thomas (1990), Mitrofanov et al. (1991), Pechkurenkov (1991), Liu et al. (1991), Nishimoto et al. (1991), Zhang et al. (1992), Bailey et al. (1992), Yasuhira et al. (1992), Reichert et al. (1992), Schnitz and Connor (1992), Kubota et al. (1992), Adams et al. (1992) and Aoki et al. (1993) have studied various aspects of fish genotoxicity using different species including, various fresh water carps, rainbow trout, gold fish, tilapia, eastern mud minnow, etc.

The majority of fish has large number of 2n chromosomes and is acrocentric type, which makes it difficult to study the genotoxic effects. However, *Channa punctatus* has small number of 2n chromosomes and majority of them are larger in size, which makes it a suitable species on genotoxic effects (Manna, 1983). The present study was

<sup>\*</sup> Present address: Senior Research Fellow, C.M.F.R.I., P.O. Box- 1603, Tatapuram P.O., Cochin - 682 014, India E-mail: ansy@rediffmail.com

envisaged to study the effect of different concentrations of mercuric nitrate on normal chromosomes of *Channa punctatus* and to compare the chromosomes of fish exposed to mercuric nitrate with those of normal *Channa punctatus*.

## Material and methods

Channa punctatus weighing 30-50 g were reared in the laboratory and maintained under standard management conditions. The genotoxic effect of mercuric nitrate was studied at 0.019, 0.038 and 0.075 mg/l concentration (Table 1). To obtain high rated mitotic metaphase spreads and preparation of permanent slides, colchicine 0.1% was used at the rate of 0.5ml/mg body weight of fish. After 3 to 4 hours, gills, kidney and liver were collected, immediately ground well and kept in KCl hypotonic solution for 30 min at room temperature. Each tissue sample along with hypotonic solution was centrifuged for 10 min at 1000 rpm. The supernatant was discarded.

A freshly prepared and chilled fixative (methanol: acetic acid (3:1) of about 5.0 ml was added to each tube slowly along the sides and the contents were resuspended and subjected to centrifugation for 10 min at 1000 rpm and the supernatant was discarded. The above step was repeated two or three times until a clear supernatant along with a mass of white cells was obtained as sediment. The final volume of the fixative was adjusted based on a pellet size and a uniform suspension was prepared. Four to five drops of the cell suspension was allowed to fall on a clean cold slide (4°C) from the height of about two to three feet. Then the slides were air dried for one hour. 4% Geimsa stain in Sorenson's phosphate buffer was used for staining the chromosomes. The slides were stained for 30 min in a cuplin jar and washed in distilled water. The slides were air dried and finally mounted with DPX. Each slide was systematically screened to identify different chromosomal abnormalities viz. chromosomal breaks, fragments, dicentric and ring types. Careful attention was taken to discriminate morphological aberrations, which might have occurred due to defective colchicine or hypotonic treatment on staining and washing procedure.

The following criteria were adapted to choose suitable metaphase spreads:

- i) Minimum chromosomal overlapping
- ii) Chromosomes of sufficient length
- iii) A distinct sharp banding pattern.

The metaphase spreads were photographed under oil immersion objective with a final magnification of 1000x. Individual chromosomes were cut from photo prints. The homologous pairs were made and arranged in the order of metacentric, submetacentric, subtelocentric and acrocentric. Chi-square test of significance was applied to detect the significant genotoxic effect of heavy metal at different doses, sex and time intervals (Pillai and Rajagopalan, 1979).

## Results and discussion

The model number of chromosomes in the present study is 2n = 32 consisting of 14 metacentric, 8 submetacentric, 6 subtelocentric and 4 acrocentric chromosomes (Plates 1 and 2). The absence of difference between male and female karyotype indicates that the heteromorphic chromosomes are absent in this species. The study revealed that mercuric nitrate at the

Table 1. Analysis of chromosomal abnormalities induced in *Channa punctatus* after exposure to various doses of mercuric nitrate for 120 h

Treatment group (mg/l)	Total number of spreads studied	Metaphase with chromosomal abnormalities	three or more	ber of spreads two aberrations	with at least one aberration	χ² value	Percentage of spreads with chromosomal abnormalities
0.019	134	86	20	19	52	39.8	67.9
control	134	49	0	0	08		34.6
0.038	127	91	32	22	32	58.9	77.7
control	110	39	0	0	13		35.5
0.075	116	99	37	23	39	102.5	85.4
control	123	31	0	0	12		35.2

Value ( $P \le 0.01$ )

concentration of 0.019 mg/l induces highly significant chromosomal abnormalities in *Channa punctatus* (Table 1). The structural abnormalities in the chromosome induced by mercuric nitrate consist of chromosomal breaks, fragments, dicentric chromosomes and ring type of chromosomes (Plate 3). With the increase in concentrations the percentage of Total Metaphase spreads with Chromosomal Abnormalities (T.M.C.A.) increased (Table 1).

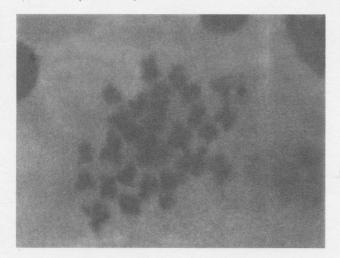


Plate 1. Normal metaphase spread of Channa punctatus

Mercuric nitrate at the concentration of 0.019 mg/l was clastogenic. The percentage of metaphase spreads with chromosomal abnormalities increased gradually as the concentration increased. It was 67.9 percent

when the exposure time was 120 h (Table 1). In all the concentrations, metaphase spreads, which contained three or more type of chromosomal abnormalities, were observed (Table 1). The percentage of metaphase spreads when the fish were exposed to mercuric nitrate (0.038mg/l, 0.075mg/l) for 120 h was 77.7 and 85.4 (Table 1). At all the concentrations, chromosomal breaks were more in number followed by fragments (Plates 3,4,5). The dicentric and ring chromosomes were in equal percent. Both male and female fish were affected equally at all three exposures. The difference between the percentages of metaphase spreads with chromosomal abnormalities between the two sexes

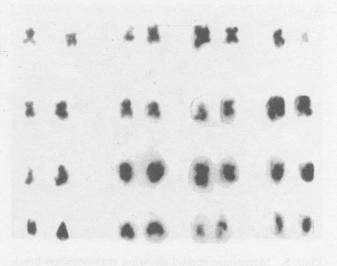


Plate 2. Normal karyotype of Channa punctatus

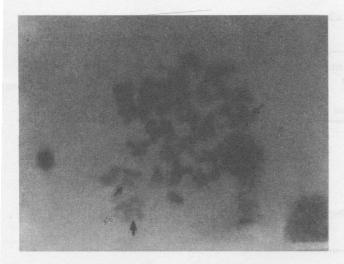


Plate 3. Metaphase spread showing chromosomal break and fragment at 0.019 mg/l for 120 h

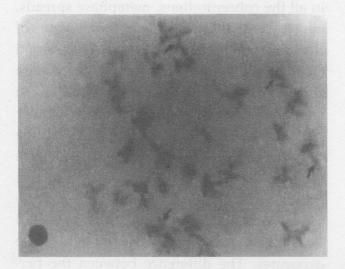


Plate 4. Metaphase spread showing chromosomal break and fragment at 0.038 mg/l for 120 h

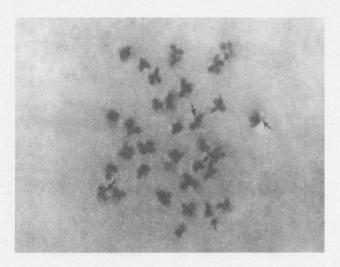


Plate 5. Metaphase spread showing chromosomal break and fragment at 0.075 mg/l for 120 h

(male and female) was not significant ( $P \le 0.01$ ). The results also indicated that both the sexes were equally susceptible to mercuric nitrate poisoning. The highly significant percentage of metaphase spreads with chromosomal abnormalities observed in fish exposed to mercuric nitrate as compared to control groups indicates that mercury acts as a potential clastogen even at a very low concentration (0.019mg/l). Therefore, it is important that fish should be reared in mercuric nitrate free water. Extreme care should be taken while selecting the brood stock. The brood stock should be selected from a pollution free environment.

The authors are indebted to Central Institute of Fisheries Education (ICAR), for granting fellowship to the first author and for providing facilities to execute the work. They are thankful to Dr. J.P. George, Dr. P. Kaladharan and Mr. K.K. Anikkuttan, of C.M.F.R.I., and to Dr. M. M. Prasad, C.I.F.T. for their valuable help.

## References

Abel, P.D. (1989) Water Pollutions Biology. Ellis Hord Wood Ltd., John Waley & Sons, NewYork, pp 168-192.

Adams, S.M., Crumbey, W.D., Greeley Jr. M.S., Shugart, L.R. and Saylor, C.F. (1992) Responses of fish populations and communities to pulp mill effluents: a holistic assessment. *Ecotoxicol. Environ. Safety*, **24**, pp 347-360.

Alex Frazer (1966) *Heredity, Genes and Chromosomes*. Mc Grain Hill Book Company, New York, **12**, pp 163-164.

Aoki, K., Nakatsuru, Y., Sakurai, J., Sato, A., Masahito, P. and Ishikawa, T. (1993) Age dependence of O<sub>6</sub>- methyl guanine DNA

methyl transferase activity and its depletion after carcinogen treatment in the teleost medaka (*Oryzias latipes*). *Mutat. Res.*, **293** pp 225-231.

- Bailey, G., Hendricks J. and Dashwood, R. (1992) Anticarcinogenesis in fish. *Mutat. Res.*, **267**, pp 243-250.
- Krishnaja, A.P. and Rege, M.S. (1982) Induction of chromosomal aberrations in fish *Boleophthalmus dussumieri* after exposed in vitro in Mitomycin C and heavy metals Mercury, Selenium and Chromium. *Mutat. Res.* **102** pp 71-82.
- Kubota, Y., Shimada, A. and Shima, A. (1992)
  Detection of gamma ray induced DNA damages in malformed dominant lethal embryos of the Japanese medaka (*Oryzias latipes*) using AP-PCR finger printing. *Mutat. Res.*, 283, pp 263-270.
- Liu, T.Y., Cheng, S.L., Ueng, T.H., Veng, Y.F., and Chi, C.W. (1991) Comparative analysis of aromatic DNA adducts in fish from polluted and unpolluted areas by the 32p-post labeling analysis. *Bull. Environ. Contam. Toxicol*, **47**, pp 783-789.
- Manna, G.K. (1986) Tilapia as a model for testing genotoxic agents. *In: Perspectives in cytology and Genetics*, pp 395-406.
- Manna, G.K. (1989) Fish cytogenetics related to taxonomy, evolution and monitoring aquatic genotoxic agents. *In: Fish Genetics in India*, Das, P., and A.G. Ghingran (Eds), Today and Tomorrows printers and publishers, New Delhi, pp 21-46.
- Mitrofanov IUA., Lesnikova, L.N., Voskanian, A.Z. and Otradnona, V.V. (1991)

- Alternative and non-alternative process of the origin of chromosome aberrations. *Radiobiologia*, **31**, pp 585-592.
- Nishimoto, M., Roubal, W.T., Stein, J.E. and Varanasi, U. (1991) Oxidative DNA damage in tissues of English sole (*Parophrys vetulus* exposed to nitrofuration. *Chem. Biol. Interact*, **80**, pp 317-326.
- Pechkurenkov, V.L. (1991) The effect of the accident at Chernobyl Atomic Electronic Power Station in 1986 on the fish population of a cooling pond. *Radiobiologia*, 31(5), pp 704-708.
- Perry, D.M., Weis, J.S. and Weis, P. (1988) Cytogenetic effects of methyl mercury in embryos of the killifish, *Fundulus heteroclitus*. *Arch*. *Environ*. *Contam*. *Toxicol*. 17, pp 569-574.
- Pillai, M.S. and Rajagopalan (1979) Statistics for Economics students. 20,pp 24-28, T 14. Progressive Corporation Pvt. Ltd., Bombay.
- Reichert, W.L., Stein, J.E., French, B., Goodwin, P. and Varanasi, U. (1992) Storage phosphor imaging technique for detection and quantification of DNA adducts measured by the 32p. Post labeling assay. *Carcinogenesis*, 13, pp 1475-1479.
- Schnitz, A.R. and O. Connor, J.M. (1992) In vivo DNA/RNA adduction of 7,12-dimethylbenz (a) anthracene (DMBA) and benzo (a) pyrene (Bap) in the liver of rainbow trout (Onchorhynchus mykiss). J. Environ. Pathol. Toxicol. Oncol., pp 229-233.

Sebastian, A. and Gerlach (1981) Marine Pollution Diagnosis and Therapy, Springer-Verlag, New York. p.125.

Thomas, P. (1990) Molecular and biochemical responses of fish to stressors and their potential use in environmental monitoring. *In: Biological indicators of stress in fish* 

Symposium 8, Bethesda, Maryland pp 9-28. Veljovic P., Dukic, D. and Soldatovic, B. (1990)
The karyotype characteristics of some fish species in the ecosystem of river Zapadna Morava. *Acta Biol. IUGOSL. E. Ichthyol.*, **22**, pp 53-56.

Adams, S.M., (Ed.) American Fisheries

Yasuhira, S.H., Mitani, A. and Shima, A. (1992) Enhancement of photo repair of ultraviolet-induced pyrimidine dimmers by pre illumination with fluorescent light in the gold fish cell line. The relationship between survival and yield of pyridine dimmers. *Photochem. Photobiol.*, **55**, pp 97-101.

Zhang, Q., Suorsa-Super, K. and Curtis, L.R. (1992) Temperature modulated aflatoxin B1 hepatic disposition and formation and persistence of DNA adducts in rainbow trout. *Toxicol. Appl. Pharmacol.*, 113, pp 253-259.