# Response of three popular varieties of wheat to arbuscular mycorrhizae grown in two common soil types of central India

Neha Chakravarty, Ashok Shukla, Anil Kumar<sup>\*</sup> and Sudhir Kumar

ICAR-Central Agroforestry Research Institute, Gwalior Road, Jhansi – 284 003, Uttar Pradesh. \*Corresponding author's E-mail: anilgargnrcaf@gmail.com

**ABSTRACT**: An experiment was conducted to study the response of three popular varieties of wheat, namely WH 147, Lok-1 and Kathiya, to ten arbuscular mycorrhizal (AM) fungi in two common soil types [alfisol (red) and vertisol (black)] of central India, under net-house conditions. The varieties recorded variable response in terms of growth and yield. Maximum yield was recorded in LOK-1, followed by WH 147 and Kathiya. AM inoculations significantly increased plant height, dry shoot weight, dry root weight and yield. The growth of different varieties was more vigorous in alfisol than vertisol. Maximum root:shoot ratio was recorded in Kathiya, which was significantly more than other two varieties. All the inoculants, except *Acaulospora mellea* and *A. scrobiculata* increased the root:shoot ratio. LOK-1 recorded maximum mycorrhizal dependency (MD), followed by Kathiya and WH 147. Plants grown in alfisol exhibited higher MD value than vertisol. Among different varieties, maximum phosphorus (P) uptake was recorded in Kathiya, followed by LOK-1 and WH 147. All AM inoculants significantly increased P uptake. Its higher value was recorded in plants grown in alfisol. Maximum root colonization index was recorded in WH 147, which varied from 32.4 to 47.3% in different AM treatments. Plants grown in alfisol recorded significantly higher colonization index. Thus, the results showed that AM fungi increased the growth, yield and P uptake of three popular varieties of wheat, which signifies that these can be utilized for inoculation of the crop under central Indian conditions.

Keywords: Arbuscular mycorrhizal fungi, central India, soil types and wheat cultivars.

Received on: 21.05.2018

#### 1. INTRODUCTION

Wheat is an important intercrop of agroforestry. It has given good results under agroforestry systems (Shukla et al., 2012a). Historically, its four species, namely Triticum aestivum, T. durum, T. dicoccum and T. sphaerococcum were under cultivation. T. sphaerococcum has now practically gone out of cultivation because of its low productivity. In total production, T. aestivum contributes approximately 95%, followed by T. durum with 4% and T. dicoccum with 1%. India is surplus in wheat production but it is predicted that an annual rate increase of 4 to 5% in wheat production would be required in relation to the rate and nature of economic growth, population expansion and income elasticity (Goyal and Singh, 2002). In order to increase the production, the technologies utilizing indigenous microbes need to be explored (Minaxi et al., 2013).

Arbuscular mycorrhiza (AM) fungi are known to occur widely under various environmental conditions and are found associated with roots of most of the crops (Jha *et al.*, 2012). By acquiring phosphate, micronutrients and water, they enhance the host nutritional status and thus their growth. In addition, AMF stimulate the production of growth substances and reduce stresses, diseases and pest attack (Shukla *et al.*, 2014; Dehariya *et al.*, 2015). For the appropriate use of these fungi, it is necessary to select the best fungi adapted to the specific environmental factors for crop productivity (Herrera-Peraza *et al.*, 2011). Soil properties can influence the efficiency of AM inoculants (Carrenho et al., 2007; Gryndler et al., 2009; Shukla et al., 2013). According to Ramadhani et al. (2015), the effectiveness of AM inoculations can be different among species or even among varieties (cultivars) in a species. Varied response of different wheat varieties to AM inoculations have been reported by several workers (Hetrick et al., 1992; Behl et al., 2003; Singh et al., 2012). Therefore, an attempt was made to study the suitability of AM fungi for inoculation of the crop in central Indian soil types. In present study, three popular varieties of wheat among local farmers, namely WH 147 (bread wheat), Lok-1 (bread wheat) and Kathiya (macaroni wheat), were screened against ten AM fungi in two common soil types.

#### 2. MATERIALS AND METHODS

The study was conducted at ICAR-Central Agroforestry Research Institute, Jhansi (25°27' N latitude, 78°35' E longitude and 271 m above mean sea level), which consisted of 11 treatments [10 AM species (*Acaulospora mellea, A. scrobiculata, Claroideoglomus etunicatum, Glomus aggregatum, G. arborense, G. cerebriforme, Rhizophagus diaphanus, R. fasciculatus, R. intraradices, Simiglomus hoi*) and a control], three wheat varieties [WH 147 (hexaploid), Lok-1 (hexaploid) and Kathiya (tetraploid)] and two soil types [red (alfisol) and black (vertisol) soils]. All treatments were replicated three times in completely randomized design (CRD). Thus, a total of 198 pots were employed in the study.

Accepted on : 12.06.2018

The soil substrates were passed through 2 mm sieves separately, moistened with water and filled in cotton bags, and autoclaved at 15 psi (121 °C for 30 minutes). After filling pots with autoclaved substrates, the mycorrhizal treatments were imposed and seeds of three wheat varieties were sown, as per treatments. The treated pots were transferred to the net-house and thinning was carried out 15 days after sowing, leaving one plant per pot. The pots were watered as and when required. The plants were harvested at maturity and observations on plant height, number of tillers plant<sup>-1</sup>, dry shoot weight, dry root weight and vield plant<sup>-1</sup> were recorded. Phosphorus (P) uptake plant<sup>1</sup> was estimated by vanado-molybdo phosphoric yellow color method (Jackson, 1973). Mycorrhizal dependency (MD) was calculated according to Plenchette et al. (1983). At the time of harvest, 1g fine roots from each plant were collected to estimate AM colonization index. Collected samples were cleared with 10% KOH and stained with acid fuchsin (0.01% in lactoglycerol) as reported by Kormanik et al. (1980) and the index was determined by gridline intersect method (Giovanneti and Mosse, 1980). The root: shoot ratio, which refers to the proportion of dry root weight to dry shoot weight, was also calculated.

All the data were subjected to three-way analysis of variance (ANOVA) for testing the effect of plant varieties, AM inoculations, soil types and interactions. Least significant difference (P<0.05) values were used to compare the treatment differences.

#### 3. RESULTS AND DISCUSSION

Different wheat varieties recorded variable response in terms of growth and yield. Maximum plant height was recorded in WH 147, dry shoot weight in LOK-1 and dry root weight in Kathiya. Maximum yield was recorded in LOK-1, which was at par with WH 147. These values were significantly higher than the value recorded in Kathiya. AM inoculations significantly increased plant height, dry shoot weight, dry root weight and yield. Values of studied growth parameters (plant height, dry shoot weight and dry root weight) were significantly higher in plants grown in alfisol than in vertisol. However, soil types did not affect the yield (Table 1).

Maximum root: shoot ratio was recorded in Kathiya, which was significantly more than other two varieties. All the inoculants, except *A. mellea* and *A. scrobiculata* significantly increased the root: shoot ratio. The differences in its values in two soil types were found non-significant. Variety LOK-1 showed maximum dependency on inoculated fungi for dry matter production, followed by Kathiya and WH 147.

MD of different wheat varieties on AM inoculants varied in a narrow range *i.e.* from 44.63-48.64%. MD in alfisol was significantly higher than MD in vertisol (Table 2). Among different varieties, maximum P uptake was recorded in Kathiya, which was at par with LOK-1. Its minimum value was recorded in WH 147. All AM inoculants significantly increased P uptake (Table 3). Its significantly higher value was recorded in plants grown in alfisol than vertisol. Maximum root colonization index was recorded in WH 147, which was at par with Lok-1 and significantly more than Kathiya. It varied from 32.37 to 47.31 in different AM treatments. Colonization index in alfisol was significantly higher than colonization index in vertisol (Table 4).

The results showed that studied wheat varieties exhibited variable growth, yield and P uptake patterns. This should be due to genetic variability. Variable response of different wheat cultivars to AM inoculations have been reported by several workers (Azcon and Ocampo, 1981; Vierheilig and Ocampo, 1991). Genetic variations for AM symbiosis have also been reported in many other crops also, like Lycopersicon esculentum (Bryla and Koide, 1998), Pisum sativum (Rivera-Becerril et al., 2002), Phaseolus vulgaris (Hacisalihoglu et al., 2005), Trifolium repens (Eason et al., 2001), Zea mays (Ortas and Akpinar, 2011), etc.

As per our results, different AM inoculants increased plant height by 11-19%, shoot dry weight by 74-90%, root dry weight by 94-129% and yield by 88-106%, over control. Increase in growth and yield of AM inoculated plants can be attributed to increase in the soil volume explored for nutrient and water uptake by the mycorrhizal plants as compared to nonmycorrhizal ones. Better nutrient, especially P, which is evident from the results obtained in our study, generally leads to increase in plant biomass (Klironomos, 2003; Shukla et al., 2012b). Beneficial effects of AM inoculations on growth and productivity of wheat have been reported by various workers (Mohammad et al., 1995; Karagiannidis and Hadjisavva-Zinoviadi, 1998; Shukla et al., 2009; Abdel-Fattah and Asrar, 2012).

Further, the results obtained in present study suggested that plants grown in alfisol showed more dependency on inoculated fungi than in vertisol, which could be due to less fertility. According to Carrenho *et al.* (2007), low fertile soil limits plant development and increases their dependency on mycorrhizal association. Ortas and Akpinar (2006) have also suggested that the response of plants to AM inoculation is generally linked with soil fertility levels

Table 1. Effect of AM inoculations on growth and yield of three varieties of wheat in two soil types

	ns Pooled
WH 147 Lok-1 Kathiya Mean WH 147 Lok-1 Kathiya Mean WH 147 Lok-1	Kathiya mean
Plant height (cm)	
A. mellea 83.3 90.0 80.0 84.4 80.7 78.0 66.0 74.9 82.0 84.0	73.0 79.7
A. scrobiculata 81.7 85.3 68.0 78.3 82.0 68.0 82.0 77.3 81.3 76.7	75.0 77.8
C. etunicatum 81.3 88.7 61.3 76.4 83.3 78.0 78.0 79.8 82.3 82.3	69.7 78.1
G. aggregatum 84.0 85.0 72.0 80.3 86.0 72.7 72.7 77.1 85.0 78.3	72.3 78.7
G. arborense 87.7 81.3 78.0 82.3 80.7 75.3 70.7 75.6 84.2 78.3	74.3 78.9
G. cerebriforme 86.0 88.0 72.0 82.0 81.3 82.0 69.3 77.6 83.7 85.0	70.7 79.8
R. diaphanus 96.3 88.0 72.0 85.4 84.0 80.7 75.3 80.1 90.2 84.3	73.7 82.7
R. fasciculatus 82.7 86.7 74.0 80.4 78.0 80.0 69.3 75.8 80.3 82.3	71.7 78.1
R. intraradices 88.7 84.0 77.3 83.3 77.3 74.0 70.0 73.8 83.0 79.0	73.7 78.6
Sim. hoi 92.0 89.7 57.3 79.7 84.0 76.7 63.3 74.7 88.0 83.2	60.3 77.2
Un-inoculated 81.7 76.0 57.3 71.7 71.3 73.2 57.3 67.3 76.5 74.6	57.3 69.5
Mean 85.9 85.3 69.9 80.4 80.8 76.2 70.4 75.8	
Pooled mean 83.4 80.8 70.2	
Dry shoot weight (g)	
A. mellea 35.4 39.2 30.8 35.1 34.1 34.1 30.7 33.2 35.1 36.7	30.8 34.2
A. scrobiculata 34.2 35.5 34.0 34.6 36.6 36.7 33.0 35.4 35.4 36.1	33.5 35.0
C. etunicatum 36.0 33.2 29.2 32.8 31.6 32.0 30.8 31.5 33.8 32.6	30.0 32.1
G. agaregatum 34.0 37.2 29.8 33.7 34.2 37.3 31.4 34.3 34.1 37.2	30.6 34.0
G. arborense 34.3 35.6 33.3 34.4 32.6 32.0 29.2 31.3 33.5 33.8	31.3 32.8
G. cerebriforme 35.0 33.4 30.7 33.0 31.0 32.8 30.6 31.5 33.0 33.1	30.7 32.3
R diaphanus 34.7 35.2 34.5 34.8 32.1 37.4 30.6 33.4 33.4 36.3	32.5 34.1
R. fasciculatus 36.3 36.7 33.0 35.3 35.1 33.4 31.5 33.3 35.7 35.1	32.3 34.4
R. intraradices 34.5 40.1 32.3 35.6 32.6 31.7 30.6 31.6 33.5 35.9	31.4 33.6
Sim hol 34 4 37 5 29 9 33 9 34 9 33 7 31 9 33 5 34 7 35 6	30.9 33.7
Un-inoculated 18.8 18.3 19.2 18.8 18.4 17.1 18.4 18.0 18.6 17.7	18.8 18.4
Mean 33.4 34.7 30.6 32.9 32.2 32.6 29.9 31.5	
Pooled mean 32.8 33.6 30.2	
Dry root weight (g)	
A mellea 10.3 10.1 23.8 14.7 7.5 9.4 21.7 12.9 8.9 9.8	22.7 13.8
A. scrobiculata 8.9 8.7 24.2 13.9 7.9 8.8 22.9 13.2 8.4 8.7	23.5 13.6
C etunicatum 97 83 255 145 95 84 275 151 96 84	26.5 14.8
G. aggregatum 8.11 9.5 22.4 13.3 9.2 12.3 22.8 14.8 8.6 10.9	22.6 14.1
<i>G</i> arborense 10.0 13.1 29.0 17.4 9.8 10.6 24.2 14.9 9.9 11.8	26.6 16.1
G cereptiforme 99 93 252 148 89 98 187 125 94 96	21.9 13.6
R. diaphanus 8.8 10.6 23.0 14.1 9.7 10.2 24.2 14.7 9.3 10.4	23.6 14.4
<i>R</i> fasciculatus 8.6 12.8 28.2 16.5 9.4 9.0 23.0 13.8 9.0 10.9	25.6 15.2
R intradices 9.5 10.2 23.9 14.6 9.3 8.1 27.3 14.9 9.4 9.2	25.6 14.7
Sim hol 98 99 240 146 97 105 193 132 97 102	21.7 13.9
Un-inoculated 4.6 5.1 11.0 6.9 6.1 4.5 10.7 7.1 5.3 4.8	10.8 7.0
Mean 8.9 9.8 23.6 14.1 8.8 9.2 22.0 13.4	
Pooled mean 8.9 9.5 22.8	
Yield (a) plant <sup>1</sup>	
<i>A mellea</i> 17 35 19 83 12 90 16 69 16 60 16 49 12 69 15 26 16 98 18 16	12.79 15.98
A scrabiculata 16.88 17.35 14.81 16.34 16.58 17.45 14.74 16.26 16.73 17.40	14.28 16.30
C etunicatum 15 10 16 04 14 75 15 30 14 56 17 64 13 97 15 39 14 83 16 84	13.86 15.34
<i>G</i> aggregation 17.76 17.97 11.32 15.35 14.69 18.38 14.87 15.98 16.23 19.51	13 10 15 67
G adorense 1658 17 10 13 55 15 74 17 23 20 33 13 97 17 18 15 90 17 88	13 76 16 46
G creptifiorme 17.35 17.85 16.29 17.16 14.10 15.30 11.15 13.50 15.70 16.57	13 72 15 33
R diaphanus 1814 1818 1325 1652 1742 1610 1431 1593 1678 1712	13.72 16.33
<i>E fascillatus</i> 17.10 15.26 13.33 15.23 16.96 15.18 12.57 14.90 17.03 15.22	12.95 15.01
R intradices 17 31 17 52 12 64 15 82 20 80 15 35 12 75 16 13 18 13 16 43	12.69 15.07
Sim hoi 16.45 16.68 13.71 15.61 15.80 17.38 12.44 15.21 16.13 16.13	13 10 15 /1
In-incluided 817 749 727 765 874 860 753 832 846 910	740 702
Maan 16.11 16.48 13.07 15.22 15.72 16.20 1.28 14.01	1.70 1.70
Pooled mean 15.91 16.34 12.95	

	Plant height	Dry shoot weight	Dry root weight	Yield					
Variety	1.3	0.8	0.6	0.73	_				
AM inoculation	2.4	1.5	1.1	1.40					
Soil types	1.0	0.6	0.5	NS					
Variety × AM inoculation	4.2	2.6	1.9	NS					
Variety × soil	1.8	NS	0.8	NS					
AM inoculation × soil types	3.4	NS	1.6	NS					
Variety × AM inoculation × soil types	5.9	NS	2.7	NS					

wheat in two soil types												
AM species	Alfisol			Vertisol				V	Pooled			
	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	mean
Root: shoot ratio												
A. mellea	0.29	0.26	0.77	0.44	0.21	0.28	0.71	0.40	0.25	0.27	0.74	0.42
A. scrobiculata	0.26	0.25	0.72	0.41	0.22	0.24	0.70	0.38	0.24	0.24	0.71	0.40
C. etunicatum	0.25	0.30	0.67	0.41	0.30	0.28	0.79	0.46	0.28	0.29	0.73	0.43
G. aggregatum	0.27	0.25	0.88	0.47	0.30	0.26	0.89	0.49	0.29	0.26	0.89	0.48
G. arborense	0.24	0.26	0.75	0.41	0.27	0.33	0.73	0.44	0.25	0.29	0.74	0.43
G. cerebriforme	0.29	0.37	0.88	0.51	0.30	0.33	0.83	0.49	0.30	0.35	0.86	0.50
R. diaphanus	0.28	0.28	0.82	0.46	0.29	0.30	0.61	0.40	0.29	0.29	0.72	0.43
R. fasciculatus	0.24	0.35	0.86	0.48	0.27	0.27	0.73	0.42	0.25	0.31	0.80	0.45
R. intraradices	0.29	0.27	0.80	0.45	0.28	0.31	0.61	0.40	0.28	0.29	0.71	0.43
Sim. hoi	0.28	0.25	0.74	0.43	0.29	0.26	0.90	0.48	0.28	0.26	0.82	0.45
Un-inoculated	0.24	0.28	0.57	0.36	0.33	0.26	0.58	0.39	0.29	0.27	0.58	0.38
Mean	0.27	0.28	0.77	0.44	0.28	0.28	0.73	0.43				
Pooled mean	0.27	0.28	0.75									
Mycorrhizal depend	ency (%)											
A. mellea	48.69	52.34	44.69	48.57	42.10	50.10	44.17	45.45	45.39	51.21	44.43	4 7.01
A. scrobiculata	45.66	46.87	48.12	46.88	44.99	52.39	47.69	48.36	45.32	49.63	47.91	47.62
C. etunicatum	45.80	48.99	47.26	47.35	41.55	54.56	46.50	47.53	43.67	51.78	46.88	47.44
G. aggregatum	48.65	43.47	44.81	45.64	40.54	46.54	49.83	45.64	44.59	45.00	47.32	45.64
G. arborense	44.36	49.79	42.10	45.40	43.63	56.20	46.15	48.66	44.00	53.00	44.10	47.03
G. cerebriforme	47.00	51.70	51.64	50.11	42.20	48.98	45.37	45.52	44.60	50.34	48.51	47.81
R. diaphanus	47.63	45.31	46.11	46.35	38.73	49.20	40.80	42.91	43.18	47.25	43.46	44.63
R. fasciculatus	47.70	52.80	50.80	50.43	45.12	49.10	46.36	46.85	46.41	50.93	48.58	48.64
R. intraradices	46.91	50.73	43.92	47.18	45.16	50.78	43.03	46.32	46.03	50.75	43.47	46.75
Sim. hoi	46.63	53.37	46.29	48.76	41.65	45.23	49.53	45.47	44.14	49.30	47.91	47.12
Mean	46.90	49.54	46.57	47.67	42.57	50.30	45.94	46.27				
Pooled mean	44.73	49.92	46.26									
						LSD						
			Ro	ot:shoot ra	atio Mycorrhizal				endency			
Variety				0.02		1 21						
AM inoculation				0.04				2.20				
Soil types				NS				0.99				
Variety × AM inoculat	ion			0.07				3.82				

 Table 2. Effect of AM inoculations on root:shoot ratio and mycorrhizal dependency (%) of three varieties of wheat in two soil types

## Table 3. Effect of AM inoculations on phosphorus uptake (mg plant<sup>-1</sup>) of three varieties of wheat in two soil types

1.71

3.12

5.40

NS

0.06

0.10

AM species		Alfi	sol		Vertisol				V	Pooled		
	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	mean
A. mellea	17.30	26.18	38.14	26.21	8.60	9.25	16.46	11.44	12.95	16.21	27.30	18.82
A. scrobiculata	12.38	17.83	45.88	25.36	10.93	10.08	21.66	14.23	11.66	13.95	33.77	19.79
C. etunicatum	16.08	47.17	23.78	29.01	12.21	13.12	14.62	13.31	14.14	30.15	19.20	21.16
G. aggregatum	25.33	34.78	28.64	29.59	12.34	11.54	14.31	12.73	18.84	23.16	21.48	21.16
G. arborense	15.37	46.84	44.49	35.57	12.80	17.43	16.36	15.53	14.10	32.14	30.42	25.55
G. cerebriforme	19.60	27.53	40.33	29.15	9.39	11.32	15.00	11.91	14.50	19.43	27.67	20.53
R. diaphanus	8.68	24.36	20.13	17.72	8.21	10.64	7.22	8.69	8.44	17.50	13.68	13.21
R. fasciculatus	16.44	47.15	54.99	39.53	10.92	13.15	17.80	13.96	13.68	30.15	36.40	26.74
R. intraradices	18.97	40.53	31.29	30.26	9.52	15.34	11.43	12.10	14.24	27.94	21.36	21.18
Sim. hoi	19.27	52.60	28.56	33.48	11.47	7.99	14.91	11.46	15.37	30.31	21.73	22.47
Un-inoculated	4.54	7.62	4.34	5.50	3.59	3.79	4.56	3.98	4.06	5.71	4.45	4.74
Mean	15.82	33.60	32.78	27.40	9.99	11.24	14.03	11.76				
Pooled mean	12.91	22.42	23.41									
					LSD <sub>0.05</sub>							
Variety					1.27							
AM inoculation					2.44							
Soil types					1.04							
Variety × AM inocula	tion				4.22							
Variety × soil					1.80							
AM inoculation × soil	types				3.45							
Variety × AM inocula	tion × soil type	S			5.97							

Variety × soil

AM inoculation × soil types

Variety × AM inoculation × soil types

AM species	Alfisol			71		Vertisol				Varietal means			
	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	Mean	WH 147	Lok-1	Kathiya	mean	
A. mellea	38.14	40.45	31.97	36.85	36.41	32.65	21.61	30.22	37.28	36.55	26.79	33.54	
A. scrobiculata	42.49	38.60	16.58	32.56	35.90	37.28	23.37	32.19	39.20	37.94	19.97	32.37	
C. etunicatum	40.45	57.23	26.97	41.55	44.31	39.86	34.84	39.67	42.38	48.55	30.91	40.61	
G. aggregatum	43.22	50.27	19.69	37.73	36.53	37.01	27.46	33.67	39.88	43.64	23.58	35.70	
G. arborense	58.70	51.56	21.84	44.03	48.89	33.11	26.96	36.32	53.80	42.33	24.40	40.18	
G. cerebriforme	45.04	56.15	38.76	46.65	46.17	47.68	27.59	40.48	45.61	51.91	33.18	43.56	
R. diaphanus	48.05	42.36	33.28	41.23	41.61	33.74	33.93	36.43	44.83	38.10	33.60	38.83	
R. fasciculatus	53.00	41.42	35.10	43.17	48.41	39.73	32.62	40.26	50.71	40.58	33.85	41.71	
R. intraradices	56.50	49.64	33.00	46.38	48.91	56.81	39.00	48.24	52.71	53.23	36.00	47.31	
Sim. hoi	46.80	38.25	35.38	40.14	55.30	40.39	40.11	45.26	51.05	39.32	37.74	42.70	
Un-inoculated	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	
Mean	42.95	42.36	26.60	37.30	40.22	36.21	27.95	34.79					
Pooled mean	41.58	39.28	27.27										
					LSD <sub>0.05</sub>								
AM inoculation					5.35								
Variety					2.79								
Soil types					2.28								
AM inoculation × vari	iety				9.27								
AM inoculation × soil types				NS									
Variety × soil					3.95								
AM inoculation × vari	iety × soil type	S			NS								

Table 4. Effect of AM inoculations on arcsine transformed value of root colonization index of three varieties of wheat in two soil types

and it is well known that P is the most influential element in mycorrhizal development and efficiency. In P-deficient soils, the yields of plant largely depend on their mycorrhizal status (Ortas, 2003; Herrera-Peraza *et al.*, 2011). P content of alfisol (olsen P: 4.0 - 5.6 kg ha<sup>-1</sup>) is comparatively less than vertisol at the study site (olsen P: 7.6 - 23.4 kg ha<sup>-1</sup>), which might explain the obtained results.

Thus, the results of present study showed that AM fungi increased the growth, yield and P uptake of three varieties of wheat. It signifies that these can be utilized for inoculation of the crop under central Indian conditions. As, the study was conducted under sterile soil conditions, the extrapolation of the results to the real field conditions should be done after their field testing. Looking to the good response of wheat to AM fungi obtained in above mentioned experiments, attempts are being made at the institute to integrate these with other bio-inoculants (*Azotobacter* and phosphate solubilizing bacteria) and chemical fertilizers.

### ACKNOWLEDGEMENTS

The authors are thankful to the Director, ICAR-CAFRI, Jhansi for facilitating the research program and constant encouragement during the study. Ashok Shukla acknowledges funding through Science and Engineering Research Board (SB/FT/LS-366/2012), Department of Science and Technology, New Delhi.

#### REFERENCES

- Abdel-Fattah, G.M. and Asrar. A.W.A. 2012. Arbuscular mycorrhizal fungal application to improve growth and tolerance of wheat (*Triticum aestivum* L.) plants grown in saline soil. *Acta Physiol. Plant.* 34: 267–277.
- Azcon, R. and Ocampo, J.A. 1981. Factors affecting the vesiculararbuscular infection and mycorrhizal dependency of thirteen wheat cultivars. *New Phytol.* 87: 677-685.
- Behl, R.K., Sharma, H., Kumar, V. and Narula, N. 2003. Interaction amongst mycorrhiza, *Azotobacter chroococcum* and root characteristics of wheat varieties. *J. Agron. Crop Sci.* 189: 151–155.
- Bryla, I.D. and Koide, R.T. 1998. Mycorrhizal response of two tomato genotypes relates to their ability to acquire and utilize phosphorus. *Ann. Bot.* 82: 849–857.
- Carrenho, R., Trufem, S.F.B., Bononi, V.L.R. and Silva, E.S. 2007. The effect of different soil properties on arbuscular mycorrhizal colonization of peanuts, sorghum and maize. *Acta Bot. Bras.* 21:723–730.
- Dehariya, K., Shukla, A., Sheikh, I.A. and Vyas, D. 2015. *Trichoderma* and arbuscular mycorrhizal fungi based biocontrol of *Fusarium udum* Butler and their growth promotion effects on pigeon pea. *J. Agric . Sci. Technol.* 17: 505-517.
- Eason, W.R., Webb, K.J., Michaelson-Yeates, T.P.T., Abberton, M.T., Griffith, G.W., Culshaw, C.M., Hooker, J.E. and Dhanoa, M.S. 2001. Effect of genotype of *Trifolium repens* on mycorrhizal symbiosis with *Glomus mosseae*. J. Agric. Sci. 137: 27-36.
- Giovannetti, M. and Mosse, B. 1980. An evaluation of techniques for measuring vesicular arbuscular mycorrhizal infection in roots. *New Phytol.* 84: 489-500.
- Goyal, S.K., Singh, J.P. 2002. Demand versus supply of food grains in India: Implications to food security. Paper presentation at the 13<sup>th</sup> International Farm Management Congress, Wageningen, The Netherlands, July 7–12, 2002, pp. 20.

- Gryndler, M., Hrselová, H., Cajthaml, T., Havránková, M., Rezá ová, V., Gryndlerová, H. and Larsen, J. 2009. Influence of soil organic matter decomposition on arbuscular mycorrhizal fungi in terms of asymbiotic hyphal growth and root colonization. *Mycorrhiza* 19: 255–266.
- Hacisalihoglu, G., Duke, E.R. and Longo, L.M. 2005. Differential response of common bean genotypes to mycorrhizal colonization. *Pro. Florida State Hort. Soc.* 118: 150–152.
- Herrera-Peraza, R.A., Hamel, C., Fernández, F., Ferrer, R.L. and Furrazola, E. 2011. Soil-strain compatibility: the key to effective use of arbuscular mycorrhizal inoculants? *Mycorrhiza* 21: 183-193.
- Hetrick, B.A.D., Wilson, G.W.T. and Cox, T.S. 1992. Mycorrhizal dependence of modern wheat varieties, landraces and ancestors. *Can. J. Bot.* 70: 2032-2040.
- Jackson, M.L. 1973. Soil Chemical Analysis. Prentice Hall of India, New Delhi, India.
- Jha, A., Kumar, A., Saxena, R.K., Kamalvanshi, M. and Chakravarty, N. 2012. Effect of arbuscular mycorrhizal inoculations on seedling growth and biomass productivity of two bamboo species. Indian J. Microbiol. 52: 281–285.
- Karagiannidis, N. and Hadjisavva-Zinoviadi, S. 1998. The mycorrhizal fungus *Glomus mosseae* enhances growth, yield and chemical composition of a durum wheat variety in 10 different soils. *Nutr. Cycl. Agroecosys.* 52: 1–7.
- Klironomos, J.N. 2003. Variation in plant response to native and exotic arbuscular mycorrhizal fungi. *Ecology* 84: 2292–2301.
- Kormanik, P.P., Bryan, W.C. and Schultz, R.C. 1980. Procedure and equipment for staining large numbers of plant root samples for endomycorrhizal assay. *Can. J. Microbiol.* 26: 536-538.
- Minaxi Saxena, J., Chandra, S. and Nain, L. 2013. Synergistic effect of phosphate solubilizing rhizobacteria and arbuscular mycorrhiza on growth and yield of wheat plants. *J. Soil Sci. Plant Nutr.* 13: 511-525.
- Mohammad, M., Pan, W.L. and Kennedy, A.C. 1995. Wheat responses to vesicular-arbuscular mycorrhizal fungal inoculation of soils from eroded toposequences. *Soil Sci. Soc. Am. J.* 59: 1086-1090.
- Ortas, I. 2003. Effect of selected mycorrhizal inoculation on phosphorus sustainability in sterile and non-sterile soils in the harran plain in south Anatolia. *J. Plant Nutr.* 26: 1-17.
- Ortas, I. and Akpinar, C. 2006. Response of kidney bean to arbuscular mycorrhizal inoculation and mycorrhizal dependency in P and Zn deficient soils. *Acta Agr. Scand. B S P* 56: 101-109.

- Ortas, I. and Akpinar, C. 2011. Response of maize genotypes to several mycorrhizal inoculums in terms of plant growth, nutrient uptake and spore production. *J. Plant Nutr.* 34: 970–987.
- Plenchette, C., Fortin, J.A. and Furlan, V. 1983. Growth responses of several plant species to mycorrhizae in a soil of moderate P fertility. I. Mycorrhizal dependency under field conditions. *Plant Soil* 70: 199-209.
- Ramadhani, R., Damanhuri and Basuki, N. 2015. A study of mycorrhizal inoculation on some genotypes of wheat (*Triticum* aestivum L.). IOSR Journal of Agriculture and Veterinary Science 8: 42-49.
- Rivera-Becerril, F., Calantzis, C., Turnau, K., Caussanel, J.P., Belimov, A.A., Gianinazzi, S., Strasser, R.J. and Gianinazzi-Pearson, V. 2002. Cadmium accumulation and buffering of cadmium-induced stress by arbuscular mycorrhiza in three *Pisum sativum* L. genotypes. *J. Exp. Bot.* 53: 1177–1185.
- Shukla, A., Dehariya, K., Vyas, D. and Jha, A. 2014. Interactions between arbuscular mycorrhizae and *Fusarium oxysporum* f. sp. *ciceris*: Effects on fungal development, seedling growth and wilt disease suppression in *Cicer arietinum* L. Arch. *Phytopathol. Plant Prot.* 48: 240-252.
- Shukla, A., Kumar, A., Jha, A., Ajit and Rao, D.V.K.N. 2012b. Phosphorus threshold for arbuscular mycorrhizal colonization of crops and tree seedlings. Biol. Fert. Soils 48: 109-116.
- Shukla, A., Kumar, A., Jha, A., Dhyani, S.K. and Vyas, D. 2012a. Cumulative effects of tree based intercropping on arbuscular mycorrhizal fungi. *Biol. Fert. Soils* 48: 899-909.
- Shukla, A., Kumar, A., Jha, A., Chaturvedi, O.P., Prasad, R. and Gupta, A. 2009. Effects of shade on arbuscular mycorrhizal colonization and growth of crops and tree seedlings in Central India. Agroforest. Syst. 76: 95-109.
- Shukla, A., Vyas, D. and Jha, A. 2013. Soil depth: an overriding factor for distribution of arbuscular mycorrhizal fungi. J. Soil Sci. Plant Nutr. 13: 23–33.
- Singh, A.K., Hamel, C., DePauw, R.M and Knox, R.E. 2012. Genetic variability in arbuscular mycorrhizal fungi compatibility supports the selection of durum wheat genotypes for enhancing soil ecological services and cropping systems in Canada. *Can. J. Microbiol.* 58: 293-302.
- Vierheilig, H. and Ocampo, J.A. 1991. Susceptibility and effectiveness of vesicular-arbuscular mycorrhizae in wheat cultivars under different growing conditions. *Biol. Fert. Soils* 11: 290-294.