



Effect of elevated ozone and carbon dioxide on growth and yield of rice (*Oryza sativa*)

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Received: 18 August 2020; Accepted: 16 April 2021

ABSTRACT

An experiment was conducted by growing Pusa 44 rice (*Oryza sativa* L.) variety under different treatments: Ambient, Elevated O₃ (EO₃: 60 ± 10 ppb), Elevated CO₂ (ECO₂: 550 ± 10 ppm) and Elevated CO₂ and O₃ (550 ± 10 ppm CO₂ and 60 ± 10 ppb EO₃) in free air carbon dioxide and ozone enrichment facility (FACE-O₃) with the objective to assess the impact of elevated tropospheric ozone (O₃) and carbon dioxide (CO₂) interaction on growth and yield of rice. The crop was exposed to elevated levels of gases from transplanting to maturity. EO₃ was found to be having a negative impact on all the growth parameters at vegetative, anthesis and maturity stages. ECO₂ was found to be having a compensatory effect over EO₃ for the different growth parameters. On an average the compensatory effect of ECO₂ over EO₃ on yield was about 40% and negative impact of elevated O₃ over control was around 26% in both the years.

Keywords: Elevated CO₂, Elevated O₃, Growth stage, Rice, Yield

Increasing ozone (O₃) and carbon dioxide (CO₂) concentration in the atmosphere are of major concern (Wang *et al.* 2017) due to its direct effect on agriculture. Ground level O₃ concentrations are increasing at the rate of 1-2% per year (Wang and Frie 2011) and may reach 60-70 ppb by 2100 (IPCC 2014), whereas atmospheric CO₂ is currently 415 ppm and is expected to reach 700 ppm by 2100 (NOAA 2019). The most rapid increase in O₃ is occurring in south Asia and may result in crop production losses (Tiwari and Agrawal 2018).

Rice (*Oryza sativa* L.) is the widely grown crop globally, occupying about 161.8 Mha area, of which 10.5 Mha lies in the rice-wheat system of the Indo-Gangetic Plains (Gupta *et al.* 2016). Ozone is known to impact stomatal conductivity (g_s), net CO₂ assimilation and carboxylation efficiency (Singh *et al.* 2013). Elevated O₃ causes physiological changes in crops, which are translated into morphological changes and reduces plant growth (Bhatia *et al.* 2013). Decline in photosynthetic rate (P_N) in the plants exposed to O₃ stress is associated with damage to photosynthetic machinery (Daripa *et al.* 2016), biophysical parameters such as g_s and internal CO₂ also play important roles in determining the photosynthetic yield and sensitivity of O₃

exposed plants. Under CO₂ rich and warm atmosphere, plants may show a tendency to reduce g_s, thus, indirectly alleviate the O₃ stress in plants (Yadav *et al.* 2019). Thus the objective of this study was to quantify the effect of elevated O₃ and CO₂ interaction on growth and yield of rice.

MATERIALS AND METHODS

Experimental site and soil: A field experiment was conducted at the farm of ICAR-Indian Agricultural Research Institute (ICAR-IARI), New Delhi by growing Pusa 44 rice genotype in *kharif* (July–Mid November) during 2016–17. The soil at the experimental site belongs to the major group of Indo-Gangetic alluvium (Typic Haplustept), non-calcareous and slightly alkaline in reaction. The climatic variables and soil properties of the experimental site were as in Bhatia *et al.* (2021).

Treatments and crop management: This experiment was carried out in under free air Carbon Dioxide and Ozone enrichment (FACE-O₃) in four 6-m diameter circular plots with perforated horizontal tubings, with one ambient plot (ACO₂ + AO₃) and three plots each receiving elevated CO₂ (ECO₂+ AO₃), elevated O₃ (EO₃ + ACO₂) and both ECO₂ and EO₃. CO₂ and O₃ were released through horizontal perforated tubing's above the soil surface at canopy level to attain the target CO₂ (550 ppm) and O₃ (60 ppb) concentration. The O₃ concentration was measured using an O₃ concentration analyzer (2B technologies) and CO₂ analyser (Fuji).

Transplanting of seedlings of rice sown on 20 June was carried out in 5 replicate crates (size 0.24 m²) on 12 July in 2016 and 16 July in 2017. EO₃ and ECO₂ exposure in

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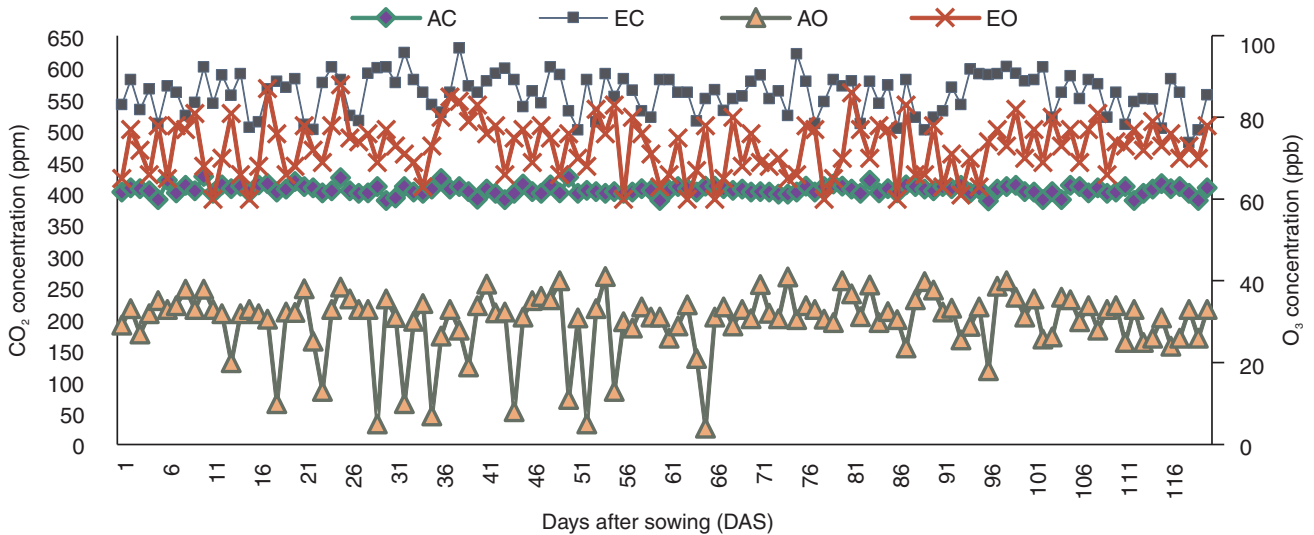


Fig 1 Daily average concentrations of ambient CO₂ (AC), elevated CO₂ (EC), ambient O₃ (AO) and elevated O₃ (EO) during rice growth period in 2016 and 2017.

rings began on 19 July 2016 and 23 July 2017 and ended on 29 October, 2016 and 1 November 2017, respectively. Urea at the rate of 12 g/m² was added in three splits of 6, 3 and 3 g/m² at 0, 35 and 70 days after transplanting (DAT). The rice soil was saturated with water till 85 DAT. Weeds, pests and diseases were controlled as required.

Plant sampling and analysis: Plant samples were collected for recording of biomass of different plant parts at three different growth stages, viz. active tillering, anthesis and maturity stage. For taking the dry weight of different plant parts, they were kept in a pre-heated hot air oven 65°C for 72 hr till a constant dry weight was achieved.

Data analysis: Statistical analysis of the data was done using SPSS 23.0 (SPSS Inc., Chicago, USA) for windows. Analysis of variance (ANOVA) was done to test whether the differences were statistically significant using completely randomized design.

RESULTS AND DISCUSSION

Concentration of CO₂ and O₃ in the rings: The season-long daytime average CO₂ concentration in the ambient and in the elevated plots was 404.6 ± 4.6 ppm and 557.8 ± 11.2 ppm, respectively. The target O₃ concentration in FACE-O₃ was higher (62.6 ± 5.5 ppb) than the ambient (30.1 ± 4.3 ppb) (Fig 1).

Leaf area: Total leaf area ranged from 1263.17 cm²/hill in ACO₂+EO₃ at tillering to 3467.88 cm²/hill in ECO₂+AO₃ at maturity stage in

2016 and 1546.92 cm²/hill in ECO₂+EO₃ during tillering to 3544.77 cm²/hill in ECO₂+AO₃ during maturity stage in 2017 (Fig 2). Negative impact of EO₃ on leaf area was most pronounced at tillering followed by anthesis and maturity stages. Elevated CO₂ has been found to increase the leaf area (Kimball *et al.* 2016) and EO₃ has been found to decrease the leaf area (Yadav *et al.* 2020). Leaf area is a basis of measuring productivity during the growth and development under different environmental condition.

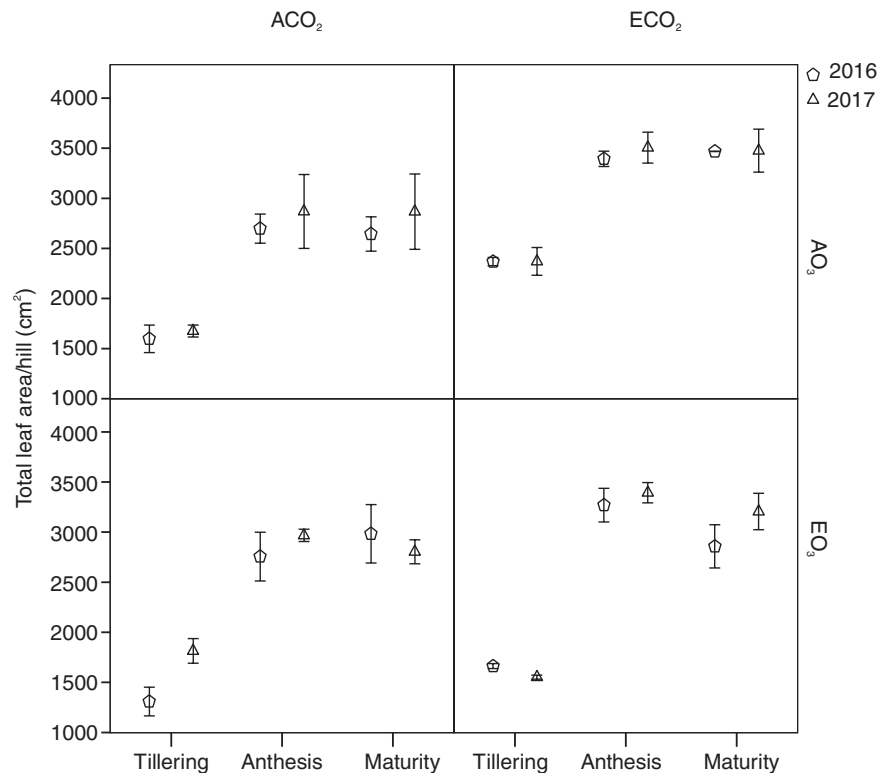


Fig 2 Effect of different treatments of CO₂ and O₃ on total leaf area/hill of rice at tillering, anthesis and maturity stages during the year 2016 and 2017. ACO₂; Ambient CO₂, ECO₂; Elevated CO₂, AO₃; Ambient O₃, EO₃; Elevated O₃.

Table 1 Effect of different treatments of CO₂ and O₃ on growth parameters of rice during the years 2016 and 2017

Treatment	DW (leaves) (g/hill)				DW (Culms) (g/hill)				DW (roots) (g/hill)				Plant height (cm)				No. of tillers/hill															
	T	A	M	T	T	A	M	T	T	A	M	T	T	A	M	T	T	A	M	T												
<i>2016</i>																																
ACO ₂ +AO ₃	12.65	15.67	19.94	11.90	10.44	13.17	30.15	29.93	28.66	84.00	102.50	100.00	27.50	33.50	34.00	8.65	15.69	20.76	8.25	10.56	16.65	8.95	15.61	21.06	83.50	96.50	103.00	23.50	28.50	28.50		
ECO ₂ +AO ₃	13.15	23.61	30.01	12.25	17.41	26.76	19.95	48.38	37.42	94.00	107.50	100.50	22.00	30.00	28.00	9.25	14.71	20.21	8.55	15.99	21.61	21.00	23.01	29.03	89.50	108.50	117.50	22.50	33.00	34.50		
SE (d)	1.33	3.40	3.78	2.06	2.01	2.18	5.02	7.28	5.29	2.90	3.12	4.07	1.50	2.72	1.81	2.90	2.10	2.10	4.50	4.39	4.76	10.94	15.88	11.53	6.32	6.80	8.87	3.26	5.93	3.95		
LSD (P=0.05)																																
CO ₂	**	**	NS	**	**	**	**	**	NS	**	**	NS	**	**	NS	**	**	NS	NS	NS	**	**	**	**	**	**	**	NS	**	**	NS	
O ₃	**	**	NS	NS	NS	**	**	**	NS	NS	**	**	**	**	**	**	**	NS	NS	NS	**	**	**	**	**	**	**	**	**	**	**	
CO ₂ × O ₃	NS	NS	NS	**	**	**	NS	NS	NS	**	**	NS	**	**	**	**	**	NS	NS	NS	**	**	**	**	**	**	**	**	**	**	**	
Impact of EO ₃ (%) over Control	-46.2	0.1	3.9	-44.2	1.1	20.9	-236.9	-91.7	-36.1	-0.6	-6.2	2.9	-17.0	-17.5	-19.3																	
Compensation effect of ECO ₂ (%) over EO ₃	6.9	-6.2	-2.6	3.6	51.4	29.8	134.6	47.4	37.8	7.2	12.4	14.1	-4.3	15.8	21.1																	
<i>2017</i>																																
ACO ₂ +AO ₃	11.05	17.18	25.00	8.70	11.00	12.44	22.35	21.00	31.46	82.00	103.50	107.00	27.00	28.50	32.50	9.90	13.05	20.39	8.25	11.96	17.34	4.60	16.81	20.65	79.50	101.50	104.00	25.00	30.50	30.00		
ECO ₂ +AO ₃	14.60	22.72	29.51	15.60	16.83	25.85	20.75	51.96	35.19	91.50	108.00	102.00	25.00	31.50	28.50	7.65	16.35	19.51	8.45	15.89	21.42	10.40	26.96	30.35	86.00	106.50	107.50	25.00	30.50	33.00		
SE (d)	0.56	0.85	1.41	0.50	0.57	1.32	1.07	1.45	1.50	2.08	2.38	2.47	1.36	1.59	1.67	1.23	1.85	3.08	1.10	1.25	2.88	2.34	3.16	3.27	4.53	5.20	5.39	2.98	3.47	3.64		
LSD (P=0.05)																																
CO ₂	**	**	NS	**	NS	**	**	**	NS	**	**	NS	**	**	NS	**	**	NS	**	**	**	**	**	**	**	**	**	**	**	**	NS	
O ₃	**	**	NS	**	**	NS	**	**	**	**	**	**	**	**	**	**	**	NS	**	**	**	**	**	**	**	**	**	**	**	**	**	NS
CO ₂ × O ₃	NS	**	**	**	**	NS	**	**	**	NS	NS	**	**	**	**	NS	**	**	**	**	**	**	**	**	**	**	**	**	**	**	**	**
Impact of EO ₃ (%) over Control	-11.6	-31.6	-22.6	-5.5	8.0	28.3	-385.9	-24.9	-52.3	-3.1	-2.0	-2.9	-8.0	6.6	-8.3																	
Compensation effect of ECO ₂ (%) over EO ₃	-22.7	25.3	-4.3	2.4	32.9	23.5	126.1	60.4	47.0	8.2	4.9	3.4	0.0	0.0	10.0																	

Values are means. NS, not significant; ** P < 0.05. T; Tilling stage, A; Anthesis stage, M; Maturity stage, ACO₂+AO₃; Ambient CO₂+Ambient O₃; ACO₂+EO₃; Ambient CO₂+Elevated O₃; ECO₂+AO₃; Elevated CO₂+Ambient O₃; ECO₂+EO₃; Elevated CO₂+Elevated O₃.

Plant height: Plant height ranged from 79.5 cm in ACO₂+EO₃ at tillering to 117 cm in ECO₂+EO₃ at maturity in the two years (Table 1 and 2). Plant height of varieties is generally genetically specified but many studies on plant height showed that the increase of O₃ concentration generally reduces the plant height of rice.

Number of tillers: The tillers/hill increased from active tillering to anthesis and then decreased at maturity stage under different treatments (Table 1 a, b). The average tillers/hill in ECO₂ rings was always higher when compared with other treatments. Reduced tiller number under O₃ stress has been also reported in other studies (Tomer *et al.* 2015). Juan *et al.* (2021) reported significant increase in number of tillers under ECO₂ condition as compared to ambient due to redistribution of nitrogen to promote tillering.

Dry matter production: EO₃ has been found to be having negative impact on root and shoot biomass. O₃ generally reduces plant growth and the amount of dry biomass (Bhatia *et al.* 2011). Under different treatments the growth parameters of dry weight (DW) of leaves/hill, DW of culms/hill and DW of roots/hill were negatively affected under EO₃. Under ECO₂+EO₃ treatment, ECO₂ was found to be having compensatory effect on EO₃ (Table 1a, b). In both the years highest compensatory effect was observed in DW of roots (134.6% in 2016 and 126.08% in 2017) at tillering. DW of roots was larger under ECO₂+AO₃ treatment at all growth stages. It has been reported that EO₃ enhances carbon allocation to reproductive parts during anthesis, compromising the growth of roots (Emberson *et al.* 2018) which was evident in this study where under EO₃ the DW of roots reduced at different growth stages. Higher O₃ levels reduce root growth by altering photosynthetic partitioning

(Bhatia *et al.* 2012). Reproductive stage is considered as most O₃ sensitive however; our results suggest that higher O₃ may reduce the dry mass of the leaf, root, and whole-plant even during the tillering stage.

Yield: Grain yield of rice ranged from 359 g/m² in ACO₂+EO₃ to 687 g/m² in ECO₂+AO₃ in during the two years. Compensatory effect of ECO₂ over EO₃ on yield was above 40% (Table 2) and negative impact of EO₃ over control was above 26%. ECO₂ interactively ameliorates the inhibition of photosynthesis induced by O₃ exposure (Kobayakawa and Imai 2011, Bhatia *et al.* 2021) that results in compensation of yield. The decline in grain yield was associated with decrease in various growth parameters under elevated O₃ conditions (Singh *et al.* 2017). O₃ from the environment enters into plants generally through stomata; encounters with apoplastic antioxidants which causes plasma membrane dysfunction which leads to membrane leakage. This causes metabolic dysfunction resulting in physiological changes including decreased photosynthesis and stomatal conductance and increased respiration rates (Bhatia *et al.* 2012). These physiological changes reduce carbon assimilation, plant growth and yield (Emberson *et al.* 2018). Harvest Index was found to decrease by approximately 6% and 4% under EO₃ condition as compared to control (Table 2). A meta-analysis by Ainsworth (2008) showed that the high O₃ concentration may decrease the harvest index in rice by 5%.

Elevated O₃ reduced the dry mass of the leaf, root, and whole-plant at both the tillering and flowering stages. Elevated O₃ significantly reduced the grain yield of rice. However, elevated CO₂ countered the yield decrease. The compensatory effect of ECO₂ over EO₃ on yield was above 40%.

Table 2 Grain yield, dry biomass and harvest index of rice during the years 2016 and 2017 under different treatments of CO₂ and O₃

Treatment	2016			2017		
	Grain yield (g/m ²)	Dry biomass (g/m ²)	Harvest Index (%)	Grain yield (g/m ²)	Dry biomass (g/m ²)	Harvest Index (%)
ACO ₂ +AO ₃	500.97	1557.22	32.17	453.33	1797.36	25.24
ACO ₂ +EO ₃	397.22	1522.36	26.09	359.03	1651.94	21.84
ECO ₂ +AO ₃	687.08	2224.31	30.93	631.25	2066.67	30.60
ECO ₂ +EO ₃	670.56	2506.67	26.76	603.06	1872.64	32.29
SE (d)	20.01	51.49	1.25	14.93	72.38	1.63
LSD (P=0.05)	41.31	106.21	2.59	30.81	149.39	3.37
<i>ANOVA results</i>						
CO ₂	**	**	**	**	**	**
O ₃	**	**	**	**	**	**
CO ₂ × O ₃	**	**	NS	**	NS	**
Impact of EO ₃ (%) over Control	-26.1	-2.3	-23.3	-26.3	-8.8	-15.6
Compensation effect of ECO ₂ (%) over EO ₃	40.8	39.3	2.5	40.5	11.8	32.4

Values are means. NS, not significant, **P<0.05. n=5. ACO₂+AO₃; Ambient CO₂+Ambient O₃, ACO₂+EO₃; Ambient CO₂+Elevated O₃, ECO₂+AO₃; Elevated CO₂+Ambient O₃, ECO₂+EO₃; Elevated CO₂+Elevated O₃

ACKNOWLEDGEMENTS

We are grateful to PG School, Dean and Director, IARI, New Delhi for providing field and facility for conducting experiments and University Grant Commission (UGC), New Delhi, for providing fellowship to the first author during his Ph D programme. The financial support by the National Innovations in Climate Resilient Agriculture project (NICRA) is highly acknowledged.

REFERENCES

- Ainsworth E A. 2008. Rice production in a changing climate: a meta-analysis of responses to elevated carbon dioxide and elevated ozone concentration. *Global Change Biology* **14**: 1642–50.
- Bhatia A, Ghosh A, Kumar V, Tomer R, Singh S D and Pathak H. 2011. Effect of elevated tropospheric ozone on methane and nitrous oxide emission from rice soil in north India. *Agriculture Ecosystems and Environment* **144**(1): 21–8.
- Bhatia A, Tomer R, Kumar V, Singh S D and Pathak H. 2012. Impact of tropospheric ozone on crop growth and productivity - a review. *Journal of Scientific and Industrial Research* **71**: 97–112.
- Bhatia A, Kumar V, Kumar A, Tomer R, Singh B and Singh S D. 2013. Effect of elevated ozone and carbon dioxide interaction on growth and yield of maize. *Maydica* **58**: 291–29.
- Bhatia A, Mina U and Kumar V. 2021. Effect of elevated ozone and carbon dioxide interaction on growth, yield, nutrient content and wilt disease severity in chickpea grown in Northern India. *Heliyon* **7**: e06049. <https://doi.org/10.1016/j.heliyon.2021.e06049>
- Daripa A, Bhatia A and Ojha S. 2016. Chemical and natural plant extract in ameliorating negative impact of tropospheric ozone on wheat crop: a case study in a part of semiarid north west India. *Aerosol and Air Quality Research* **16**: 1742–56.
- Emberson L D, Pleijel H, Ainsworth E A, Van den Berg M, Ren W, Osborne S, Mills G, Pandey D, Dentener F, B ker P and Ewert F. 2018. Ozone effects on crops and consideration in crop models. *European Journal of Agronomy* **100**: 19–34.
- Gupta D K, Bhatia A, Kumar A, Das T K, Jain N, Tomer R, Malyan S K, Fagodiya R K, Dubey R and Pathak H. 2016. Mitigation of greenhouse gas emission from rice-wheat system of the Indo-Gangetic plains: Through tillage, irrigation and fertilizer management. *Agriculture Ecosystem Environment* **230**: 1–9.
- IPCC. 2014. Climate change The physical science basis. (In) *Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Stocker T F, Qin D, Plattner M, Tignor S K, Allen J, Boschung A, Nauels Y, Bex V, Midgley P M (Eds.). Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, p 1535.
- Juan Z, Yingbo G, Junpeng W, et al. 2021 Elevated atmospheric CO₂ concentration triggers redistribution of nitrogen to promote tillering in rice. *Plant Environment Interactions*. **2**: 125–136.
- Kobayakawa H and Imai K. 2011. Effects of the interaction between ozone and carbon dioxide on gas exchange, photosystem II and antioxidants in rice leaves. *Photosynthetica* **49**(2): 227–38.
- Kimball BA. 2016. Crop responses to elevated CO₂ and interactions with H₂O, N, and temperature. *Current Opinion in Plant Biology* **31**: 36–43.
- NOAA. 2019. Trends in atmospheric carbon dioxide- Earth System Research Laboratory. Global Monitoring Division - NOAA. <http://www.esrl.noaa.gov/gmd/ccgg/trends/> (accessed December 2019).
- Paoletti E and Grulke N E. 2005. Does living in elevated CO₂ ameliorate tree response to ozone? A review on stomatal responses. *Environmental Pollution* **137**: 483–93.
- Shurpali N, Agarwal A K and Srivastava V K. 2019. Introduction to greenhouse gas emissions. (In) *Greenhouse gas emissions: challenges, technologies and solutions*. Energy, environment and sustainability series, Agarwal A K and Pandey A (Eds.). https://doi.org/10.1007/978-981-13-3272-2_1
- Singh S, Bhatia A, Tomer R, Kumar V, Singh B and Singh S D. 2013. Synergistic action of tropospheric ozone and carbon dioxide on yield and nutritional quality of Indian mustard (*Brassica juncea* (L.) Czern.). *Environment Monitoring and Assessment* **185**: 6517–29.
- Singh R N, Mukherjee, J, Sehgal V K and Bhatia A. 2017. Effect of elevated ozone, carbon dioxide and their interaction on growth, biomass and water use efficiency of chickpea (*Cicer arietinum* L.). *Journal of Agrometerology* **19**: 301–05.
- Tiwari S and Agrawal M. 2018. Tropospheric ozone and its impacts on crop plants: A threat to future global food security. Springer.
- Tomer R, Bhatia A, Kumar V, Kumar A, Singh R, Singh B and Singh S D. 2015. Impact of elevated ozone on growth, yield and nutritional quality of two wheat species in Northern India. *Aerosol and Air Quality Research* **15**: 329–40.
- Wang B, Shugart H H and Lerdau M T. 2017. Sensitivity of global greenhouse gas budgets to tropospheric ozone pollution mediated by the biosphere. *Environmental Research Letters* **12**(8): 084001.
- Wang Y and Frei M. 2011. Stressed food–The impact of abiotic environmental stresses on crop quality. *Agriculture Ecosystems and Environment* **141**: 271–86
- Yadav A, Bhatia A, Yadav S, Kumar V and Singh B. 2019. The effects of elevated CO₂ and elevated O₃ exposure on plant growth, yield and quality of grains of two wheat cultivars grown in north India. *Heliyon* **5**: e02317.
- Yadav A, Bhatia A, Yadav S, Singh A, Tomer R, Kumar V and Singh B. 2020. Growth, yield and quality of maize under ozone and carbon dioxide interaction in North West India. *Aerosol and Air Quality Research* **20**: <https://doi.org/10.4209/aaqr.2020.05.0194>