GC-MS analysis of metabolites in tritrophic interaction of yeast (*Pichia Kudriavzevii*) against soft rot pathogen of carrot (*Daucus carota*)

S LAVANYA¹, P MUTHULAKSHMI² and I JOHNSON³

Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu 641 003, India

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Carrot (Daucus carota L.) is an important root vegetable cultivated worldwide, valued for their high levels of alpha and beta carotene, essential vitamins A, K, and B₆, as well as minerals that enhance eye health. They are widely used in various culinary dishes (Kitinoja et al. 2018). However, carrots are susceptible to postharvest diseases during transportation, handling, and storage. Yeasts have been utilized to control these diseases, supported by successful reports from various studies. Yeasts are beneficial microorganisms known for their strong antagonistic properties, environmental friendliness, and safety for human consumption (Zhang et al. 2020). These single-celled fungi thrive in diverse environments, do not produce harmful metabolites, and can be easily cultured in large quantities. They can colonize fruit surfaces for extended periods and resist chemical pesticides commonly used in postharvest disease management (Spadaro and Droby 2016). Liu et al. (2020) demonstrated that the yeast strain Pichia kudriavzevii effectively inhibited the growth of Botrytis cinerea, Alternaria alternata, and Rhizopus stolonifer in cherry tomatoes. In carrots, the primary diseases are soft rot caused by Pectobacterium carotovorum subsp. carotovorum and sour rot caused by Geotrichum candidum. This study focused on the metabolites extracted from soft rot pathogen, antagonist, and their interaction, which were analyzed through GC-MS. The metabolites from the effective yeast isolate were found to play a role in managing postharvest diseases in carrots.

A study was carried out during 2022–23 at Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu. Bacterial strain *P. carotovorum* subsp. *carotovorum*, isolated from carrot soft rot symptoms, and yeast strain *P. kudriavzevii*, isolated from the carrot surface, were cultivated separately in YEPDA broth. Both isolates were then co-inoculated in YEPDA broth and incubated for 48 h with continuous

¹Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu. *Corresponding author email: muthupathology@gmail.com

agitation. For metabolite extraction, the actively growing cultures were centrifuged at 12,000 rpm for 15 min. The supernatant was collected and filtered through Whatman No. 1 filter paper. The filtrate was mixed with an equal volume of ethyl acetate and shaken overnight. The ethyl acetate layer was separated using a separating funnel and collected in a separate container. This layer was then evaporated using a vacuum flask evaporator at 60°C and 80 rpm, resulting in a concentrated residue. The residue was dissolved in HPLC-grade methanol for analysis with Gas Chromatography-Mass Spectrometry (GC-MS) (Sasidharan *et al.* 2012).

Effect of crude metabolites by effective yeast isolates against P. carotovorum subsp carotovorum: The antagonistic activity of P. kudriavzevii against P. carotovorum subsp. carotovorum was tested using the agar well diffusion method. A petri dish containing Nutrient Agar (NA) medium was prepared, and a bacterial lawn was established by evenly spreading a bacterial suspension with an L-shaped rod and allowing it to air-dry. Four wells, each 1 cm from the edge of the plate, were made using a cork borer. The extracted metabolite was added to the wells in volumes of 50, 100, 150, and 200 μL, while sterile distilled water served as a control. An inhibition zone was observed around the wells containing the metabolite (Chiranjeevi et al. 2021).

Detection of crude metabolites through GC-MS analysis: The extracted crude metabolites were analysed using GC-MS. An Elite-5MS column (30 mm \times 0.25 mm, 0.25 μ m thickness) was used, with helium as the carrier gas at a flow rate of 1 ml/min. The column temperature was initially set at 40°C for 2 min, then increased to 160°C at 6°C/min, and finally raised to 260°C at 10°C/min, where it was held for 4 min. The ion source temperature was 230°C, with electron energy of 70 eV, and the mass range scanned from 40–450 Da. Compound identification was performed by comparing the spectra with the NIST MS Data library using computer-based sources (Mulatu et al. 2022).

Statistical analysis: The experiments were analysed through completely randomized design (CRD). The mean differences of the treatment were compared using Duncan's Multiple Range-Test (DMRT) at 5% significance level

(Gomez and Gomez 1984). All the data were analysed statistically using SPSS software (version 16).

Effect of crude metabolites of P. kudriavzevii YKP 12 against P. carotovorum subsp. carotovorum: The study investigated the impact of crude metabolites from Pichia kudriavzevii YKP 12 on P. carotovorum sub sp. carotovorum using an agar well diffusion assay. The metabolites exhibited varying levels of inhibition, with the highest inhibition zone of 18.33 mm observed at 100 μ L and the lowest of 5.66 mm at 50 μ L against P. carotovorum subsp. carotovorum (Table 1, Fig. 1). Previous studies, such as those by Sheeba et al. (2019), also utilized the agar well diffusion method to assess similar antibacterial activity.

Detection of crude metabolites through GC-MS analysis: GCMS analysis of metabolites extracted from P. carotovorum subsp. carotovorum: GC-MS analysis of bacterial isolates identified three major compounds: Carbamic acid phenyl ester, 1-Hexanol 2-ethyl, and Hexadecanoic acid methyl ester. Among these, 1-Hexanol 2-ethyl had the highest peak area at 13.56% with a retention time of 4.8 min, while 1-Octanol 2, 7-dimethyl had the lowest peak area at 0.5% with a retention time of 5 min.

Similar findings by Meziani *et al.* (2015) linked compounds like hexadecenoic acid trimethylsilyl ester to pathogenesis in potato tubers infected with *P. atrosepticum*.

Marquez-Villavicencio *et al.* (2021) noted that *P. carotovorum* subsp. *carotovorum* strain PCC21 from Chinese cabbage exhibited butanoate metabolism, contributing to a rotting odor.

GCMS analysis of metabolites extracted from yeast YKP12 isolate: GC-MS analysis of Pichia kudriavzevii identified three major compounds: 2,4-bis (1,1-dimethylethyl) phenol, 1,2-benzenedicarboxylic acid, and hexamethyl cyclotrisiloxane. The most prominent compound was 1, 2-benzenedicarboxylic acid, which accounted for 13.09% of the peak area and had a retention time of 16.130 min. The compound with the smallest peak area, Cyclotrisiloxane (hexamethyl), was recorded at 0.67% with a retention time of 15.643 minutes (Fig. 2). Research by Zhao et al. (2023) on the aroma-producing yeast Wickerhamomyces anomalus identified metabolites such as phenylethyl alcohol, isobutyl alcohol, isothiocyanate esters, myrcene, and dimethyl disulfide, highlighting the best ether compound found in an ethyl acetate mixture. Additionally, Hannaella sinensis has demonstrated

Table 1 Effect of crude metabolites of *P. kudriavzevii* YKP 12 against *P. carotovorum* sub sp *carotovorum* by agar well diffusion assay

Yeast isolate	Concentrations (µl)	Inhibition zone (mm)
YKP12	25	5.66 ^d
	50	12.67 ^c
	75	15.66 ^b
	100	18.33 ^a
	Control	$0.00^{\rm e}$

Mean followed by a common letter as superscript does not differ significantly at 5% level by DMRT. Values in parentheses are are sine transformed values.

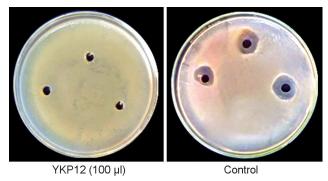


Fig. 1 Effect of crude metabolites of *P. kudriavzevii* YKP 12 against *P. carotovorum* sub sp *carotovorum*.

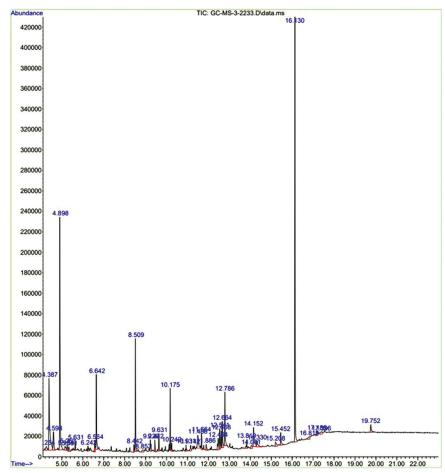


Fig. 2 GCMS analysis of metabolites extracted from Pichia kudriavzevii YKP 12 isolate.

Molecular Compound RT Area Molecular Structure Biological activity Reference (%)weight formula 1-Hexanol, 2-ethyl-13.89 130.2279 $C_8H_{18}O$ Antifungal, Nematicidal (Hewavitharana et 4.898 al. 2014) Heptocosane 5.154 0.60 380.7 $C_{27}H_{56}$ Antimicrobial (Marrufo et al. 2013) Antioxidant Tricosanoic acid, 13.819 0.77 368.6367 $C_{24}H_{48}O_{2}$ Antimicrobial (Valiei et al. 2011) methyl ester C₁₂H₂₂Si₂ 1,2-Bis (trimethylsiyl) 15.208 0.80 222.47 Antioxidant Antimicrobial (Momin and benzene Thomas 2020) $C_{12}H_{16}N_2O_3$ Cyclobarbital 14.041 0.65 236.27 Antioxidant Antimicrobial (Joshi et al. 2020) Pyrrolo [1,2-a]pyrazine-12.664 2.34 244.29 $C_{10}H_{16}N_2O_2$ Antioxidant (Manimaran and 1,4-dione, hexahydro-Kannabiran 2017) 3-(phenyl methyl)-Compound RT Molecular Molecular Structure Biological activity Reference Area (%) weight formula Cyclopentane 13.819 0.22 268.50 $C_{18}H_{32}O_{2}$ Antibacterial (Kawahara et al.

 $C_{16}H_{34}$

Table 2 GCMS analysis of metabolites extracted from interaction of *P. kudriavzevii* YKP 12 isolate and *P. carotovorum* sub sp. carotovorum

effectiveness in managing post-harvest diseases in apples by degrading patulin (PAT) through intracellular enzymes (Ma *et al.* 2023).

12.430

0.17

226.44

tridecanoic acid

Hexadecane

GCMS analysis of metabolites extracted from the interaction of Pichia kudriavzevii YKP12 isolate and P. carotovorum subsp carotovorum: In the bipartite interaction analysis, four bioactive compounds were identified: 1-Hexanol, 2-ethyl, Heptacosane, and Pyrrolo [1,2-a] pyrazine-1,4-dione, as well as 1,2-Bis(trimethylsilyl) benzene. Among these, 1-Hexanol, 2-ethyl exhibited the highest peak area at 13.89% with a retention time of 4.89 min. Pyrrolo [1,2-a] pyrazine-1,4-dione showed a peak area of 2.34% and a retention time of 12.66 min, while Heptacosane had the lowest peak area at 0.60% (Table 2).

SUMMARY

Carrot a fleshy edible root vegetable, abundant in bioactive compounds such as carotenoids and dietary fibres. The market value of carrots is impacted by post-harvest illnesses, which have emerged as a serious production constraint. A promising advancement in the battle against post-harvest illnes is yeast-based biocontrol, because of its colonization on dry surfaces for longer periods and low nutrient utilization and can multiply easily. A study was carried out during 2022-23 at Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu to evaluate the GC-MS analysis of metabolites in tritrophic interaction of yeast (Pichia Kudriavzevii) against soft rot pathogen of carrot. The tritrophic interaction of soft rot pathogen Pectobacterium carotovorum subsp. carotovorum and yeast isolate YKP 12 Pichia kudriavzevii was analyzed through GCMS analysis and the metabolite 1-Hexanol, 2-ethyl showed highest peak area of 13.89% with the retention time of 4.89 min. The tri-trophic interaction results revealed that the compound 1, 2-Benzenedicarboxylic acid monoester had both antibacterial and antifungal activity.

Antibacterial, Antioxidant (Yogeswari et al.

1996)

REFERENCES

Chiranjeevi N, Reddi Kumar M, Padmodaya B, Venkateswarlu N C, Sudhakar P, Sarada Jayalakshmi Devi R and Aruna Sri P. 2021. Studies on extraction, evaluation of crude metabolite extract from endophytic *Bacillus subtillis* and its mechanistic effect on chickpea dry root rot causing pathogen *Rhizoctonia bataticola* (Taub.) Butler. *Pharma Innovation* 10: 898–905.

Gomez K A and Gomez A A. 1984. Statistical Procedures for Agricultural Research, 2nd edn, pp. 680. John Wiley and Sons, New York.

Hewavitharana S S, Ruddell D and Mazzola M. 2014. Carbon source-dependent antifungal and nematicidal volatiles derived during anaerobic soil disinfestation. *European Journal of Plant Pathology* **140**: 39–52.

Joshi S, Amatya S, Pandey R D, Khadka P and Bhattarai J. 2020. Antimicrobial, antioxidant, antidiabetic, cytotoxic activities and GC-MS analysis of methanolic extract of *Elaeocarpus* sphaericus leaves from Nepal. International Journal of Advanced Research in Chemical Science 8(1): 11–23.

Kawahara H, Hirai F and Obata H. 1996. Antibacterial effect of ω-Cyclopentane fatty acids synthesized by *Bacillus subtilis* 626 from cyclopentane carboxylic acid. *Biocontrol Science* 1(1): 69–72.

Kitinoja L, Tokala V Y and Brondy A. 2018. Challenges and opportunities for improved postharvest loss measurements in plant-based food crops. *Journal of Postharvest Technology* **6**(4): 16–34.

Liu X, Gao Y, Yang H, Li L, Jiang Y, Li Y and Zheng J. 2020. *Pichia kudriavzevii* retards fungal decay by influencing the

- fungal community succession during cherry tomato fruit storage. *Food Microbiology* **88**: 103404.
- Ma J, Godana E A, Yang Q and Zhang H. 2023. Effect of the antagonistic yeast *Hannaella sinensis* on the degradation of Patulin. *Biological Control* 178: 105134.
- Manimaran M and Kannabiran K. 2017. Marine Sp. VITMK1 derived Pyrrolo [1, 2-A] Pyrazine-1, 4-Dione, Hexahydro-3-(2-Methylpropyl) and its free radical scavenging activity. *The Open Bioactive Compounds Journal* **5**(1).
- Marquez-Villavicencio M D P, Weber B, Witherell R A, Willis D K and Charkowski A O. 2011. The 3-hydroxy-2-butanone pathway is required for *Pectobacterium carotovorum* pathogenesis. *PloS One* 6(8): e22974.1.
- Marrufo T, Nazzaro F, Mancini E, Fratianni F, Coppola R, De Martino L and De Feo V. 2013. Chemical composition and biological activity of the essential oil from leaves of *Moringa oleifera* Lam. cultivated in Mozambique. *Molecules* 18(9): 10989–11000.
- Meziani S, Oomah B D, Zaidi F, Simon-Levert A, Bertrand C and Zaidi Yahiaoui R. 2015. Antibacterial activity of carob (*Ceratonia siliqua* L.) extracts against phytopathogenic bacteria *Pectobacterium atrosepticum*. *Microbial Pathogenesis* **78**: 95–102.
- Momin K and Thomas S C. 2020. GC-MS analysis of antioxidant compounds present in different extracts of an endemic plant *Dillenia scabrella* (Dilleniaceae) leaves and barks. *International Journal of Pharmaceutical Sciences and Research* 11: 2262–73.

- Mulatu A, Megersa N, Tolcha T, Alemu T and Vetukuri R R. 2022. Antifungal compounds, GC-MS analysis and toxicity assessment of methanolic extracts of *Trichoderma* species in an animal model. *PloS One* 17(9): e0274062.
- Sasidharan K, Soga T, Tomita M and Murray D B. 2012. A yeast metabolite extraction protocol optimised for time-series analyses. e44283.
- Sheeba H, Ali M S and Anuradha V. 2019. Bioactive compounds and antimicrobial activity of fungal crude extract from medicinal plants. *Journal of Pharmaceutical Sciences and Research* 11(5): 1826–33.
- Spadaro D and Droby S. 2016. Development of biocontrol products for postharvest diseases of fruit: The importance of elucidating the mechanisms of action of yeast antagonists. *Trends in Food Science and Technology* **47**: 39–49.
- Yogeswari S, Ramalakshmi S, Neelavathy R and Muthumary J Y. 2012. Identification and comparative studies of different volatile fractions from *Monochaetia kansensis* by GCMS. *Global Journal of Pharmacology* **6**(2): 65–71.
- Zhang X, Li B, Zhang Z, Chen Y and Tian S. 2020. Antagonistic yeasts: A promising alternative to chemical fungicides for controlling postharvest decay of fruit. *Journal of Fungi* 6(3): 158.
- Zhao Q, Shi Y, Ngea G L N, Zhang X, Yang Q, Zhang Q and Zhang H. 2023. Changes of the microbial community in kiwifruit during storage after postharvest application of Wickerhamomyces anomalus. Food Chemistry 404: 134593.