



## Characterization of biogenic volatile organic compounds and membrane lipid peroxidation during leaf ageing in maize (*Zea mays*)\*

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Plants evolved a series of morphological and physiological adaptation mechanism to cope with various biotic and abiotic stresses. The secondary metabolism substance played important roles in these processes. Based on this, McKey (1974) put forward the optimal defense theory (ODT), which assumed that it is valuable for plant to produce these substances to cope with neighbour plants competition, herbivores and pathogen infection (Strauss *et al.* 2002). Recently, the physiological and ecological effects of low molecular weight biogenic volatile organic components (BVOCs) were intensively investigated and the mysterious of BVOCs were unveiled (Vickers *et al.* 2009, Loreto and Schnitzler 2010). For example, the universal green leaf volatiles (GLVs) can induce direct and indirect defense response (Bate and Rothstein 1998, Kishimoto *et al.* 2005). Terpenoid (Terp), another group of universal BVOCs, had antioxidant function (Peñuelas and Llusà 2002, Vickers *et al.* 2009) and could protect leaves against singlet oxygen (Velikova *et al.* 2004).

Given the important role of BVOCs in plant life, the spatial-temporal distribution of these metabolism substances was uneven. Besides the external environment, such as light (Rasulov *et al.* 2009), temperature (Velikova *et al.* 2009), water and nitrogen availability (Gouinguene and Turlings 2002, Penuelas *et al.* 2009; Ormeño *et al.* 2009) had great influence on the biosynthesis and emission rate of BVOCs; the developmental and senescence process also had significant effect on BVOCs biosynthesis and emission (Ucar and Ucar 2010, Niinemets *et al.* 2010), which may the cause of age-dependent resistance, as the un-senescence tissue or organs are more important for the plant based on the ODT (Radhika *et al.* 2008). However, most of these studies were focused on

perennial woody plant. For annual cereal crops, researchers pay more attention to BVOCs ecological function, and crop seedlings were used frequently in their studies. While BVOCs emission pattern during the late growth stage of these cereal crops, which might be really different from seedling stage and play a more important role in their life-cycle, is neglected. Here, we used the most important cereal crop maize (*Zea mays* L.) to explore BVOCs emission pattern during leaf senescence in parallel with monitoring leaf chlorophyll (Chl) degradation and malonyldiadehyde (MDA) accumulation.

Leaves used in this study were harvested in the scientific and education park of Henan Agricultural University in Zhengzhou city (China) in 2010. To create different light environment, high and low plant population densities were selected in parallel. Row-to-row distance was 0.7m for both plant population densities; while plant-to-plant distances were 0.16m and 0.32m for high and low plant population, respectively. The mid part of the second uppermost leaf (Up), ear leaf (Mid) and the seventh leaf (Down) were sampled at tasselling stage (T<sub>1</sub>), 10 days after tasselling (DAT) (T<sub>2</sub>), 20 DAT (T<sub>3</sub>), 30 DAT (T<sub>4</sub>) and 45 DAT (T<sub>5</sub>). Sun irradiance at each sampling canopy level were measured by using SunScan System and photosynthetically active radiation (PAR) distribution character at each canopy level were presented by relative light transmittance in Fig 1.

Leaf volatile were collected by 100 µm PDMS SPME fiber (Supelco) as described by Kollner (2008). Chl and MDA content were determined spectrophotometrically by the method of Lichtenthaler (1987) and Du *et al.* (2011) respectively. Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) was measured according to Alexieva *et al.* (2001). All the quantification was based on three replications. Statistics were done by using SPSS 16.0. The effects of population density, leaf position and leaf age were assessed with ANOVA separately, and the interaction among them was analyzed using a GLM-univariate ( $P=0.05$ ). Post hoc analyses were done by Duncan's multiple-range test at 0.05 probability levels.

\*Short note

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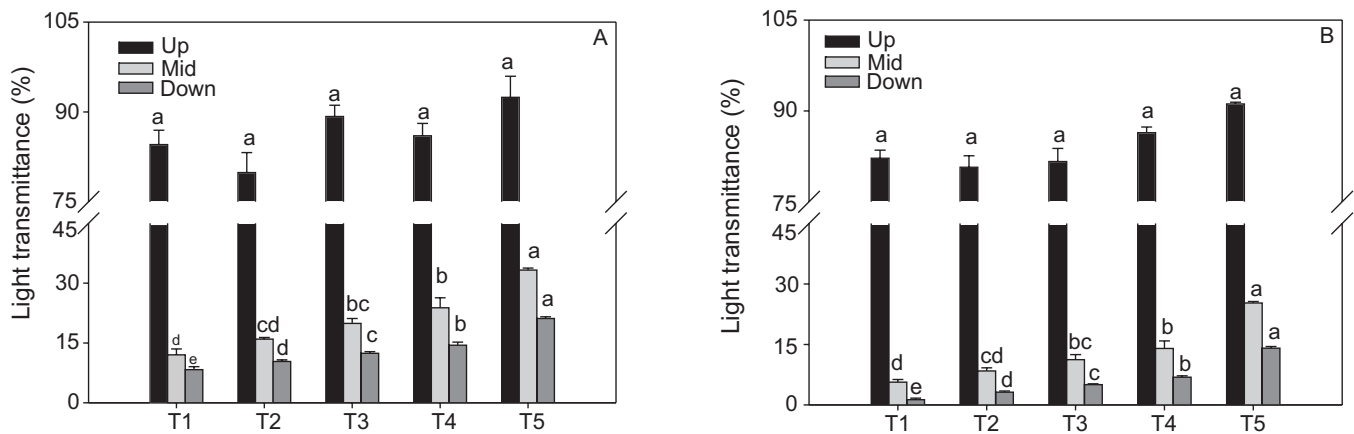


Fig 1 Relative light transmittance of low (A) and high (B) population density at different canopy level. Different letters above the columns indicate statistically different means at  $P < 0.05$

Light transmittance decreased dramatically along the vertical gradient in the canopy. And light transmittance of high population density decreased more than that of low population density. At  $T_1$  stage, light transmittances of low population density and high population density were 12.03% and 5.63% at ear level; and were 8.31% and 1.29% at the seventh leaf level respectively. At  $T_5$  stage, light transmittances of low population density and high population density were 33.26% and 25.27% at ear level and were 21.12% and 14.06% at the seventh leaf level respectively (Fig 1). Leaves at lower canopy level received less PAR than upper level leaves, and at the same canopy level, leaves in low population density received more PAR than high population density.

GLVs and Terp were two major compositions of total BVOCs in the present study. Total GLVs emission rate of the second uppermost leaves and ear leaves in both low and high density populations changed little with leaf aging. There are significant interaction between sampling stage and leaf position ( $P < 0.001$ , Table 1). Total GLVs emission rate of the seventh leaf decreased significantly with leaf aging. GLVs emission rate of the seventh leaf was significantly lower than that of the second uppermost and ear leaves in both low and high density populations at the same sampling time ( $P < 0.01$ , Table 1). This might be partially caused by the reduction of PAR (Gouinguene and Turlings 2002).

Terp emission rate of the second uppermost and ear leaves in both low and high density populations first slightly increased and then decreased with leaf aging. Terp emission rate of the seventh leaf in low density population decreased rapidly with leaf aging, while kept rarely low in a similar emission rate in the high density population (Table 1). Terp emission rate of ear leaves was highest among the three leaf position, and then the second uppermost leaves, the seventh leaves comes the last (Table 1). Terp can protect leaves by scavenging reactive oxygen and reduce pest damage via tritrophic interactions. Ear leaf is closest to the reproductive

organ. By the economic analogy, ear leaf is more important for kernel growth and development at the reproduce stage to keep that species passed on (Bloom *et al.* 1985). Invest in synthetic these volatiles to protect the ear leaf are valuable based on ODH (McKey 1974) with enough substrate and energy for the metabolism.

Variation of Chl *a* and Chl *b* content exhibited a similar character as total GLVs emission rate. Chl *b* decreased more rapidly than Chl *a*, which led a decrease of Chl *a/b*. MDA and  $H_2O_2$  content show an opposite regularity (Table 1). In general, Chl content decreased with leaf aging. On the contrary, the content of MDA and  $H_2O_2$ , which caused by the excess reactive oxygen species and reflect senescence, increased with leaf aging, is in accordance with Hung (1997) and Prochazkova *et al.* (2001). Lipid peroxidation aggravation is accompanied by the decrease of BVOCs emission rate. Besides antioxidant enzyme system, several BVOCs composition also have effect on reactive oxygen species quenching (Affek and Yakir 2002, Velikova *et al.* 2004, Vickers *et al.* 2009). BVOCs emission profile from deferent development stage and the decline of BVOCs emission rate of plants with aging process has been recognized (Zhang *et al.* 2008, Ucar and Ucar 2010, Niinemets *et al.* 2010).

For the low density population, GLVs and Terp emission rate of ear leaves was higher than the second uppermost leaves. However, for the high density population, GLVs and Terp emission rate of ear leaves were a little lower than the second uppermost leaves at the late stage (Table 1). The difference of BVOCs emission patterns of the second uppermost and ear leaves between low and high population density indicate higher emission rate is probably caused by the sufficient PAR. Insufficient PAR led a rapid senescence of lower canopy position leaves (Monneveux *et al.* 2005). As we can see that ear leaves from high density population exhibit relatively low Chl content and high MDA and  $H_2O_2$  content at all sampling stage (Hung and Kao 1997). BVOCs emission rate of leaves at the lower canopy level was

Table 1 BVOCs emission rate, Chl, MDA and H<sub>2</sub>O<sub>2</sub> content at different canopy level with leaf aging

		Low density			High density		
		Up	Mid	Down	Up	Mid	Down
GLVs	T <sub>1</sub>	16.6±0.7a	20.0±0.2a	16.1±0.7a	15.9±0.6b	18.9±0.6a	14.3±0.9a
	T <sub>2</sub>	19.1±0.8a	19.0±0.3a	14.2±2.3ab	19.5±1.4a	19.6±0.6a	13.1±1.1ab
	T <sub>3</sub>	19.9±0.6a	20.2±1.1a	12.8±1.6abc	19.8±0.4a	19.9±0.5a	10.7±1.3ab
	T <sub>4</sub>	20.7±1.0a	21.9±1.1a	10.7±1.5bc	20.5±0.5a	19.8±1.5a	8.9±1.3b
	T <sub>5</sub>	17.8±1.9a	18.2±0.7a	9.5±0.6c	18.1±1.1ab	17.1±0.6a	
Terp	T <sub>1</sub>	38.4±2.8c	72.9±6.7ab	37.1±2.9a	36.9±4.6c	43.7±2.0c	28.8±5.9a
	T <sub>2</sub>	68.3±5.2ab	85.6±13.4a	30.8±4.2a	57.2±2.0b	55.7±3.6ab	26.1±3.2a
	T <sub>3</sub>	82.2±3.9a	75.3±1.3ab	27.2±2.2ab	78.5±0.7a	61.0±3.4a	16.9±5.0a
	T <sub>4</sub>	65.2±10.1ab	69.1±6.6ab	17.5±0.2bc	55.9±4.8b	47.6±1.5bc	9.2±3.2a
	T <sub>5</sub>	58.7±4.4b	58.0±6.1b	12.5±6.2c	53.5±4.7b	39.3±3.3c	
Chl <i>a</i> (mg/g FW)	T <sub>1</sub>	2.3±0.0b	3.4±0.1a	2.3±0.1a	2.2±0.1b	3.2±0.0a	1.7±0.0a
	T <sub>2</sub>	3.1±0.4a	3.4±0.0a	2.0±0.0b	2.9±0.1a	2.8±0.0b	1.4±0.0b
	T <sub>3</sub>	3.2±0.0a	3.1±0.4ab	1.8±0.1b	2.8±0.2a	2.7±0.0c	1.2±0.0c
	T <sub>4</sub>	2.9±0.2a	2.9±0.1b	1.6±0.2c	2.4±0.0b	2.5±0.0d	1.1±0.0d
	T <sub>5</sub>	2.0±0.2b	2.3±0.1c	1.2±0.0d	1.7±0.0c	1.9±0.1e	
Ch <i>b</i> (mg/g FW)	T <sub>1</sub>	0.7±0.0c	1.1±0.1a	0.8±0.0a	0.6±0.0c	0.9±0.0a	0.6±0.0a
	T <sub>2</sub>	1.0±0.1a	1.1±0.1a	0.7±0.0b	0.9±0.0ab	0.9±0.0b	0.6±0.0a
	T <sub>3</sub>	1.0±0.0a	1.0±0.1ab	0.6±0.0c	0.9±0.1a	0.8±0.0c	0.5±0.0b
	T <sub>4</sub>	0.9±0.0b	1.0±0.0b	0.6±0.0c	0.8±0.0b	0.8±0.0c	0.5±0.0b
	T <sub>5</sub>	0.7±0.0c	0.8±0.0c	0.5±0.0d	0.6±0.0c	0.7±0.0d	
Chl <i>a/b</i>	T <sub>1</sub>	3.4±0.0a	3.1±0.3a	2.9±0.1a	3.5±0.2a	3.5±0.0a	2.8±0.2a
	T <sub>2</sub>	3.2±0.3a	3.1±0.2a	2.9±0.1a	3.3±0.1ab	3.2±0.0b	2.5±0.0bc
	T <sub>3</sub>	3.2±0.1a	3.1±0.2a	3.0±0.2a	3.1±0.1b	3.2±0.1b	2.6±0.1ab
	T <sub>4</sub>	3.2±0.2a	3.0±0.1a	2.7±0.3a	2.9±0.1c	3.0±0.0c	2.4±0.1bc
	T <sub>5</sub>	3.1±0.3a	2.9±0.1a	2.6±0.1a	2.7±0.0d	2.8±0.1d	
MDA (imol/g FW)	T <sub>1</sub>	24.5±2.1c	36.5±3.4d	37.0±2.8d	23.0±3.9d	38.2±2.8d	40.6±1.5c
	T <sub>2</sub>	31.7±3.8b	39.2±2.6cd	41.7±3.5cd	34.6±1.5c	39.3±3.1cd	42.5±2.4bc
	T <sub>3</sub>	39.0±1.7a	42.0±3.8bc	43.9±1.9bc	42.5±2.3b	45.0±4.6bc	47.3±4.4ab
	T <sub>4</sub>	41.5±1.4a	45.4±2.2ab	47.0±1.6b	47.4±4.9b	49.7±3.0ab	53.0±2.7a
	T <sub>5</sub>	42.4±2.1a	48.2±0.5a	53.4±3.0a	53.7±2.4a	53.1±1.6a	
H <sub>2</sub> O <sub>2</sub> (imol/g FW)	T <sub>1</sub>	146.3±15.4d	196.5±11.2c	285.7±16.7c	164.7±4.9d	274.6±25.4c	332.4±21.8c
	T <sub>2</sub>	236.2±7.1c	304.8±15.5b	336.8±12.1b	369.1±11.4c	389.6±15.9b	412.0±31.6b
	T <sub>3</sub>	328.9±30.1b	318.6±24.9b	360.5±21.4b	411.8±28.2b	419.3±10.1b	451.0±16.2ab
	T <sub>4</sub>	356.6±18.0b	365.3±22.0a	422.0±46.0a	425.0±24.7b	464.4±24.1a	472.0±35.2a
	T <sub>5</sub>	395.1±17.8a	391.0±31.5a	437.1±24.5a	501.1±30.8a	474.3±26.7a	

Different letters within the same row indicate mean values statistically different at  $P < 0.05$

significantly lower than the mid and up leaves also provide evidence for this. This is in agreement with the prior results that some BVOCs emission is light dependent (Schuh *et al.* 1997, Gouinguene and Turlings 2002, Rasulov *et al.* 2009).

Our result also indicates that leaf aging also remarkably affects BVOCs emission pattern. On one hand, juvenile leaves metabolism are sharper, which can provide sufficient substrate for BVOCs synthesis; on the other hand, juvenile leaves is the growth center, more assimilates redistribution

or translocation to this center organ (Bloom *et al.* 1985, Radhika *et al.* 2008).

### SUMMARY

A study was conducted to access BVOCs emission pattern during leaf senescence in parallel with monitoring leaf Chl degradation and MDA accumulation. BVOCs emission rate and Chl content of leaves in the lower canopy position were significantly lower than leaves in the mid and up positions. While MDA and H<sub>2</sub>O<sub>2</sub> content of the lower position leaves were significantly higher than the up and mid position leaves. Positive correlation between leaf volatile emission rate and chlorophyll content and negative correlation between leaf volatile emission rate and MDA and H<sub>2</sub>O<sub>2</sub> content were observed. VOCs emission pattern during the cereal crop aging could provide more evidence for age-related resistance though the results here we show are *in vitro*.

### REFERENCES

- Affek H P and Yakir D. 2002. Protection by isoprene against singlet oxygen in leaves. *Plant Physiology* **129**: 269–77.
- Alexieva V, Sergiev I, Mapelli S and Karanov E. 2001. The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant Cell & Environment* **24**: 1337–44.
- Bate N J and Rothstein S J. 1998. C<sub>6</sub>-volatiles derived from the lipoxygenase pathway induce a subset of defense-related genes. *Plant Journal* **16**: 561–9.
- Bloom A J, Chapin F S and Mooney H A. 1985. Resource limitation in plants. an economic analogy. *Annual Review of Ecological System* **16**: 363–92.
- Du J B, Yuan S, Chen Y E, Sun X, Zhang Z W, Xu F, Yuan M, Shang J and Lin H H. 2011. Comparative expression analysis of dehydrins between two barley varieties, wild barley and Tibetan hulless barley associated with different stress resistance. *Acta Physiology Plantarum* **33**: 567–74.
- Farang M A and Pare P W. 2002. C-6-green leaf volatiles trigger local and systemic VOC emissions in tomato. *Phytochemistry* **61**: 545–54.
- Gouinguene S P and Turlings T C J. 2002. The effects of abiotic factors on induced volatile emissions in corn plants. *Plant Physiology* **129**: 1296–307.
- Hung K T and Kao C H. 1997. Lipid peroxidation in relation to senescence of maize leaves. *Journal of Plant Physiology* **150**: 283–6.
- Kishimoto K, Matsui K, Ozawa R and Takabayashi J. 2005. Volatile C<sub>6</sub>-aldehydes and Allo-ocimene activate defense genes and induce resistance against *Botrytis cinerea* in *Arabidopsis thaliana*. *Plant Cell Physiology* **46**: 1093–102.
- Kollner T G, Held M, Lenk C, Hiltbold I, Turlings T C J, Gershenzon J and Degenhardt J. 2008. A maize (E)-beta-caryophyllene synthase implicated in indirect defense responses against herbivores is not expressed in most American maize varieties. *The Plant Cell* **20**: 482–94.
- Lichtenthaler H K. 1987. Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. *Methods Enzymology* **148**: 350–82.
- Loreto F and Schnitzler J P. 2010. Abiotic stresses and induced BVOCs. *Trends in Plant Science* **15**: 154–66.
- McKey D. 1974. Adaptive patterns in alkaloid physiology. *American Naturalist* **108**: 305–20.
- Monneveux P, Zaidi P H and Sanchez C. 2005. Population density and low nitrogen affects yield-associated traits in tropical maize. *Crop Science* **45**: 535–45.
- Niinemets Ü, Arneeth A, Kuhn U, Monson R K, Peñuelas J and Staudt M. 2010. The emission factor of volatile isoprenoids: stress, acclimation, and developmental responses. *Biogeosciences* **7**: 2203–23.
- Ormeño E, Olivier R, Mévy J P, Baldy V and Fernandez C. 2009. Compost may affect volatile and semi-volatile plant emissions through nitrogen supply and Chlorophyll fluorescence. *Chemosphere* **77**: 94–104.
- Peñuelas J and Llusà J. 2002. Linking photorespiration, monoterpenes and thermotolerance in *Quercus*. *New Phytologist* **155**: 227–37.
- Peñuelas J, Filella I, Seco R and Llusia J. 2009. Increase in isoprene and monoterpene emissions after re-watering of droughted *Quercus ilex* seedlings. *Biology Plants* **53**: 351–4.
- Prochazkova D, Sairam R K, Srivastava G C and Singh D V. 2001. Oxidative stress and antioxidant activity as the basis of senescence in maize leaves. *Plant Science* **161**: 765–71.
- Radhika V, Kost C, Bartram S, Heil M and Boland W. 2008. Testing the optimal defence hypothesis for two indirect defences: extrafloral nectar and volatile organic compounds. *Planta* **228**: 449–57.
- Rasulov B, Huve K, Valbe M, Laisk A and Niinemets U. 2009. Evidence that light, carbon dioxide, and oxygen dependencies of leaf isoprene emission are driven by energy status in hybrid aspen. *Plant Physiology* **151**: 448–60.
- Schuh G, Heiden A C, Hoffmann T, Kahl J, Rockel P, Rudolph J and Wildt J. 1997. Emissions of volatile organic compounds from sunflower and beech: dependence on temperature and light intensity. *Journal of Atmosphere Chemistry* **27**: 291–318.
- Strauss S Y, Rudgers J A, Lau J A and Irwin R E. 2002. Direct and ecological costs of resistance to herbivory. *Trends in Ecological Evolution* **17**: 278–85.
- Ucar G and Ucar M B. 2010. Natural decline of volatile acids due to aging of living trees. *Wood Science Technology* **44**: 465–74.
- Velikova V, Edreva A, Loreto F. 2004. Endogenous isoprene protects *Phragmites australis* leaves against singlet oxygen. *Physiologia Plantarum* **122**: 219–25.
- Velikova V, Tsonev T, Barta C, Centritto M, Koleva D, Stefanova M, Busheva M and Loreto F. 2009. BVOC emissions, photosynthetic characteristics and changes in Chloroplast ultrastructure of *Platanus orientalis* L. exposed to elevated CO<sub>2</sub> and high temperature. *Environmental Pollution* **157**: 2629–37.
- Vickers C E, Gershenzon J, Lerdau M T and Loreto F. 2009. A unified mechanism of action for volatile isoprenoids in plant abiotic stress. *Nature Chemical Biology* **5**: 283–91.
- Zhang P Y, Chen K S, He P Q, Liu S H and Jiang W F. 2008. Effects of crop development on the emission of volatiles in leaves of *Lycopersicon esculentum* and its inhibitory activity to *Botrytis cinerea* and *Fusarium oxysporum*. *Journal of Integrative Plant Biology* **50**: 84–91.