



Effect of trap colour, installation height and size on insect capture in grapevine (*Vitis vinifera*) ecosystem

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ABSTRACT

Effects of optimum colour, installation height and size of sticky traps were studied for monitoring insects in grapes. *Aphis* spp., *Amrasca biguttula biguttula* Ishida, *Rhipiphorothrips cruentatus* Hood, *Scirtothrips dorsalis* Hood and *Stethorus rani* Kapur were the major insect species trapped. Yellow was most effective colour for trapping all these five insect species as compared to blue. Optimum installation height was at near-ground for *Aphis* spp. and *A. biguttula biguttula* and above-canopy for *S. rani*. Optimum size of traps was found to be 11×30 square cm over 22×30 square cm sizes. When analysed for interaction effects, yellow sticky traps of 11×30 square cm installed at near-ground level provided most effective trapping for *A. biguttula biguttula* (1.47, 2.46 and 1.22 and 1.14, 2.12 and 0.95 catches per cell during 2011-12, 2012 and 2012-13 in bower and Y, respectively) and *Aphis* spp. (5.92, 0.37 and 1.42 and 4.31, 0.37 and 1.11 catches per cell during 2011-12, 2012 and 2012-13 in bower and Y, respectively). Yellow sticky traps (11×30 square cm size) installed at above-canopy were most effective in trapping *R. cruentatus* (0.03, 0.86 and 0.47 catches per cell during 2011-12, 2012 and 2012-13) in Y system, however at near-ground in bower system (0.92 and 0.727 catches per cell during 2012 and 2012-13). Yellow sticky traps installed at above-canopy were most effective in trapping *S. rani* in both bower (0.027 and 0.010 catches per cell during 2011-12 and 2012-13) and Y system (0.058 and 0.007 catches per cell during 2011-12 and 2012-13).

Key words: Colour, Grapes, Insect monitoring, Sticky trap

Grape (*Vitis vinifera* L.) has aroused considerable interest in recent years about its cultivation in India due to its prolific yield and good monetary returns (Mani *et al.* 2014). It is also important from export point of view as it has great demand in the foreign markets such as United Kingdom, Saudi Arabia and United Arab Emirates fetching valuable foreign exchange (Mani *et al.* 2014). The major part of the grape area is in peninsular states of India where winters are mild and the grapevines do not undergo dormancy and remain evergreen throughout the year. Therefore, the grapevines remain available for the pest development all the year round which leads to high pest infestations (Yadav *et al.* 2012). Among arthropod pests; thrips, *Scirtothrips dorsalis* Hood and *Rhipiphorothrips cruentatus* Hood; mealybugs, *Maconellicoccus hirsutus* (Green) and red spider mites, *Tetranychus* spp. are major pests in peninsular India. Leafhoppers were earlier considered minor pests in grapes,

however, outbreak of *Amrasca biguttula biguttula* Ishida in grapes in state of Maharashtra was observed during October-November, 2011 (Sharma *et al.* 2013). Since then, *A. biguttula biguttula* remained as one of the major pests in viticulture.

Monitoring is a basic tool for decision making in pest management. The first step in managing pests is to determine the practically implementable, accurate and effective monitoring system to measure their population levels. The sticky traps are very efficient and important monitoring tools alerting growers to pests before damage is observed in crops (Bethke *et al.* 2009). The sticky traps are also very useful when other trapping methods such as pheromone or light traps are either unavailable or economically or practically not feasible.

Trap height (Atakanand Canhilal 2004, Pilkington *et al.* 2004) and colour (Clare *et al.* 2000, Roubos and Liburd 2008) can also influence the catch of insects in sticky traps. Blue traps captured more *Frankliniella occidentalis* than yellow traps in plum (Allsopp 2010), broccoli (Chen *et al.* 2004) and lettuce and onion (Natwick *et al.* 2007). Demirel and Yildirim (2008) found yellow sticky traps effective in trapping leafhoppers. Among beneficial insects, blue attracted more *Allograptus bliqua* (S.) than yellow or white traps in broccoli (Chen *et al.* 2004). However, *Chrysoperla carnea* (S.) in alfalfa showed a variable seasonal response

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to orange, red, black hues and clear traps (Blackmer *et al.* 2008). Esker *et al.* (2004) evaluated effect of installation heights (0.15, 0.30, 0.45, 0.60, and 0.90m) on sticky trap insect catch in corn field.

In grapes, yellow sticky traps were recommended by Scatoni *et al.* (2007) against *F. occidentalis* and Akbarzadeh Shoukat and Shayesteh (2006) against *Rubiothrips vitis*. Blue traps were suggested by Tsitsipis *et al.* (2003), Allsopp (2010) and Lucas and Ceron (2011) against *F. occidentalis*. Mani *et al.* (2008) recommended installation of yellow or blue sticky traps for monitoring thrips. Sufficient information is not available on optimum colour, size and height of sticky traps for installation in vineyard ecosystems for monitoring various insect species in peninsular India. Therefore, the present study was carried out to find out optimum installation height, colour and size of sticky traps for insect catch to develop insect monitoring method using sticky traps in grapevine ecosystem.

Further, two training systems, viz. bower and extended Y, are commonly followed in Indian viticulture. In the bower system the vines are trained at the height of about 6.5 feet with horizontal canopy. However, in case of the extended Y system, the cordons are trained horizontally with vertical shoot positioning which helps in harvesting of maximum sunlight required for effective fruit bud differentiation. As these different training systems affect the canopy architecture, they may also affect the optimum height of the traps. Thus, the study was carried out in both of these training systems separately.

MATERIALS AND METHODS

Commercially available sticky traps of yellow and blue colours were used. To define colours of sticky traps RGB (red, green, blue) values of the coloured traps were measured by method described by Rodriguez-Saona *et al.* (2012). Digital images of the coloured sticky traps were taken with a Sony Cybershot DSC-TX9 digital camera at 4000 × 2248 pixel resolution and 24-bit colour during 1130 to 1230 hours from the vineyard. Internet software described by Byers (2006) was used to analyse the RGB attributes of pixels.

The experiments were conducted in table grape, *Vitis vinifera* on Thompson Seedless variety on Dog Ridge rootstock for two years from 2011-13 in 12-14 years old vineyards with bower and Y training systems. The location of the vineyards was at ICAR-National Research Centre for Grapes, Pune, India.

Effect of colour, size and height of sticky traps was assessed during two fruiting seasons during 2011-12 and 2012-13 and one vegetative season during 2012 for catch of insects. A 2×3×2 levels for colour (main factor) × height (sub factor) × size (sub-sub factor) factorial design with five replicates was used in both the training systems separately. Traps set at different heights were placed in plant row and separated by a distance of at least 15 m for the same colour. The traps for different sizes were placed in a single plot for colour with minimum distance of one meter. To

create height variations, traps for a given colour and size were tied to a thin bamboo stick at different positions, viz., above canopy level, inside canopy and near ground (30 cm above ground surface) with the help of plastic threads. The positions, at which the traps were set, were determined relative to the ground surface for near ground level; and to the height of the canopy of the vines for inside canopy and above canopy levels.

The traps were replaced from 1000 to 1200 hours at every 10 day intervals. The traps were removed and brought to the laboratory for visual observations on both of the adhesive sides of the traps. The visual observations were taken on total number of thrips and leafhoppers per trap under stereo-zoom-microscope. We also counted the number of other insects trapped such as aphids, coccinellid predators, parasitic wasps, etc. Insect predators were identified to the order level and, when possible, to family and species. The sticky substance was dissolved with acetone to remove the insects for identification whenever necessary.

To bring uniformity in insect counts per unit area in traps of 11×30 cm² size with 22×30 cm² size, the insect numbers for equivalent unit area of 2.50 × 2.50 cm² were calculated for data analysis and this area was considered as one cell. A three-factor analysis of variance (2×3×2 for colour × height × size) was performed on the data to determine the optimum colour, height and size of sticky traps and the best interaction. When significant, the means were separated by least significant difference. All the statistical analyses were carried out using statistical software SAS Institute 9.3 (SAS Institute 2012).

RESULTS AND DISCUSSION

Aphid, *Aphis* spp.; leafhopper, *Amrasca biguttula biguttula*; chilli thrips, *Scirtothrips dorsalis*; grapevine thrips, *Rhipiphorotheirus cruentatus* Hood and coccinellid beetle, *Stethorus rani* were the major insect species trapped in decreasing order during fruiting season 2011-12. During 2012 vegetative season, it was *A. biguttula biguttula*, *R. cruentatus*, *Aphis* spp. and *S. dorsalis* and *S. rani* in decreasing order. During 2012-13 fruiting season, the decreasing order of catch of insects was *Aphis* spp, *A. biguttula biguttula*, *R. cruentatus*, *S. dorsalis* and *S. rani*. However, higher numbers of *S. dorsalis* during 2011-12 fruiting season can be attributed to the fact that the population of *R. cruentatus* was found lower during the season (NRCG 2012). Therefore, the conclusion that which insect species showed more attraction towards sticky traps cannot be drawn on the basis of present data. The differences in number of different insect species per cell only indicate their population levels in the vineyards.

Individual effects of colour, height and size of sticky traps

When analysed separately, colour, height and size significantly affected capture of insects. In all three seasons, yellow was the most effective trap in descending order to *Aphis* spp, *A. biguttula biguttula*, *R. cruentatus*, *S. dorsalis* and *S. rani* as compared to blue traps in both bower and

and 2012 in bower and Y system, *A. biguttula biguttula* during 2012 in bower, *S. dorsalis* during 2012-13 in Y system. Therefore, the 22×30 cm² size traps can be cut into half to make smaller size of 11×30 cm² can be used to cover larger area for monitoring purpose thus providing cost-effectiveness.

Interaction effects of colour, height and size of sticky traps

During fruiting season 2011-12 in bower, yellow+near ground+11×30 cm² interaction was most effective in trapping *A. biguttula biguttula* and *S. dorsalis* and yellow+near ground+22×30 cm² interaction was most effective for *Aphis* spp. (Table 2). Yellow+above canopy+11×30 cm² interaction was most effective for *R. cruentatus* and *S. rani*. This interaction was at par with yellow+above canopy+22×30 cm² for *S. rani*. In Y training system during the same season, yellow+near ground+11×30 cm² interaction was most effective in trapping *A. biguttula biguttula*, *S. dorsalis* and *Aphis* spp. Yellow+above canopy+11×30 cm² interaction was most effective for *R. cruentatus* and *S. rani*. This interaction was at par with yellow+above canopy+22×30 cm² for *S. rani*.

During vegetative season 2012, yellow+near ground+11×30 cm² and yellow+near ground+22×30 cm² interactions were most effective in trapping *A. biguttula biguttula* and *Aphis* spp. in both bower and Y systems (Table 3). Yellow+above canopy+11×30 cm² interaction was most effective for *S. dorsalis* in both training systems. This interaction was at par with yellow+inside canopy+11×30 cm² for bower. Yellow+near ground+11×30 cm² interaction was most effective for *R. cruentatus* in bower. However, yellow+above canopy+11×30 cm² and yellow+inside canopy+11×30 cm² interactions were most effective and

at par with each other in Y system.

During fruiting season 2012-13, yellow+near ground+11×30 cm² interaction was most effective in trapping *A. biguttula biguttula* and *Aphis* spp. in both training systems (Table 4). Yellow+above canopy+11×30 cm² interaction was most effective for *S. dorsalis* in Y system and *R. cruentatus* in bower. Yellow+near ground+11×30 cm² interaction was most effective for *S. dorsalis* in bower and *R. cruentatus* in Y system. Yellow+above canopy+11×30 and 22×30 cm² and yellow+inside canopy+11×30 and 22×30 cm² were most effective and at par with each other in trapping *S. rani* in Y system. However, yellow+above canopy+11×30 cm² interaction was most effective for *S. rani* in bower system.

Yellow+near ground+11×30 cm² was most effective interaction for *A. biguttula biguttula* and *Aphis* spp. in both training systems. *R. cruentatus* and *S. rani* preferred yellow+above canopy+11×30 cm². *S. dorsalis* showed equal preference towards yellow+nearground+11×30 cm² and yellow+above canopy+11×30 cm².

Single-lens eyes are able to focus on objects at different distances. However, insect compound eyes have the same angular resolution at far and close distances, therefore, the insects are unable to resolve spatial details of distant objects, though they can use vision at extremely close distances (Hempel de Ibarra *et al.* 2015). It was also emphasized that insects could not distinguish small objects or small-scaled variations of shapes and patterns over long distances, e.g. swallowtail butterfly, *Papilio xuthus* (Takeuchi *et al.* 2006). In the present experiment, the smaller sticky trap size of 11×30 cm² was physiological sufficient for *A. biguttula biguttula* and *Aphis* spp. to detect for yellow+near ground+11×30 cm² interaction, and mostly for *S. dorsalis*, *R. cruentatus* and *S. rani* to detect for yellow+above

Table 2 Mean number of insect catches during fruiting season 2011-12

Trap color	Trap height	Trap size (cm ²)	Mean number of insect catches per cell									
			<i>A. biguttula</i>		<i>S. dorsalis</i>		<i>R. cruentatus</i>		<i>Aphis</i> spp.		<i>S. rani</i>	
			Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y
Yellow	Above canopy	22×30	0.51 c	0.264 cd	0.143 c	0.071 c	0.077 b	0.025 bc	1.58 c	0.74 d	0.025 a	0.031 ab
		11×30	0.40 de	0.298 c	0.145 c	0.108 b	0.105 a	0.036 a	1.53 c	0.67 d	0.027 a	0.058 a
	Inside canopy	22×30	0.39 e	0.235 d	0.083 e	0.055 d	0.044 c	0.011 e	1.39 d	0.73 d	0.008 cd	0.012 b
		11×30	0.46 cd	0.226 d	0.108 d	0.078 c	0.045 c	0.029 b	1.24 e	0.82 cd	0.014 b	0.023 b
	Near Ground	22×30	1.31 b	0.829 b	0.168 b	0.116 b	0.029 cde	0.011 e	6.55 a	4.01 b	0.005 cd	0.013 b
		11×30	1.47 a	1.141 a	0.199 a	0.130 a	0.041 cd	0.018 d	5.92 b	4.31 a	0.009 bcd	0.019 b
Blue	Above canopy	22×30	0.06 fg	0.078 e	0.036 fg	0.034 f	0.003 g	0.005 ef	0.26 g	0.81 cd	0.008 cd	0.007 b
		11×30	0.09 fg	0.097 e	0.033 fg	0.052 d	0.020 efg	0.022 cd	0.26 g	0.81 cd	0.010 bcd	0.018 b
	Inside canopy	22×30	0.06 g	0.061 e	0.029 g	0.037 ef	0.011 fg	0.004 f	0.22 g	0.66 d	0.005 cd	0.003 b
		11×30	0.09 fg	0.074 e	0.045 f	0.047 de	0.025 def	0.007 ef	0.30 g	0.73 d	0.011 bc	0.011 b
	Near Ground	22×30	0.08 fg	0.078 e	0.038 fg	0.039 ef	0.019 efg	0.006 ef	0.44 f	0.75 cd	0.004 d	0.003 b
		11×30	0.11 f	0.098 e	0.072 e	0.050 d	0.012 efg	0.002 f	0.44 f	0.93 c	0.009 bcd	0.010 b
		F-value	439.99	203.71	123.97	70.12	18.44	17.93	1631.15	311.35	8.82	1.82
		df (model, error)	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44
		Pr>F	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	0.0618

Table 3 Mean number of insect catches during vegetative season 2012

Trap color	Trap height	Trap size (cm ²)	Mean number of insect catches per cell									
			<i>A. biguttula</i>		<i>S. dorsalis</i>		<i>R. cruentatus</i>		<i>Aphis</i> spp.		<i>S. rani</i>	
			Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y
Yellow	Above canopy	22×30	1.796 b	1.55 b	0.122 ab	0.12 b	0.56 d	0.626 c	0.264 b	0.32 b	0	0.000 b
		11×30	1.499 c	1.93 a	0.127 a	0.14 a	0.71 c	0.864 a	0.255 b	0.33 b	0	0.000 b
	Inside canopy	22×30	0.910 d	0.29 c	0.100 c	0.10 c	0.66 c	0.601 c	0.202 c	0.24 c	0	0.000 b
		11×30	1.381 c	0.34 c	0.126 a	0.13 b	0.85 b	0.866 a	0.237 b	0.25 c	0	0.000 b
	Near Ground	22×30	2.416 a	1.90 a	0.104 c	0.08 d	0.65 c	0.767 b	0.391 a	0.39 a	0	0.000 b
		11×30	2.468 a	2.12 a	0.112 bc	0.06 e	0.92 a	0.790 b	0.379 a	0.37 a	0	0.000 b
Blue	Above canopy	22×30	0.386 e	0.14 c	0.027 de	0.03 gh	0.03 e	0.039 d	0.090 de	0.09 e	0	0.000 b
		11×30	0.315 e	0.14 c	0.038 d	0.05 f	0.05 e	0.051 d	0.103 d	0.10 de	0	0.000 b
	Inside canopy	22×30	0.191 e	0.08 c	0.023 e	0.02 h	0.03 e	0.035 d	0.073 e	0.08 e	0	0.000 b
		11×30	0.195 e	0.10 c	0.034 de	0.04 g	0.04 e	0.063 d	0.090 de	0.11d	0	0.000 b
	Near Ground	22×30	0.188 e	0.17 c	0.022 e	0.02 h	0.03 e	0.028 d	0.081 de	0.09 e	0	0.000 b
		11×30	0.182 e	0.15 c	0.033 de	0.03 gh	0.04 e	0.050 d	0.104 d	0.09 de	0	0.004 a
	F-value		104.61	60.35	77.73	83.51	212.67	184.45	105.85	133.32		31.25
	df (model, error)		15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44
	Pr>F		<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001

Table 4 Mean number of insect catches during fruiting season 2012-13

Trap color	Trap height	Trap size (cm ²)	Mean number of insect catches per cell									
			<i>A. biguttula</i>		<i>S. dorsalis</i>		<i>R. cruentatus</i>		<i>A. craccivora</i>		<i>S. rani</i>	
			Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y
Yellow	Above canopy	22×30	0.477 e	0.366 e	0.139 c	0.102 c	0.282 e	0.355 c	0.61 de	0.62 d	0.0066 bc	0.0058 abcde
		11×30	0.551 d	0.436 d	0.193 a	0.128 b	0.426 c	0.474 a	0.68 c	0.75 c	0.0104 a	0.0078 a
	Inside canopy	22×30	0.484 e	0.401 de	0.106 e	0.071 e	0.386 d	0.342 c	0.54 f	0.41 f	0.0072 bc	0.0066 abcd
		11×30	0.628 c	0.562 c	0.158 b	0.091 cd	0.533 b	0.425 b	0.64 cd	0.55 e	0.0080 b	0.0076 ab
	Near Ground	22×30	0.891 b	0.800 b	0.087 f	0.075 de	0.559 b	0.347 c	1.12 b	0.95 b	0.0042 de	0.0047 cde
		11×30	1.228 a	0.955 a	0.122 d	0.327 a	0.727 a	0.424 b	1.42 a	1.11 a	0.0081 b	0.0051 bcde
Blue	Above canopy	22×30	0.037 f	0.048 f	0.020 gh	0.025 fgh	0.020 f	0.016 d	0.28 hi	0.33 g	0.0033 e	0.0037 ef
		11×30	0.057 f	0.045 f	0.027 g	0.029 fg	0.033 f	0.022 d	0.34 h	0.43 f	0.0059 cd	0.0072 abc
	Inside canopy	22×30	0.042 f	0.042 f	0.009 h	0.010 h	0.020 f	0.018 d	0.23 i	0.23 h	0.0032 e	0.0042 def
		11×30	0.053 f	0.059 f	0.016 gh	0.015 gh	0.027 f	0.030 d	0.29 h	0.35 g	0.0038 e	0.0076 ab
	Near Ground	22×30	0.062 f	0.062 f	0.012 h	0.011 gh	0.025 f	0.029 d	0.45 g	0.42 f	0.0026 e	0.0020 f
		11×30	0.061 f	0.064 f	0.027 g	0.034 f	0.032 f	0.036 d	0.57 ef	0.54 e	0.0081 b	0.0053 abcde
	F-value		632.16	416.69	195.16	138.51	336.37	557.90	211.13	258.92	11.20	3.06
	df (model, error)		15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44	15,44
	Pr>F		<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	0.0020

canopy+11x30 cm² interaction. Higher capture of *A. biguttula biguttula* on sticky traps at near ground height in both bower and Y training systems present indications of movement away from canopy towards ground during day time possibly due to higher temperatures which needs further investigations.

The analyses of RGB will allow comparisons of the trap colours with data during future studies which may utilize similar measurements (Table 5). Further, the batches of plastic traps from commercial sources can also be checked

in future for consistency with earlier used traps so that the attractive properties can be standardized and reliable (Rodriguez-Saona *et al.* 2012).

On the basis of these observations, it can be concluded that the sticky traps can be used in grapevine ecosystems for the monitoring of *Aphis* spp., *A. biguttula biguttula*, *R. cruentatus*, *S. dorsalis* and *S. rani* when installed using most effective colour, at right installation height and optimum size. The future area of research calls for standardization of sticky trap based economic threshold levels (ETL) for

Table 5 Mean (\pm standard deviation) red (R), green (G), blue (B), trichromatic percentages (% R, G, B) and H:S:L values (Hue: Saturation: Luminosity) from areas of digital photos of sticky traps taken under vineyard conditions in Bower and Y training system

Trap colour	Trap level	R \pm SD		R% ^a		G \pm SD		G%		B \pm SD		B%		H:S:L	
		Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y	Bower	Y
Yellow	Near ground	153 \pm 4	165 \pm 4	50.00	49.69	141 \pm 4	148 \pm 4	46.07	44.57	12 \pm 3 \pm 4	19 \pm 4	3.92	5.72	0.15:0.85:032	0.15:0.79:0.36
	Inside canopy	164 \pm 3	158 \pm 3	48.95	49.68	157 \pm 4	141 \pm 3	46.86	44.33	14 \pm 5 \pm 4	19 \pm 4	4.17	5.97	0.15:0.84:034	0.15:0.78:0.35
	Above canopy	164 \pm 3	164 \pm 3	46.72	48.95	152 \pm 3	147 \pm 3	43.30	43.88	35 \pm 9 \pm 4	24 \pm 4	9.97	7.16	0.15:0.64:039	0.15:0.74:0.37
Blue	Near ground	19 \pm 6	17 \pm 3	4.65	4.27	148 \pm 8	145 \pm 2	36.27	36.43	241 \pm 7	236 \pm 2	59.06	59.29	0.90:0.77:0.51	0.90:0.86:0.50
	Inside canopy	13 \pm 4	20 \pm 3	3.77	5.18	121 \pm 6	138 \pm 3	35.17	35.75	210 \pm 5	228 \pm 3	61.04	59.06	0.91:0.88:0.44	0.90:0.84:0.49
	Above canopy	15 \pm 4	18 \pm 3	4.34	4.81	121 \pm 7	134 \pm 3	35.07	35.82	209 \pm 8	222 \pm 2	60.57	59.35	0.91:0.87:0.44	0.90:0.85:0.47

^aRepresents the percent red $\{R/(R + G + B) \times 100\}$, similarly %G and %B was also determined.

decision making in grapes.

REFERENCES

- Akbarzadeh Shoukat G H A and Shayesteh N. 2006. Thrips species found in West Azarbaijan (Orumieh) vineyards and seasonal abundance of the predominant species (*Rubiothrips vitis*). *Journal of Agricultural Science and Technology* **8(2)**: 133–9.
- Allsopp E. 2010. Investigation into the apparent failure of chemical control for management of western flower thrips, *Frankliniella occidentalis* (Pergande), on plums in the Western Cape Province of South Africa. *Crop Protection* **29(8)**: 824–31.
- Amala U and Yadav D S. 2013. Study on life table parameters and predatory potential of *Stethorus rani* Kapur on red spider mite, *Tetranychus urticae* Koch. *Biopesticides International* **9(2)**: 113–9.
- Atakan E and Canhilal R. 2004. Evaluation of yellow sticky traps at various heights for monitoring cotton insect pests. *Journal of Agricultural and Urban Entomology* **21(1)**: 15–24.
- Bethke J A. 2009. Monitoring with Sticky Traps. UC IPM Pest Management Guidelines: Floriculture and Ornamental Nurseries. UC ANR Publication 3392.
- Blackmer J L, Byers J A and Rodriguez-Saona C. 2008. Evaluation of color traps for monitoring *Lygus* spp.: Design, placement, height, time of day, and non-target effects. *Crop Protection* **27(2)**: 171–81.
- Byers J A. 2006. Analysis of insect and plant colors in digital images using Java software on the Internet. *Annals of the Entomological Society of America* **99**: 865–74.
- Chen T Y, Chu C C, Fitzgerald G, Natwick E T and Henneberry T J. 2004. Trap evaluations for thrips (Thysanoptera: Thripidae) and hoverflies (Diptera: Syrphidae). *Environmental Entomology* **33(5)**: 1416–20.
- Childers C C and Brecht J K. 1996. Coloured sticky traps for monitoring *Frankliniella bispinosa* (Morgan) (Thysanoptera: Thripidae) during flowering cycle in citrus. *Journal of Economic Entomology* **89**: 1 240–9.
- Clare G N, Suckling D M, Bradley S J, Walker J T S, Shaw P W, Daly J M, McLaren G F and Wearing C H. 2000. Pheromone trap colour determines catch of non-target insects. *New Zealand Plant Protection* **53**: 216–20.
- Demirel N and Yildirim A E. 2008. Attraction of various sticky colour traps to *Thrips tabaci* Lindeman (Thysanoptera: Thripidae) and *Empoasca decipiens* Paoli (Homoptera: Cicadellidae) in cotton. *Journal of Entomology* **5(6)**: 389–94.
- Dowell R V and Cherry R H. 1981. Survey traps for parasitoids, and coccinellid predators of the citrus black fly, *Aleurocanthus woglumi*. *Entomologia Experimentalis et Applicata* **29**: 356–62.
- Esler PD, Obrycki J and Nutter FW. 2004. Trap height and orientation of yellow sticky traps affect capture of *Chaetocnema pulicaria* (Coleoptera: Chrysomelidae). *Journal of Economic Entomology* **97(1)**: 145–9.
- Hempel de Ibarra N, Langridge K V and Vorobyev M. 2015. More than colour attraction: behavioural functions of flower patterns. *Current Opinion in Insect Science* **12**: 64–70.
- Lessio F and Alma A. 2004. Dispersal patterns and chromatic response of *Scaphoideustitanus* Ball (Homoptera: Cicadellidae) vector of the phytoplasma agent of grapevine flavescencedoree. *Agricultural and Forest Entomology* **6(2)**: 121–7.
- Lucas Espadas A and Hermosilla Ceron A. 2011. Population control of thrips (*Frankliniella occidentalis*) in table grapes, using capture MASIVA sticky traps and aggregation pheromone [Spanish]. *Agricola Vergel Fruticultura Horticultura Floricultura Citricultura Vid Arroz* **30**: 174–80.
- Mani M, Kulkarni N S, Banerjee K and Adsule P G. 2008. Pest management in grapes, Extension bulletin no. 2, NRC for grapes, Pune, 50 p.
- Mani M, Shivaraju C and Kulkarni N S. 2014. *The Grape Entomology*. Springer, India.
- Natwick E T, Byers J A, Chu C, Lopez M and Henneberry T J. 2007. Early detection and mass trapping of *Frankliniella occidentalis* and *Thrips tabaci* in vegetable crops. *Southwestern Entomologist* **32(4)**: 229–38.
- NRCG. 2012. Annual Report 2011-12, National Research Centre for Grapes, Pune, India, p 116.
- Pearsall I A. 2002. Daily flight activity of the western flower thrips (Thysanoptera, Thripidae) in nectarine orchards in British Columbia, Canada. *Journal of Applied Entomology* **126(6)**: 293–302.

- Pilkington L J, Gurr G M, Fletcher M J, Elliott E, Nikandrow A and Nicol H I. 2004. Reducing the immigration of suspected leafhopper vectors and severity of Australian lucerne yellows disease. *Australian Journal of Experimental Agriculture* **44(10)**: 983–92.
- Raja K M and Arivudainambi S. 2004. Efficacy of sticky traps against bhendi leaf hopper, *Amrasca biguttula biguttula* Ishida. *Insect Environment* **10**: 32–2.
- Roditakis N E, Lykouressis D P and Golfopoulou N G. 2001. Colour preference, sticky trap catches and distribution of western flower thrips in greenhouse cucumber, sweet pepper and egg plant crops. *Southwestern Entomologist* **26**: 227–37.
- Rodriguez-Saona C R, Byers J A and Schiffhauer D. 2012. Effect of trap color and height on captures of blunt-nosed and sharp-nosed leafhoppers (Hemiptera: Cicadellidae) and non-target arthropods in cranberry bogs. *Crop Protection* **40**: 132–44.
- Roubos C R and Liburd O E. 2008. Effects of trap colour on captures of grape root borer (Lepidoptera: Sesiidae) males and non-target insects. *Journal of Agricultural and Urban Entomology* **25(2)**: 99–109.
- SAS Institute. 2012. PROC user's manual, version 9.3 ed 2011. SAS Institute Inc., Cary, NC.
- Saxena K N and Saxena R C. 1975. Patterns of relationships between certain leafhoppers and plants, Part III. Range and interaction of sensory stimuli. *Entomologia Experimentalis et Applicata* **18**: 194–206.
- Scatoni I B, Mujika M V, Franco J, Nunez S and Bentncourt C M. 2007. Population fluctuation of *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae) on *Vitis vinifera* L. cv. Italia in the South of Uruguay [Spanish]. *Boletín de sanidad vegetal. Plagas* **33**: 457–7.
- Sharma J, Upadhyay A K, Adsule P G, Sawant S D, Sharma A K, Satisha J, Yadav D S and Ramteke S D. 2013. Effect of climate change on grape and its value added products. (In) *Climate-Resilient Horticulture: Adaptation and Mitigation Strategies*. Singh et al. (Eds). Springer, p 67.
- Takeuchi Y, Arikawa K and Kinoshita M. 2006. Color discrimination at the spatial resolution limit in a swallowtail butterfly, *Papilio xuthus*. *Journal of Experimental Biology* **209**: 2 873–9.
- Tsitsipis J A, Roditakis N, Michalopoulos G, Palivos N, Pappas D, Zarpas K D, Jenser G, Vaggelas J and Margaritopoulos J T. 2003. A novel scarring symptom on seedless grapes in the Corianth region (Peloponnese, southern Greece) caused by the western flower thrips, *Frankliniella occidentalis*, and pest control tests. *Integrated Protection and Production in Viticulture IOBC/wprs Bulletin* **26(8)**: 259–63.
- Wallis D R and Shaw P W. 2008. Evaluation of coloured sticky traps for monitoring beneficial insects in apple orchards. *New Zealand Plant Protection* **61**: 328–32.
- Yadav D S, Kamte A S and Jadhav R S. 2012. Bio-efficacy of cyantraniliprole, a new molecule against *Scelodonta strigicollis* Motschulsky and *Spodoptera litura* Fabricius in grapes. *Pest Management in Horticultural Ecosystems* **18**: 128–34.