



Effect of storage period on special DUS characters in extant varieties of rice (*Oryza sativa*) for establishing distinctiveness

ARCHANA SANYAL¹, MONIKA A JOSHI², SUDIPTA BASU³ and SHIV K YADAV⁴

ICAR-Indian Agricultural Research Institute, New Delhi 110 012

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ABSTRACT

Under the PPV and FR Act 2001, a number of rice varieties have been registered based on morphological and biochemical DUS descriptors. In the present study, 61 extant varieties of rice (*Oryza sativa* L.) were characterized based on 55 morphological descriptors at different stages of plant growth. However, for the assessment of biochemical characters, namely; phenol reaction of lemma, amylose content and gelatinization temperature, the assessment is based on ripened seed onwards, and even after the seed is stored. Hence, there is a need to determine optimum period after seed ripening for recording biochemical characters to avoid confusion during grouping of varieties for establishing distinctiveness. For this, all the 61 varieties were examined to see the effect of storage period on biochemical characters. The observations were recorded beginning from freshly harvested seed (0 months) vis-à-vis stored seed up to one year (12 months). The data were recorded after every three months up to a period of one year. Results revealed that significant changes begin to occur during six months of storage and continue for remaining period. Phenol reaction of lemma colour in 26 varieties showed significant changes in colour intensity after six months of storage. Gelatinization temperature (GT) and amylose content are important cooking quality traits for rice. Among 61 varieties, 26 showed increase in Gelatinization temperature after six months of storage as ASV decreased from 6.7 to 6.0, while the remaining varieties showed constant ASV. Further, the average percentage of amylose content significantly increased after six months of storage period in all varieties. Hence, storage period significantly influences the character expression of biochemical traits. Therefore, it is recommended that the varieties should be evaluated for the biochemical assessment within six months of harvesting.

Key words: Amylose content of endosperm, Biochemical assessment, Gelatinization temperature, *Oryza sativa*, Phenol reaction of lemma, Rice, Storage period

Cultivated rice (*Oryza sativa* L.) belongs to family Poaceae and at present sustains two thirds of the world's population. Rice is a rich source of carbohydrates (71%) and proteins (7.3%) and provides vitamins, minerals and fiber. It provides 21% of the human per capita energy, and 15% of protein globally (FAO). UPOV models of plant variety protection were not suitable for Indian requirements, thus the Government of India enacted *sui generis* system on the Protection of Plant Varieties and Farmers Act (PPV and FRA) in 2001 for providing protection to plant varieties based on distinctiveness, uniformity and stability (DUS) test along with novelty (Patra 2000). The act envisages that the new and notified/extant plant varieties will be registered and protected on the basis of their morphological and biochemical characters as

listed in the DUS guideline. Thus, it has become necessary for all the organizations to document and characterize all the notified/extant varieties that are in active commerce in their territories.

Under the PPV and FR Act 2001, a number of rice varieties have been registered based on morpho-physiological DUS descriptors. The plant morphological characters are recorded as per the stage of observation listed in DUS guidelines. However, for the assessment of biochemical characters, viz. amylose content, gelatinization temperature and aroma, the assessment is based on ripened seed onwards, and even after the seed is stored, which leads to biochemical changes. Hence, there is an urgent need to determine optimum period after ripening of grain for recording these observations. Biochemical parameters, viz. phenol reaction of lemma, gelatinization temperature and amylose content are the critical parameters for assessment of distinctiveness of genotypes for varietal protection, especially in case of essentially derived varieties, where there is no difference in morphological traits. Hence the present study was aimed to determine the optimum stage for assessment of

¹Ph D Scholar (e-mail: archana0811bhu@gmail.com),
²Principal Scientist (e-mail: monikakshat622@yahoo.com),
³Principal Scientist (e-mail: sudipta_basu@yahoo.com), ⁴Principal Scientist (e-mail: sky_sst@yahoo.com), Division of Seed Science and Technology

biochemical characteristics and to study the influence of storage period on these characters.

MATERIALS AND METHODS

Breeders' seed of sixty-one extant varieties of rice (Table 1) belonging to non-basmati and basmati group was procured from the Breeder Seed Production unit of IARI New Delhi, and raised at the Seed Technology Farm in plot size of five rows with each row of four-meter length. Row to row and plant-to-plant spacing was maintained at 30 × 20 cm during *kharif* 2013 and 2014. The material was replicated thrice and all the recommended agronomic practices were followed to raise a good crop. Freshly harvested seed of all the sixty-one varieties was cleaned and packed in polythene bags of 700 gauges (moisture vapor proof material). Further the seed was stored under low temperature-low humidity (15°C-17°C and 60-70% RH) conditions for storage study. All these biochemical characters were observed up to one year of storage at three months interval, i.e. freshly harvested, three months of storage followed by observation at six, nine and twelve months of storage period. Observations were taken for following biochemical characteristics: Phenol reaction of lemma using 1.5% phenol solution (Chang and Bardenas 1965), gelatinization temperature with 1.7% KOH solution (Little *et al.* 1958) and amylose content (Juliano 1971).

Table 1 List of genotypes/extant varieties of rice

| Genotype | Genotype | Genotype | Genotype |
|-------------------------|-----------------|--------------|-------------------|
| Vikramarya | NDR 359 | Poornima | Sambha Mahsuri |
| Jaya | Pusa Sugandh 2 | Mahamaya | PR 106 |
| PR 113 | Pusa Sugandh 3 | Shymla | Pusa 2-21 |
| CSR 27 | Pusa Sugandh 5 | Saraswati | Pusa 44 |
| Kasturi | Mandya Vijya | IR 8 | Pusa 33 |
| Pant Dhan 11 | Pant Dhan 12 | Lochit | PNR 546 |
| Vasumati | Nidhi | Krishna Veni | PNR 162 |
| Pusa Basmati 1 | Suraksha | Aruna | Sabarmati |
| Kranthi | Salivahana | Sugandhamati | Pusa 834 |
| Improved Pusa Basmati 1 | Taraori Basmati | CSR 13 | Pusa Basmati 1121 |
| Vikash | Annand | HMT PKV | PNR 519 |
| Pant Dhan 4 | CSR 10 | Govind | JD 13 |
| Basmati 370 | VL Dhan 206 | Heera | Pusa Basmati 1401 |
| Jyoti | Vivek Dhan 62 | VL Dhan 221 | PNR 381 |
| Makom | ASD 20 | Rasi | JD 6 |
| | | | Ravi |

The data were analyzed with two factor analyses by using OPSTAT (Online Statistical Analysis Tool).

RESULTS AND DISCUSSION

Phenol reaction of lemma

The phenol colour test, which is an index of polyphenol oxidase activity, is a simple method for grouping the rice varieties into different groups on the basis of lemma colour (Nil, Light Brown, Brown, Dark Brown, Black) (Sripunitha and Sivasubramaniam 2014, Singh *et al.* 2011, Gupta *et al.* 2007, Mor *et al.* 2006). During phenol colour test, phenol gets oxidized into dark colour melanin catalyzed primarily by the enzyme 'tyrosinase' in the seed coat and is under simple genetic control (Bhowal *et al.* 1969). But there is no information about influence of storage period on lemma colour during phenol test for varietal characterization. Hence all the sixty-one varieties were examined on the basis of colour changes after every three months of storage beginning from freshly harvested seed up to a period of one year. All the 61 varieties showed intra-varietal variation and the final colour was recorded on the basis of predominant coloration of replicates by using Wilson colour chart (Royal Horticulture Society). The numerical scoring was done as 0 for nil, 1 for light brown, 2 for brown, 3 for dark brown and 4 for black respectively. Out of 61 extant rice varieties, 35 genotypes (Table 2) showed no colour change throughout the storage period, while remaining 26 genotypes exhibited significant colour change with different rate and variable time period during and after six months of storage (Table 3). The mean value of colour intensity increased from 2.07 (brown) to 3.33 (dark brown) during storage. The maximum colour intensity score (3.59) was observed in Kranthi, Pant Dhan-4, Makom, Mahamaya, Shymla, and Pusa Basmati 1401. While least (1.59) was observed for Vasumati, Taraori Basmati and VL Dhan-206. This variation could be genetically influenced as reported by Bhowal *et al.* (1969). The change in colour intensity was observed at six months in all 26 varieties during experiment. Similar study showing changes in enzymatic activity takes six months had also been reported by Ten Ching- Lee *et al.* (1965). This

Table 2 Grouping of 61 extant rice varieties based on phenol reaction of lemma

| Group | Lemma reaction | Color intensity | No. of genotype | Varieties |
|-------|----------------|--------------------------|-----------------|---|
| I | Present | not changed | 35 | 3,5,8,13,14,16,17,22,23,24,26,27,29,30,31,34,36,37,39,38,40,41,42,43,44,45,46,50,51,52,53,55,56,59,60 |
| II | Present | changed after six months | 26 | 1,2,4,6,7,9,10,11,12,15,18,19,20,21,25,28,32,33,35,47,48,49,54,57,58,61 |

Table 3 Influence of storage period on colour intensity of lemma pooled for two years

| Varieties | Storage period (months)/Average colour intensity | | | | | Mean |
|-------------------------|--|------|------|------|------|------|
| | 0 | 3 | 6 | 9 | 12 | |
| Vikramarya | 2 | 2 | 3 | 3 | 3 | 2.63 |
| Jaya | 1 | 1 | 2 | 2 | 2 | 1.62 |
| CSR 27 | 2 | 2 | 3 | 3 | 3 | 2.61 |
| Pant Dhan 11 | 1 | 1 | 3 | 3 | 3 | 2.27 |
| Vasumati | 1 | 1 | 2 | 2 | 2 | 1.59 |
| Kranthi | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Improved Pusa Basmati 1 | 2 | 2 | 3 | 3 | 3 | 2.61 |
| Vikash | 1 | 1 | 2 | 2 | 3 | 1.81 |
| Pant Dhan 4 | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Makom | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Pusa Sugandh 3 | 1 | 2 | 3 | 3 | 3 | 2.43 |
| Pusa Sugandh 5 | 1 | 1 | 2 | 2 | 3 | 1.81 |
| Mandya vijya | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Pant Dhan 12 | 1 | 1 | 2 | 3 | 4 | 2.21 |
| Taraori Basmati | 1 | 1 | 2 | 2 | 2 | 1.59 |
| VL Dhan 206 | 1 | 1 | 2 | 2 | 2 | 1.59 |
| Mahamaya | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Shymla | 3 | 3 | 4 | 4 | 4 | 3.59 |
| IR 8 | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Samba Mahsuri | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Pusa 2-21 | 2 | 2 | 4 | 4 | 4 | 3.15 |
| Pusa 44 | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Pusa 834 | 3 | 3 | 3 | 4 | 4 | 3.41 |
| JD 13 | 2 | 2 | 3 | 3 | 3 | 2.61 |
| Pusa Basmati 1401 | 3 | 3 | 4 | 4 | 4 | 3.59 |
| Ravi | 1 | 1 | 2 | 2 | 3 | 1.85 |
| Mean | 2.07 | 2.10 | 3.10 | 3.17 | 3.33 | |

could be possible reason for variation in colour showed by lemma in various period of time interval.

Gelatinization temperature (GT)

Gelatinization temperature is an important cooking quality trait for the varietal characterization in rice. GT measures starch gelatinization temperature which is a chemical reaction process in which ungelatinized part of starch is changed to gelatinized part (Yamamoto *et al.* 2004). Interpretation of gelatinization temperature can be done with the help of Alkali spreading value (ASV) of starch endosperm. A low ASV corresponds to the high gelatinization temperature (Pachauri *et al.* 2013). The gelatinization parameters are important to breeders who select lines with specific starch physicochemical characteristics. All 61 genotypes were grouped in two broad classes, viz. varieties having constant ASV throughout

storage period and varieties exhibiting changes in ASV value (Table 4).

It was observed that during entire storage period, 35 genotypes showed constant ASV, viz. Vikramarya, Jaya, CSR 27, Rasi while remaining 26 genotypes showed significantly decrease in ASV at six months of storage (viz. Suraksha, ASD 20) and continued up to remaining period (Table 5). The mean ASV value for 26 genotypes decreased from 6.7 to 4.5 during storage period. The ASV decreased significantly during six months of storage (6.7 to 6) resulting in more harder grain at the end of study (Table 5). The gelatinization parameters are influenced by amylopectin structure (chain length distribution), which can vary by cultivar, location and crop year (Cameron *et al.* 2008, Singh *et al.* 2006). The maximum decrease in ASV was observed in Nidhi (4.19) while least (6.25) was found in ASD 20, Poornima, PNR 162, Pusa 834 and JD 13.

Zhou *et al.* (2002, 2003) suggested that during storage, lipid hydrolysis produces free fatty acid and the complexation of free fatty acid with amylose probably occurred after gelatinization which explains the influence of storage on viscosity. Faruqa *et al.* (2015) and Wiset *et al.* (2005) also reported same findings as in present study, that the grain becomes harder and gives high GT after significant storage period (5-6 months).

Amylose content in endosperm

Amylose content is one of the important biochemical traits for varietal characterization. In DUS guideline (National DUS guideline for rice SG/01/2007), varieties are classified in to five groups on the basis of amylose content in endosperm namely: very low (<10%), low (10-19%), medium (20-25%), high (26-30%) and very high (>30%) for varietal registration under PPV and FRA 2001. In the present study, out of 61 genotypes, 22 genotypes had low amylose content (16.2 to 19.6%). Further, 37 genotypes had medium (21.2 to 25.5%) amylose content while only two genotypes, viz. Mahamaya and Jyothi were grouped in

Table 4 Grouping of sixty-one genotypes on the basis of ASV

| Group | ASV test | Consistency In ASV | No. of genotype | Varieties |
|-------|--------------------|---------------------------|-----------------|---|
| I | + | Constant | 35 | 1, 2, 4, 5, 6, 7, 9, 15,17, 25, 33, 34, 35, 36, 37, 38, 39, 40,41,42,43,44,46,45, 47, 48, 49,50,51,52, 54,56,57,59, ,60 |
| II | + | Decrease after six months | 26 | 3,8, 10,11, 12, 13,14, 16, 18,19, 20, 21, 22, 23, 24, 26,27, 28, 29, 30, 31, 32,53, 55, 58 61 |
| | Factors | C.D. | SE(d) | SE(m) |
| | Genotype (A) | 0.064 | 0.033 | 0.023 |
| | Storage period (B) | 0.028 | 0.014 | 0.01 |
| | Factor (A × B) | 0.144 | 0.073 | 0.051 |

Table 5 Influence of storage period on ASV (Alkaline Spreading Value), pooled for two years

| Varieties | Storage period (months)/Alkali spreading value | | | | | Mean |
|--------------------|--|-----|-------|-----|-------|------|
| | 0 | 3 | 6 | 9 | 12 | |
| PR 113 | 7 | 7 | 7 | 5 | 5 | 6.19 |
| Pusa Basmati 1 | 7 | 7 | 7 | 5 | 5 | 6.19 |
| Improved Pusa | 7 | 7 | 5 | 5 | 5 | 5.77 |
| Basmati 1 | 7 | 7 | 5 | 5 | 5 | 5.79 |
| Vikash | 7 | 7 | 7 | 5 | 5 | 6.19 |
| Pant Dhan 4 | 7 | 7 | 7 | 5 | 5 | 6.17 |
| Basmati 370 | 7 | 7 | 5 | 3 | 3 | 4.97 |
| Makom | 5 | 5 | 5 | 3 | 3 | 4.22 |
| Pusa Sugandh 2 | 7 | 7 | 5 | 5 | 5 | 5.83 |
| Pusa Sugandh 3 | 7 | 7 | 7 | 5 | 5 | 6.15 |
| Pusa Sugandh 5 | 7 | 7 | 5 | 5 | 5 | 5.79 |
| Mandya Vijya | 7 | 7 | 5 | 5 | 5 | 5.79 |
| Pant Dhan 12 | 7 | 7 | 5 | 5 | 5 | 5.79 |
| Nidhi | 5 | 5 | 5 | 3 | 3 | 4.19 |
| Suraksha | 7 | 7 | 5 | 5 | 3 | 5.42 |
| Taraori Basmati | 7 | 7 | 7 | 5 | 5 | 6.23 |
| Annada | 7 | 7 | 7 | 5 | 5 | 6.24 |
| CSR 10 | 7 | 7 | 7 | 5 | 5 | 6.23 |
| VL Dhan 206 | 5 | 5 | 5 | 3 | 3 | 4.18 |
| Vivek Dhan 62 | 5 | 5 | 5 | 3 | sss3 | 4.17 |
| ASD 20 | 7 | 7 | 7 | 5 | 5 | 6.25 |
| Poornima | 7 | 7 | 7 | 5 | 5 | 6.25 |
| PNR 162 | 7 | 7 | 7 | 5 | 5 | 6.25 |
| Pusa 834 | 7 | 7 | 7 | 5 | 5 | 6.25 |
| JD 13 | 7 | 7 | 7 | 5 | 5 | 6.25 |
| Ravi | 7 | 7 | 5 | 5 | 5 | 5.85 |
| Mean | 6.7 | 6.7 | 6.0 | 4.6 | 4.5 | |
| Factor | C.D. | | SE(d) | | SE(m) | |
| Genotype (A) | 0.06 | | 0.03 | | 0.021 | |
| storage period (B) | 0.026 | | 0.013 | | 0.009 | |
| Factor (A × B) | 0.134 | | 0.068 | | 0.048 | |

Scale 1-7 was used for measuring ASV as per DUS guideline

to high category with 26.6 % and 26.1% amylose content, respectively (Table 6). The mean value of amylose content of freshly harvested paddy varieties ranged from 16-26% with the genotypic variation. The differences in amylose content were non-significant during first 3-5 months of storage; however, significant variation was recorded during and after six months of storage period (Table 7). Faruqa *et al.* (2015) also highlighted the same trend in their studies. The average increase in amylose content value was 2.5% which is significantly high enough to bring genotype under higher descriptor category. This up gradation of genotype can create problem during characterization of variety for registration under PPV and FRA 2001. In the present study, significant changes were recorded after six months of storage which was continued further. The mean value of amylose content was 21.5 % at harvesting time which increased by 22.1 % during six months of storage. However, the value of amylose content continued to increase up to 22.6 % at 12 months of storage period. Similar result was reported by Kanlayakrit and Maweang (2013), who reported that after four months of storage period, enhanced enzymatic activity occurs, which contributes in increasing amylose content, and hence, effects the hardness, gel consistency and amylograph viscosity values for cooked rice. Further, amylose content is considered as single most important characteristic for predicting rice cooking and processing quality (Juliano 1985, Webb 1985 and Lii *et al.* 1996). Amylose content is directly related to water absorption, volume expansion, fluffiness and separation of cooked grains (Delwiche *et al.* 1996) while inversely correlated to cohesiveness, tenderness and glossiness (Tester and Morrison 1990).

Hence, from the present study it was concluded that storage temperature and time are the most influential factors on the chemical, physical, and functional qualities of rice during post-harvest storage. The rate and nature of the change is primarily temperature dependent. Generally, the outer layers of seed coat are more susceptible to these reactions than the endosperm of the rice kernel. Although the physicochemical properties of paddy may exhibit greater changes, milled characteristics can also be altered during storage. Quality shifts generally occur faster with increasing temperature. Therefore, the biochemical assessment in rice

Table 6 Grouping of varieties on the basis of amylose content

| Class/state | Low (10-19%) | Medium (20-25%) | High (26-30%) |
|-------------|--|---|--------------------|
| Genotypes | Vikramarya, CSR 27 Pusa Sugandh 3, CSR 10 Pusa Sugandh 2, Vivek Dhan 62, ASD 20, Saraswati, Lochit, Krishna Veni, PR 106 Pusa 221, Pusa 33, PNR 162 Sabarmati, PNR 519, PNR 381, Pant Dhan 11, Vasumati Pant Dhan 4, Salivahana JD 6 | Jaya, PR113, Kasturi, PB 1, IMP. PB 1, Jyothi Vikashbasmati 370, Makom NDR 359, Pusa Sugandha 5, Mandya Vijya, Pant Dhan 12 Nidhi, Surakshatarori Basmati, Annada, VL Dhan 206, Poornima, Shymlair 8, Aruna, sugandhamati. CSR 13, HMT PKV, Govind, Heera, VL Dhan 221, Rasi, Sambha Mahsuri, Pusa 44 PNR 546, Pusa 834 Pusa Basmati 1121, JD 13, Pusa Basmati 1401, Ravi, Kranthi | JYOTHI MAHAMAYA |
| Total | 22 | 37 | 2 |

Table 7 Influence of storage period on average amylose content (%) pooled for two years

| Varieties (V) | Storage period (months)/amylose content (%) | | | | | |
|----------------|---|--------|--------|--------|--------|--------|
| | 0 | 3 | 6 | 9 | 12 | Mean |
| Vikramarya | 18.800 | 18.800 | 19.300 | 19.300 | 20.100 | 19.260 |
| Jaya | 23.400 | 23.650 | 23.900 | 23.900 | 24.400 | 23.850 |
| PR 113 | 22.700 | 22.700 | 22.900 | 22.900 | 23.400 | 22.920 |
| CSR 27 | 18.600 | 18.600 | 19.600 | 19.600 | 20.100 | 19.300 |
| Kasturi | 24.300 | 24.300 | 24.800 | 24.800 | 25.000 | 24.640 |
| Pant Dhan 11 | 19.600 | 19.600 | 20.300 | 20.300 | 20.800 | 20.120 |
| Vasumati | 19.300 | 19.300 | 20.000 | 20.000 | 20.700 | 19.860 |
| PB 1 | 24.100 | 24.100 | 24.500 | 24.500 | 25.900 | 24.620 |
| Kranthi | 25.100 | 25.100 | 25.600 | 25.600 | 25.900 | 25.460 |
| IMP. PB1 | 24.100 | 24.100 | 24.500 | 25.000 | 25.700 | 24.680 |
| Vikash | 25.000 | 25.000 | 25.100 | 25.100 | 25.700 | 25.180 |
| Pant Dhan 4 | 19.300 | 19.300 | 19.900 | 19.900 | 20.400 | 19.760 |
| Basmati 370 | 22.730 | 22.730 | 23.400 | 23.400 | 24.200 | 23.292 |
| Jyothi | 26.600 | 26.600 | 26.800 | 26.800 | 28.900 | 27.140 |
| Makom | 21.640 | 21.640 | 22.400 | 22.400 | 23.400 | 22.296 |
| NDR 359 | 22.850 | 22.850 | 23.100 | 23.100 | 24.100 | 23.200 |
| Pusa Sugandh 2 | 16.300 | 16.300 | 16.900 | 16.900 | 17.000 | 16.680 |
| Pusa Sugandh 3 | 18.300 | 18.300 | 18.900 | 18.900 | 19.000 | 18.680 |
| Pusa Sugandh 5 | 23.230 | 23.230 | 24.100 | 24.100 | 25.000 | 23.932 |
| Mandya Vijya | 24.500 | 24.500 | 24.700 | 24.700 | 25.000 | 24.680 |
| Pant Dhan 12 | 22.300 | 22.300 | 22.600 | 22.600 | 23.000 | 22.560 |
| Nidhi | 22.940 | 22.940 | 23.100 | 23.100 | 23.700 | 23.156 |
| Suraksha | 24.190 | 24.190 | 25.200 | 25.200 | 25.600 | 24.876 |
| Salivahana | 19.450 | 19.450 | 20.400 | 20.400 | 20.600 | 20.060 |
| Tarori Basmati | 24.100 | 24.100 | 25.200 | 25.200 | 25.600 | 24.840 |
| Annada | 24.700 | 24.700 | 25.100 | 25.100 | 25.700 | 25.060 |
| CSR 10 | 17.240 | 17.240 | 18.300 | 18.300 | 18.900 | 17.996 |
| VL Dhan 206 | 20.040 | 20.040 | 21.500 | 21.500 | 21.900 | 20.996 |
| Vivek Dhan 62 | 16.710 | 16.710 | 17.800 | 17.800 | 18.000 | 17.404 |
| ASD 20 | 17.600 | 17.600 | 18.400 | 18.400 | 18.900 | 18.180 |
| Poornima | 25.500 | 25.500 | 26.400 | 26.400 | 26.700 | 26.100 |
| Mahamaya | 26.100 | 26.100 | 26.300 | 26.300 | 26.800 | 26.320 |
| Shymla | 23.360 | 23.360 | 24.100 | 24.100 | 24.600 | 23.904 |
| Saraswati | 16.200 | 16.200 | 17.500 | 17.500 | 17.900 | 17.060 |
| IR 8 | 22.300 | 22.300 | 23.300 | 23.300 | 23.700 | 22.980 |
| Lochit | 17.800 | 17.800 | 18.000 | 18.000 | 18.400 | 18.000 |
| Krishna Veni | 18.800 | 18.800 | 19.200 | 19.200 | 20.300 | 19.260 |
| Aruna | 23.500 | 23.500 | 24.100 | 24.100 | 24.500 | 23.940 |
| Sugandhamati | 25.500 | 25.500 | 25.900 | 25.900 | 25.700 | 25.700 |
| CSR 13 | 24.900 | 24.900 | 25.000 | 25.000 | 25.300 | 25.020 |
| HMT PKV | 24.500 | 24.500 | 25.000 | 25.000 | 25.000 | 24.800 |
| Govind | 20.900 | 20.900 | 21.000 | 21.000 | 21.400 | 21.040 |
| Heera | 22.600 | 22.600 | 23.000 | 23.000 | 23.400 | 22.920 |

Contd.

Table 7 (Concluded)

| Varieties (V) | Storage period (months)/amylose content (%) | | | | | |
|-------------------|---|--------|--------|--------|--------|--------|
| | 0 | 3 | 6 | 9 | 12 | Mean |
| VL Dhan 221 | 21.200 | 21.200 | 22.100 | 22.100 | 22.500 | 21.820 |
| Rasi | 24.500 | 24.500 | 24.900 | 24.900 | 25.000 | 24.760 |
| Sambha Mahsuri | 24.800 | 24.800 | 24.900 | 24.900 | 25.000 | 24.880 |
| PR 106 | 17.900 | 17.900 | 18.000 | 18.000 | 18.500 | 18.060 |
| Pusa 221 | 16.260 | 16.260 | 16.900 | 16.900 | 17.000 | 16.664 |
| Pusa 44 | 22.300 | 22.300 | 22.700 | 22.700 | 22.900 | 22.580 |
| Pusa 33 | 18.700 | 18.700 | 19.000 | 19.000 | 19.400 | 18.960 |
| PNR 546 | 22.300 | 22.300 | 23.000 | 23.000 | 23.600 | 22.840 |
| PNR 162 | 18.900 | 18.900 | 19.000 | 19.000 | 19.400 | 19.040 |
| Sabarmati | 18.900 | 18.900 | 19.000 | 19.000 | 19.400 | 19.040 |
| Pusa 834 | 22.800 | 22.800 | 23.000 | 23.000 | 23.100 | 22.940 |
| Pusa Basmati 1121 | 21.900 | 21.900 | 22.000 | 22.000 | 22.300 | 22.020 |
| PNR 519 | 17.300 | 17.300 | 18.000 | 18.000 | 18.400 | 17.800 |
| JD 13 | 22.100 | 22.100 | 23.000 | 23.000 | 23.400 | 22.720 |
| Pusa Basmati 1401 | 24.100 | 24.100 | 24.800 | 24.800 | 24.900 | 24.540 |
| PNR 381 | 20.200 | 20.200 | 20.700 | 20.700 | 21.300 | 20.620 |
| JD 6 | 19.100 | 19.100 | 19.800 | 19.800 | 20.100 | 19.580 |
| Ravi | 20.400 | 20.400 | 20.600 | 20.600 | 20.900 | 20.580 |
| Mean | 21.563 | 21.567 | 22.107 | 22.115 | 22.580 | |
| Factors | V | M | V × M | | | |
| CD | 0.004 | 0.001 | 0.008 | | | |

should be carried out within first six months of harvesting the produce for establishing distinctiveness among genotypes.

REFERENCES:

- Bhowal J G, Banerjee S K and Joshi M G.1969. The evolution of phenol colour reaction gene in wheat. *Japan Journal of Genetics* **44**: 123–8.
- Cameron D K, Wang Y J and Moldenhauer K A. 2008. Comparison of physical and chemical properties of medium grain rice cultivars grown in California and Arkansas. *Journal of Food Science* **73**(2): 72–8.
- Chang T T and Bardensa E A. 1965. The morphology and varietal characters of the rice plant. Technical Bulletin 4, IRRI, Philippines, p 40.
- Delwiche S R, McKenzie K S and Webb B D.1996. Quality characteristics in rice by near-infrared reflectance analysis of whole-grain milled samples. *Cereal Chemistry* **73**: 257–63.
- Faruqa G, Hossain Z, Prodhana A and Nezhadahmadi A. 2015. Effects of ageing on selected cooking quality parameters of rice. *International Journal of Food Properties* **18**(4): 922–33.
- Gupta N, Joshi M, Sarao A, Navraj K and Sharma R C. 2007. Utility of phenol test in varietal characterization. *Crop Improvement* **34**(1): 77–81.
- Juliano B O. 1971. A simplified assay for milled rice amylose. *Cereal Science Today* **16**: 334–8.

- Juliano B O. 1985. Criteria and tests for rice grain qualities. (In) *Rice Chemistry and Technology*, pp 443-524. (Juliano B O (Ed)., Am. Assoc. Cereal Chem., St Paul, MN, USA.
- Lii C Y, Tsai M L and Tseng K H. 1996. Effect of amylose content on the rheological property of rice starch. *Cereal Chemistry* **73**: 415–20.
- Little R R, Hildre G B and Dawson E H. 1958. Differential effect of dilute alkali on 25 varieties of milled white rice. *Cereal Chemistry* **35**: 111–26.
- Mor R S, Battan K R and Mehla B S. 2006. Identification of rice varieties by laboratory techniques. *Journal of Plant National Improvement* **8**(1): 28–31.
- National Dus Test Guidelines. 2007. Guidelines for the conduct of tests for distinctness, uniformity and stability-Rice (*Oryza sativa* L.). SG/ 01/ 2007.
- Pachauri V, Taneja N, Vikram P, Singh N K and Singh S. 2013. Molecular and morphological characterization of Indian farmers rice varieties (*Oryza sativa* L.). *Agricultural Journal Crop Sciences* **7**(7): 923–32.
- Patra B C. 2000. Collection and characterization of rice genetic resources from Keonjhar district of Orissa. *Oryza* **34**: 324–6.
- Singh D, Kumar A, Kumar N, Kumar V, Kumar S, Bhardwaj A, Sirohi P and Chand S. 2011. Comparison of traditional physico-chemical methods and molecular marker assays for characterization of Basmati rice (*Oryza sativa* L.). *African Journal of Biotechnology* **10**(62): 13390–8.
- Singh N, Kaur L, Sandhu S K, Kaur J and Nishinari K. 2006. Relationships between physicochemical, morphological, thermal, rheological properties of rice starches. *Food Hydrocolloids* **20**(4): 532–42.
- Sripunitha A and Sivasubramaniam K. 2014. Varietal characterization rice varieties based on chemical methods. *Trends in Biosciences* **7**(20) : 3139–46
- Ten Ching Lee, Wu W T and Williams V. R. 1965. The effect of storage time on the compositional patterns of rice fatty acids. *Cereal Chemistry* **42**: 498.
- Tester R F and Morrison W R. 1990. Swelling and gelatinization of cereal starches. I. Effects of amylopectin, amylose, and lipids. *Cereal Chemistry* **67**: 551–7.
- Webb B D. 1985. Criteria of rice quality in the United States. (In) *Criteria of Rice Quality in the United States*, pp 403-42. Webb BD (Ed). Am. Assoc. Cereal Chem., St Paul, MN, USA.
- Wiset L, Kongkiattikajorn J and Potchanachai S. 2005. Effect of free fatty acid contents on pasting behavior of postharvest brown rice flour. 31st Congress on Science and Technology of Thailand at Suranaree University of Technology, pp 18-20.
- Yamamoto H, Makita E, Oki Y and Otani M. 2004: Flow characteristics and gelatinization kinetics of rice starch under strong alkali conditions. *UNI EN ISO 11746:2012 Riso – Determinazione delle caratteristiche biometriche dei grani*.
- Zhou Z, Robards K, Helliwell S and Blanchard C. 2002. Ageing of stored rice: changes in chemical and physical attributes. *Journal of Cereal Science* **35**: 65–78.
- Zhou Z, Robards K, Helliwell S. and Blanchard C. 2003. Effect of rice storage on pasting properties of rice flour. *Food Research International* **36**: 625–34.