

# Effect of diets containing oak kernel on the rumen fermentation and digestibility, blood metabolites and liver enzymes in Khuzestani buffalo

TAHEREH MOHAMMADABADI\*, MOJTABA SHEIKH AZADI and MEHDI BABAEI

University of Khuzestan, P.O. Box 63517-73637, Mollasani, Ahvaz, Iran

Received: 3 September 2019; Accepted: 3 October 2019

#### ABSTRACT

This study was conducted to investigate the effect of different levels of oak kernel on rumen digestibility and fermentation, blood metabolites and liver enzymes of Khuzestani buffalo. Sixteen male buffaloes, one year old were fed with experimental diets containing different levels of oak kernel (0, 15.8%, 31.58% and 47.37% equal to 0, 1.5, 3 and 4.5% tannin, respectively) in a completely randomized design for 35 days. In end of experiment, dry matter intake, digestibility, rumination behaviour, rumen and blood parameters were measured. The results showed that oak kernel increased dry matter intake and digestibility of dry matter and organic matter but the digestibility of protein, NDF and ADF was decreased. Incorporation of 47.37% oak kernel in the diet caused significant decrease in ammonia-N and protozoa population of buffalo rumen, but *p*H and total fatty acids were not affected. The rumination activity in diet containing 47.37% oak kernel was the lowest. With the increasing oak kernel in the buffalo diet, blood cholesterol level decreased, but there were no significant changes in blood glucose, urea, triglycerides, alkaline phosphatase, aspartate and alanin amino transferase enzymes between the experimental treatments. Therefore, even if the digestibility of ADF and CP decreased by adding of oak kernel but the using of oak kernel up to 47.37% in buffalo diet had no negative effect on the rumen fermentation characteristics, digestibility and blood metabolites, hence it may be used in buffalo's diet.

Keywords: Blood metabolites, Buffalo, Digestibility, Fermentation, Oak kernel

One of feed items that can be used as an alternative to livestock in autumn and winter is oak kernel (Rajablo 2009). The main part of the oak kernel is starch (about 55%). Therefore it can be used as a source of energy, and as a substitute for concentrates in ruminant's nutrition (Kilic *et al.* 2010).

The approximate analysis of oak kernel shows that the chemical composition of oak fruits is similar to cereals (Ozcan 2006). The use of oak kernel is restricted due to the presence of anti-nutritional agents (8.8% tannins which 57% is hydrolysable) (Jafari *et al.* 2018).

Researchers reported that oak consumption probably reduces protein degradation in the rumen and growth of proteolytic bacteria. Studies have shown that tannins prevent digestion of lignocellulosic material, and the binding of microbes to feed particles, and consequently reduce digestibility (Rivera-Méndez *et al.* 2017).

Information on use of oak kernel tannin in buffalo nutrition is not sufficient; hence this study was conducted to investigate the effect of oak kernel on the digestibility, rumen fermentation characteristics and blood metabolites of Khuzestani buffalo.

\*Corresponding author e-mail: mohammadabadi@asnrukh. ac.ir

## MATERIALS AND METHODS

Sixteen male khuzestani buffaloes (average weight 290 kg) in the research center of Safiabd, Dezful, Iran were selected. Animals were randomly divided into 4 groups with 4 replications and placed in individual pens places. Drinking water and daily feed were provided separately. Daily meals were administered two times in the morning (08:00 h) and afternoon (16:00 h) to the livestock. The buffaloes diets were formulated on the weight of the animals and according to the dietary requirements (NRC 2000). Treatments consisted of control (without oak) and three levels of oak kernel (15.8, 31.58 and 47.37% equal to 1.5, 3 and 4.5% tannin, respectively). The chemical analysis of oak kernel is as follows: fat 5%, protein 4.8%, carbohydrate 64%, and total tannin 8.5%. Total tannins (TT) were conducted in three replicates as described by Makkar (2000).

Each treatment was randomly assigned to each group of animals for 35 days. All experimental diets were isoenergetic and iso-nitrogenous (Table 1).

In order to determine the apparent digestibility of nutrients, during the last five days of the experiment, feed, feces, and residual feed per animal was collected. The samples were dried in an oven at 60°C for 48 h, milled, then NDF (nautral detergent fibre), ADF (acid detergent fibre) and dry matter were measured by standard methods

Table 1. Ingredient (%) and chemical composition of experimental diets

Ingredient	Treatment			
	1	2	3	4
Corn	25.00	4.00	4.00	0.00
Barely	13.00	19.00	8.00	0.00
Wheat bran	12.00	11.00	6.42	4.00
Oak kernel	0.00	15.80	31.58	47.35
Wheat straw	17.00	12.00	5.50	0.00
Alfalfa hay	18.00	24.00	37.50	49.00
Corn silage	14.00	13.00	6.00	0.00
Salt	0.50	0.50	0.50	0.50
Mineral and vitamin supplement	0.50	0.50	0.50	0.50
Net energy (Mcal/kg)	1.52	1.47	1.44	1.41
Crude protein (%)	10.58	10.61	10.30	10.40
Neutral detergent fibre (%)	37.76	42.76	43.95	45.96
Acid detergent fibre (%)	22.29	26.52	29.53	32.64
Ether extract (%)	2.55	2.54	3.19	3.76
Calcium (%)	0.48	0.57	0.72	0.85
Phosphorous (%)	0.44	0.41	0.33	0.26

<sup>1, 2, 3</sup> and 4 were diets containing 0, 15.8, 31.58 and 47.37% oak kernel equal to 0, 1.5, 3 and 4.5% tannin, respectively.

(AOAC 1990) then digestibility of nutrients was measured. Blood samples were collected 2 h after morning feeding in the tubes containing EDTA and centrifuged (3,000 rpm for 15 min). Then glucose, urea, cholesterol, triglyceride and liver enzymes were measured using Pars Azmon chemical kits and Auto Analyzer (BS200). Also for determination of ammonia nitrogen (phenol hypochlorite method) and *pH* (*pH* meter), rumen fluid was taken, 3 h after morning meal, by using stomach tube.

For rumen protozoa count, the rumen fluid fixed by using a stabilizing solution of 10% formaldehyde. Methylene blue coloring and counting were performed with optical microscopy (Yanez 2004).

The estimation of total volatile fatty acids in this experiment was carried out according to Getachew *et al.* (2002). The activities of rumination, chewing, resting and eating were recorded every 5 min with in 24 h. Statistical analysis of the obtained data was done in a completely randomized design using General Linear Models (GLM) procedure in SAS software, using the following statistical model:

$$Y_{ij} = \mu + T_i + e_{ijk}$$

where  $Y_{ij}$ , observation;  $\mu$ , general mean,  $T_i$ , effect of experimental treatments and  $e_{ijk}$ , standard error term. The means were compared using Duncan's multiple range tests at 5% level of significance.

## RESULTS AND DISCUSSION

The results of this experiment showed that the dry matter intake of buffaloes (Table 2) increased with increasing of the oak kernel in the diet (P<0.05). In agreement of this study, Rivera-Méndeza *et al.* (2017) reported tannin

Table 2. Effect of different levels of oak kernel on dry matter intake, nutrient digestibility and rumen parameters in buffaloes

Parameter		Treatn	SEM	P value		
	1	2	3	4	=	
Dry matter intake (g/d)	7125 <sup>b</sup>	8250 <sup>ab</sup>	8350 <sup>ab</sup>	8500a	314.49	0.03
Digestibility						
Dry matter	78.33 <sup>b</sup>	81.18 <sup>a</sup>	83.32a	85.05a	1.62	0.04
Crude protein	43.9	30.12	26.8	23.31	4.71	0.03
Organic matter	72.24 <sup>b</sup>	76.32ab	79.64ab	83.51a	2.51	0.03
NDF	55.7	52.11	48	45.21	4.09	0.2
ADF	44	39.1	38.2	33.1	2.2	0.012
Rumen parameters						
pН	7.07	6.77	6.67	6.63	0.05	0.05
Ammonia-N (mg/dL)	16.62a	13.23 <sup>b</sup>	8.28 <sup>b</sup>	6.81 <sup>b</sup>	0.62	0.02
VFA (mmol/L)	75.1	77.1	76	78	0.3	0.4
Protozoa (Cell/ mL rumen fluid*10 <sup>4</sup>	6.2ª	5.1 <sup>b</sup>	4.8 <sup>b</sup>	3.4 <sup>b</sup>	0.27	0.02

1, 2, 3 and 4 were diets containing 0, 15.8, 31.58 and 47.37% oak kernel equal to 0, 1.5, 3 and 4.5% tannin, respectively.

SEM, standard error of the averages; There are significant differences in the numbers in each row with non-identical alphanumeric characters (P<0.05).

supplementation tended to increase dry matter intake in steers during the finishing feed lot phase. But Henke *et al.* (2017) reported quebracho tannin extract did not have any effect on feed intake of dairy cows and intake of organic matter (Al-Kindi *et al.* 2016). Shakeri and Fazaeli (2005) reported that the 30% dried pistachio byproducts in the diet of lamb's decreased feed intake

Increasing oak kernel level in diet cause to increase dry matter and organic matter digestibility and the highest dry matter digestibility was for treatment containing 4.5% oak tannin. But the digestibility of protein, NDF and ADF was decreased (P<0.05).

One of the reasons for increasing digestibility of dry matter in diets containing oak kernel may be attributed to the higher amounts of non-structural carbohydrates (NSC) in these diets. Ghasemi *et al.* (2010) also indicated that replacement of pistachio hulls (7.7% phenolic compounds) up to 50% in Baluchi sheep's, increased the digestibility of dry matter and organic matter, due to the higher content of NSC in these diets in compared to alfalfa diet.

The recent results showed the protein, NDF and ADF digestibility was reduced by oak kernel. Besharati *et al.* (2008) reported that raisin residue as a tannin source in Ghezel sheep reduces the protein digestibility. But Henke *et al.* (2017) reported quebracho tannin extract did not influence N digestibility. Tannins reduce the fibre digestion through the formation of a complex with lignocellulose, binding to microorganisms and extracellular enzymes of microorganisms.

Mohammadabadi and Jolazadeh (2017) reported that NDF and ADF digestibility after 24 and 48 h by rumen

bacteria was not affected by inclusion of subabul leaves (containing tannin) in goat's diet. Aguerre *et al.* (2016) reported that quebracho-chestnut tannin extracts reduced digestibility of dry matter, organic matter, crude protein and NDF.

The ammonia nitrogen concentration (NH<sub>3</sub>-N) and protozoa population significantly decreased by increasing oak kernel level in the buffalo diet (P<0.05). Bhatta *et al.* (2012) reported tannin sources reduce the ammonia nitrogen content in the rumen.

Mohammadabadi and Chaji (2012) reported the NH<sub>3</sub>-N in media culture decreased by processing sunflower meal with 30 g/kg DM oak fruit tannin. Also quebracho-chestnut tannin extracts reduced ruminal NH<sub>3</sub>-N concentration (Aguerre *et al.* 2016). Sunflower meal treated with pistachio hull extract decreased *in vitro* ruminal NH<sub>3</sub>-N and pH (Jolazadeh and Mohammadabadi 2017). In general, the reduction in the NH<sub>3</sub>-N indicates that tannin of oak kernel protects the protein against rumen microbial digestion.

With the increase of oak kernel levels in the diet, the pH did not change. But Maldar et al. (2010) reported that the consumption of oak leaves and Jolazadeh and Mohammadabadi (2016) concluded sunflower meal treated with pistachio hull tannin decreased rumen pH.

Considering that with the increase of oak level in the diet, the cell wall content (especially NDF) decreased and by decreasing the rumen stimulation; salivary secretion and pH was reduced. Also one of the causes of decrease of rumen *p*H with increasing oak level in the diet is high carbohydrates of oak kernel.

The increase of oak kernel levels in the diet caused to decrease the protozoa population. Also incorporation of oak a corn to the diets decreased total protozoa (Aghamohamadi *et al.* 2014).

One of the reasons for the decrease in the protozoa population can be reduction of rumen pH in the diet. Probably other reasons for reduction in rumen protozoa population can be the presence of high amounts of starch in these diets and reducing the growth of proteolytic bacteria by tannins (Maldar *et al.* 2010).

The amount of total volatile fatty acid was not affected by the oak kernel tannin (P>0.05). In contrary to the current experiment, tannins extract of pistachio hull decreased ruminal VFA concentration (Jolazadeh and Mohammadabadi 2017).

Quebracho-chestnut tannin extracts reduced ruminal total branched-chain volatile fatty acid concentration (Aguerre *et al.* 2016). Tannin inclusion did not affect rumen total volatile fatty acid concentration and total protozoa populations (Aboagye *et al.* 2018).

The rumination time in diet containing 15.8% oak kernel was higher than other treatments and decreased linearly with the increase oak kernel in the diets (P<0.05). The highest resting time was observed in diet containing 47.37% oak kernel. The total chewing and eating time/dry matter, linearly decreased with increasing oak kernel in the diet (P<0.05) (Table 3).

Table 3. Effect of different levels of oak kernel on feed intake behavior in buffaloes

Parameter	Treatment				SEM	P value
	1	2	3	4		
Eating (min/day)	385.0	317.5	280.0	264.0	34.20	0.40
Resting (min/day)	670.0 <sup>b</sup>	685.0 <sup>b</sup>	717.5 <sup>a</sup>	745.0 <sup>a</sup>	31.70	0.02
Rumination (min/day)	437.7a	442.5a	415.0 <sup>a</sup>	385.0 <sup>b</sup>	45.85	0.03
Chewing	822.7a	760.0a	695.0 <sup>b</sup>	649.1 <sup>b</sup>	31.70	0.03
Activity for dry m	atter					
Eating	45.1a	44.1a	$33.1^{b}$	$30.0^{c}$	0.30	0.03
Rumination	60.0	50.0	47.0	47.0	5.10	0.40
Chewing	105.0a	94.1 <sup>b</sup>	80.0°	77.0 <sup>d</sup>	0.20	0.03

1, 2, 3 and 4 were diets containing 0, 15.8, 31.58 and 47.37% oak kernel equal to 0, 1.5, 3 and 4.5% tannin, respectively. SEM, standard error of the averages. Figures with non-identical alphanumeric characters in each row differ sigificantly (P<0.05) from each other.

Lower eating times in treatment 4, is probably due to decrease of NDF or increase of non-structural carbohydrates (NSC). Linear decrease in rumination time and chewing was associated with increase in the level of pistachio byproducts and decreasing the amount of NDF, reduction in feed size in the diets, and linear decrease in cell wall digestibility. The small size of the particles causes faster passage rate from the rumen and reduces the digestibility of fibre and dry matter, and reducing the particles size in the diets with the same fibre content reduces the chewing time (Krause and Combs, 2003).

There was no significant difference in blood glucose, urea, and triglyceride levels among experimental treatments (P>0.05). The lowest cholesterol was for diet containing 4.5% oak kernel tannin (Table 4). Sinclair *et al.* (2009)

Tabel 4. Effect of different levels of oak kernel on blood parameters in buffaloes (mg/dl)

Parameter		Trea	SEM	P value		
	1	2	3	4		
Glucose	73.5	77.5	79.5	83.1	8.27	0.20
Urea	31.6	26.3	24.7	19.2	5.8	0.40
Cholesterol	113.0a	87.0 <sup>b</sup>	81.5c	$62.0^{d}$	1.1	0.02
Triglyceride	14.1	12.0	11.1	9.4	3.7	0.30
Alanine amino- transferase	21.67	18.1	14.3	11.2	8.2	1.03
Aspartate amino transferase	188.5	176.0	165.5	153.5	17.8	1.30
Alkaline phosphatase	627.5	602.0	591.0	580.5	52.2	1.10

<sup>1, 2, 3</sup> and 4 were diets containing 0, 15.8, 31.58 and 47.37% oak kernel equal to 0, 1.5, 3 and 4.5% tannin, respectively. SEM, standard error of the averages. Figures with non-identical alphanumeric characters in each row differ sigificantly (P<0.05) from each other.

reported that high levels of tannin in the diet, did not change blood glucose levels, but in lower levels, glucose increased, especially when the level of dietary protein was low.

The results of many experiments show that tannin caused a significant decrease in ammonia concentration due to decreasing protein degradation rate, followed by decreasing plasma urea nitrogen (Ben Salem *et al.* 2005). Sinclair *et al.* (2009) stated increasing of tannin levels, along with increase of blood urea concentrations.

The results showed that the use of different levels of oak kernel had significant effect on serum cholesterol. Flavonoids, phenolic compounds and tannins reduce blood lipids by increasing LDL receptors in the liver, as well as binding to apolipoprotein B, thereby increasing the ability of the liver to remove LDL, and decrease cholesterol. Also, lowering blood cholesterol levels is probably due to excessive excretion of bile by excessive secretion due to stimulation of phenols and tannins, which can reduce serum cholesterol and triglyceride levels (Clover and Vandermeer 1993). Animal studies indicate that secondary plant metabolites may decrease cholesterol by reducing the activity of lipogenic hepatic enzymes and cholesterogenetic enzymes such as glucose 6-phosphatase dehydrogenase and 3-hydroxy-3-methyl-glutaryl coenzyme A. Goats fed diets containing oak acorn had a lower plasma glucose, urea and triglyceride compared with those fed the control diet (Jafari et al. 2018).

The results of this study showed that alkaline phosphatase, aspartate and alanine aminotransferase were not affected significantly by the treatments (P>0.05). Aspartate aminotransferase (ALT) is used to evaluate the protein balance in cows and buffaloes, also synthesized in response to increased protein intake and tissue protein catabolism (Zurek *et al.* 1995). Jafari *et al.* (2018) reported animals fed oak acorn containing diets had higher plasma ALT than those fed the control diet.

Results of one study showed that the plant extract containing antioxidant compounds can enhance and protect the liver tissue and reduce liver transaminases by inhibiting the destructive effects of oxidative compounds, and can reduce the degradation of the liver tissue and thereby reduce the leakage of enzymes. The decrease in liver enzymes is probably due to the oxidative antistress effects of these plants in comparison with the effects of oxidative stress on liver tissue (El Damdash *et al.* 2005).

Thus, even if the digestibility of ADF and CP is decreased by oak kernel; incorporation of oak kernel up to 47.37% in buffalo diet had no negative effect on the fermentation characteristics, digestibility and blood metabolites, hence may be used in buffalo's diet.

#### REFERENCES

Aboagye I A, Oba M, Castillo A R, Koenig K M, Iwaasa A D and Beauchemin K A. 2018. Effects of hydrolyzable tannin with or without condensed tannin on methane emissions, nitrogen use, and performance of beef cattle fed a high-forage diet. *Journal of Animal Science* **96**(12): 5276–86.

- Aghamohamadi N, Hozhabri F and Alipour. D. 2014. Effect of oak acorn (*Quercus persica*) on ruminal fermentation of sheep. *Small Ruminant Research* **120**(1): 42–50.
- Aguerre M J, Capozzolo M C, Lencioni P, Cabral C and Wattiaux M A. 2016. Effect of quebracho-chestnut tannin extracts at 2 dietary crude protein levels on performance, rumen fermentation, and nitrogen partitioning in dairy cows. *Journal of Dairy Science* **99**(6): 4476–86.
- Al-Kindi A, Dickhoefer U, Schlecht E, Sundrum A and Schiborra A. 2016. Effects of quebracho tannin extract (*Schinopsis balansae* Engl.) and activated charcoal on nitrogen balance, rumen microbial protein synthesis and faecal composition of growing Boer goats. *Archives of Animal Nutrition* 70(4): 307–21.
- Association of Official Analytical Chemists (AOAC). 2000. *Official methods of analysis of AOAC international*. 17th ed. AOAC International, Gaithersburg (MD).
- Ben Salem H, Makkar H P S, Nefzaoui A, Hassayoun L and Abidi S. 2005. Benefit from the association of small amounts of tannin-rich shrub foliage (*Acacia cyanophylla* Lindl.) with soybean meal given as supplements to Barbarian sheep fed on oaten hay. *Animal Feed Science Technology* **122**: 173–86.
- Besharati M and Taghizadeh A. 2009. Evaluation of dried grape by-product as a tanniniferous tropical feedstuff. *Animal Feed Science and Technology* **152**: 198–203.
- Bhatta R, Baruash L, Saravanan M, Suresh K P and Sampath K T. 2012. Effect of medicinal and aromatic plants on rumen fermentation, protozoa population and methanogenesis in vitro. Journal of Animal Physiology and Animal Nutrition 97: 446– 56
- El-Demerdash F M, Yousef M I and El-Naga N I. 2005. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. *Food Chemistry and Toxicology* **43**(1): 57–63.
- Getachew G, Blümmel M, Makkar H and Becker K. 1998. *In vitro* gas measuring techniques for assessment of nutritional quality of feeds: A review. *Animal Feed Science Technology* **72**: 261–81.
- Ghasemi S, Naserian A, Valizadeh R and Behgar M. 2010. The effect of phenolic compounds of pistachio hull on digestibility and some rumen fermentative properties of Baluchi sheep. Fourth Iranian Congress of Animal Science, Faculty of Agriculture, University of Tehran (Campus-Karaj).
- Henke A, Dickhoefer U, Westreicher-Kristen E, Knappstein K, Molkentin J, Hasler M and Susenbeth A. 2017. Effect of dietary Quebracho tannin extract on feed intake, digestibility, excretion of urinary purine derivatives and milk production in dairy cows. Archives of Animal Nutrition 71(1): 37–53.
- Jafari H, Fatahnia F, Khatibjoo A, Taasoli G and Fazaeli H. 2018. Effect of oak acorn level on colostrum composition and plasma immunoglobulin G of late-pregnant goats and their kids. *Animal* 12(11): 2300–09.
- Jolazadeh A and Mohammadabadi T. 2017. Effect of treated sunflower meal with tannin extracted from pistachio hulls on in vitro gas production and ruminal fermentation. Veterinary Research Forum 8(3): 203–08.
- Kilic U, Boga M and Guven I. 2010. Chemical composition and nutritive value of oak (*Quercus mbur*) nut and leaves. *Journal of Applied Animal Research* **38**: 101–104.
- Krause K M and Combs D K. 2003. Effects of forage particle size, forage source and grain fermentability on performance and ruminal pH in mid lactation cows. *Journal of Dairy Science* **86**: 1382–97.

- Makkar H P S. 2000. *Quantification of Tannins Tree and Shrub Foliage*. A Laboratory Manual for the FAO/IAEA Coordinated Research Project on Use of Nuclear and Related Techniques to Develop Simple Tannin Assays for Predicting and Improving the Safety and Efficiency of Feeding Ruminants on Tanniniferous Tree Foliage. Joint FAO/IAEA of Nuclear Techniques in Food and Agriculture. Animal Production and Health Sub-Programme, FAO/IAEA Working Document, IAEA, Vienna, Austria.
- Maldar S M, Roozbehan Y and Ali Pour D. 2010. The effect of adaptation period to oak leaves on digstibility and rumen parameters of Alamut goats. *Iranian Journal of Animal Science* **41**(43): 243–52.
- Mohammadabadi T and Chaji M. 2012. The influence of the plant tannins on *in vitro* ruminal degradation and improving nutritive value of sunflower meal in ruminant. *Pakistan Veterinary Journal* **32**(2).
- Mohammadabadi T and Jolazadeh A. 2017. Replacement of alfalfa hay (*Medicago sativa* L.) with subabul (Leucaena leucocephala) leaf meal in diets of Najdi goats: effect on digestion activity of rumen microorganisms. *Tropical Animal Health and Prodution* 49(6): 1309–16.
- NRC. 2000. *Nutrient Requirements of Beef Cattle*. Seventh Revised Edition (National Academies Press).
- Ozcan T. 2006. Total protein and amino acid compositions in the acorns of Turkish *Quercus*. L Taxa. *Genetic Resources and Crop Evolution* **53**: 419–29.
- Rajablo M. 2009. The use of oak fruit in feeding of livestock.

- Newsletter inside the Jihad Agriculture.
- Reed J D. 1995. Nutritional toxicology of tannins and related polyphenols in forage legumes. *Journal of Dairy Science* 73: 1516–28.
- Rivera-Méndez C, Plascencia A, Torrentera N and Zinn R A. 2017. Effect of level and source of supplemental tannin on growth performance of steers during the late finishing phase. *Journal of Applied Animal Research* **45**(1): 199–203.
- Shakeri P and Fazaeli H. 2005. Effect of diets contained pistachio by-product on the performance of fattening lambs. IV International Symposium on pistachio and almond—ISHS-Tehran-Iran. May 22–25.
- Sinclair L A, Hart K J, Wilkinson R G and Huntington J A. 2009. Effects of inclusion of whole-crop pea silages differing in their tannin content on the performance of dairy cows fed high or low protein concentrates. *Livestock Science* 124: 306–13.
- Van Soest P J. 1994. Nutritional Ecology of the Ruminant. 2 ed. Comstock Publ. Assoc., Ithaca, NY.
- Yanez Ruiz D R, Moumen A, Mart'ýn Garc'ýa A I and Molina Alcaide E. 2004. Ruminal fermentation and degradation patterns, protozoa population, and urinary purine derivatives excretion in goats and wethers fed diets based on two-stage olive cake: effect of PEG supply. *Journal of Animal Science* 82: 2023–32.
- Zurek E, Foxcroft G R and Kennelly J J. 1995. Metabolic status and interval to first ovulation in postpartum dairy cows. *Journal of Dairy Science* **78**: 1909–192.