

Molecular characterization of sheep and human isolates of *Echinococcus* granulosus from temperate region of India

A B BEIGH^{1⊠}, M M DARZI¹, S BASHIR², P A DAR¹, B A BHAT³, S NAZKI⁴ and I AHMAD¹

Sher-e-Kashmir University of Agricultural Sciences and Technology, Jammu and Kashmir 190 006 India

Received: 5 December 2019; Accepted: 24 December 2019

Keywords: Cystic Echinococcosis, Genotyping, Human, ITS1, PCR-RFLP, Sheep

Cystic Echinococcosis (CE) is a zoonotic infection of serious public health concern, caused by larval forms (metacestodes) of the tapeworm Echinococcus granulosus. Echinococcosis has carnivores as definitive hosts, and the herbivores and omnivores as intermediate hosts. Humans are infected accidently and are not a part of the natural life cycle of parasite. Echinococcosis is diagnosed by different ways using X-ray, CT scan, immunological and serological tests including modern diagnostic technique, i.e. polymerase chain reaction (PCR), which is very sensitive and specific in detecting echinococcosis infection (Beigh et al. 2017). Further, PCR has also been used in genotyping of E. granulosus to facilitate treatment and vaccination. PCRbased technique, have been used widely for strain characterization within E. granulosus (Arbabi et al. 2017). To the date, ten distinct genetic types (G1-G10) of E. granulosus sensulato (s.l.) have been characterized (Bowles and Mc Manus 1993a, Hassan et al. 2016, Snabel et al. 2016, Hammada et al. 2018). RFLP (Restriction fragment length polymorphism), is a perfect technique by which Echinococcus are identified on the basis of sequence and size of the nuclear genomic region rDNA ITS 1. It is also important technique for genotyping of Echinococcus spp (Bowles and McManus 1993b, Arbabi et al. 2017).

E. granulosus sensu stricto (G1-3) has the widest global distribution (McManus et al. 2012). Different strains of the E. granulosus (i.e. G1, G2, G3 and G5) are observed in animals from Western parts of India (Bhattacharya et al. 2008, Pednekar et al. 2009) and G1 and G3 strains infect the livestock of North India (Singh et al. 2012). The myriad of biologic variety in E. granulosus influences the lifecycle patterns, antigenicity, pathology, transmission dynamics and their sensitivity to drugs (Carmena and Cardona 2014, Carlos and Tamara 2016). The early diagnosis of Echinococcus species might be of importance for the prevention and control measures, diagnostic assays and drug

Present address: ¹Sher-e-Kashmir University of Agricultural Sciences and Technology, Jammu and Kashmir, India. ²Hamdard University, New Delhi, India.³University of Otago, New Zealand.⁴College of Veterinary Medicine, Chonbuk National University, South Korea. [™]Corresponding author email: beighab@gmail.com

therapy (McManus 2010).

Enough studies have not been carried out on the molecular and genetic variations of *E. granulosis* in Kashmir valley. Therefore the present study was designed to find out the genotypes of *E. granulosus* currently infecting Sheep and humans in Kashmir valley, using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) and to estimate the genetic variability within the strains by sequencing rDNA-ITS1 gene.

Sample collection: The present study was conducted from the year 2013–2016 on locally reared sheep, including both slaughtered and naturally dead cases in different regions of Kashmir valley and hydatid cyst fluid collected from human beings operated for hydatidosis in SMHS-Hospital, Srinagar. A total of 2100 sheep were screened. 87 isolates were collected, 85 from sheep and 2 isolates from human beings.

Collection of parasite: Fertile cysts of *E. granulosus* were recognized on the basis of Protoscolices presence. Protoscolices were isolated from the fertile cysts. Prior to DNA extraction, Protoscoleces were washed almost three times using distilled water and preserved in 70% alcohol and then stored in refrigerator until used.

DNA extraction: DNA was isolated from ethanol preserved, frozen or fresh samples using standard phenol/chloroform extraction method and ethanol precipitation (Sambrook *et al.* 1989).

PCR amplification and Restriction fragment length polymorphism (RFLP-PCR): The RFLP-PCR was performed as described by Bowles and McManus (1993c) in the rDNA-ITS1 region of the parasite.

Nucleotide sequencing: DNA derived from individual hydatid cysts was subjected to sequencing by the primers employed in the PCR. The purified PCR product was sequenced in Macrogen Inc. Lab. (Geumcheon-gu, Seoul, Korea). Multiple sequence alignment was done using the MUSCLE (v3.8.31) configured for highest accuracy (MUSCLE with default settings). Data obtained were compared with the NCBI nucleotide gene bank (National Center for Biotechnology Information; www.ncbi.nlm.nih. gov/BLAST/).

In the present study, the region ITS1-PCR and linked

ITS1-PCR-RFLP were used to characterize genotypes of *E*. granulosus. DNA isolated from hydatid cyst Protoscolices recovered from sheep and human isolates in the Kashmir Province. The rDNA-ITS1 fragment of samples including 85 from sheep and 2 of human origin was amplified with BD1 / 4S primers (Bowles and McManus 1993b). The length of amplified fragment for all isolated samples with sheep origin was 1000 bp and was between 1,000 bp and 1,100 bp in human origin samples. However, no amplification was observed in the negative controls. Similar results were reported by other workers (Bowles and McManus 1993b, Bhattacharya et al. 2008, Gholami et al. 2012, Hanifian et al. 2013, Dousti et al. 2013, Hassan et al. 2016, Arbabi et al. 2017). The PCR product obtained was subsequently digested with four restriction enzyme (Rsa 1, Alu, Msp 1 and Taq 1). Rsa 1, Alu 1, Msp 1 yielded identical fragments, 300 and 700 bp in sheep and 325 and 700 bp in humans. TaqI restriction enzyme had no effect on PCR product and after digestion intact 1000 bp fragment was seen, which is in accordance with Bowles and McManus (1993b), Gholami et al. (2012), Arbabi et al. (2017) who reported similar results. Molecular analysis by PCR-RFLP of ITS1 of cattle, buffalo and sheep showed similar patterns with Msp1 and Rsa1 (Bhattacharya et al. 2008). Hanifian et al. (2013) reported that Rsa1 showed two bands approximately 655 bp and 345 bp. Alu1 yielded 800 bp and 200 bp and *Taq1* had no effect on PCR product. Dousti et al. (2013) reported that by digesting amplified ITS1 fragment with Alu1 restriction enzyme, yielded 200 and 800 bp fragments; with RsaI, 345 and 655 bp fragments but the TaqI restriction enzyme had no effect on PCR product. Arbabi et al. (2017) using PCR-RFLP of ribosomal ITS1 gene, reported the existence of common strain (G1) from sheep, cattle, humans and goat. The isolates from all the sheep strains (G1) gave similar RFLP patterns despite of being diverse in their hosts or geographical origin. Hassan et al. (2016) examined human, sheep and goat ITS1 region of E. granulosus isolates by the use of PCR-RFLP and reported that the isolates from the human, sheep and goat samples belong to the same genotype. In India, Pednekar et al. (2009) reported four different genotypes of E. granulosus namely G1, G2, G3 and G5 genotypes in Maharashtra and Western parts of India. Singh et al. (2012) reported 2 genotypes, (G3) and (G1) from Punjab. Similarly, Sharma et al. (2013) have reported 3 genotypes of E. granulosus to infect the livestock in north India namely G1, G2 and G3 genotypes.

Sequencing and phylogenetic analysis: The ITS1 gene fragments of hydatid cyst were sequenced. GenBank (http://www.ncbi.nlm.nih.gov/) was searched for similar sequences (Bowles et al. 1995, Van Herwerden et al. 2000, Bhattacharya et al. 2007, Huttner et al. 2009, Arbabi et al. 2017) with the BLAST program and a significant homology was detected with E. granulosus sequences. All of the isolates examined (GenBank accession nos. KY129666, KY129667, KY129668, KY129669 and KY129670) were identified as corresponding to the sheep strain (G1) of E. granulosus and no other genotypes were detected. The

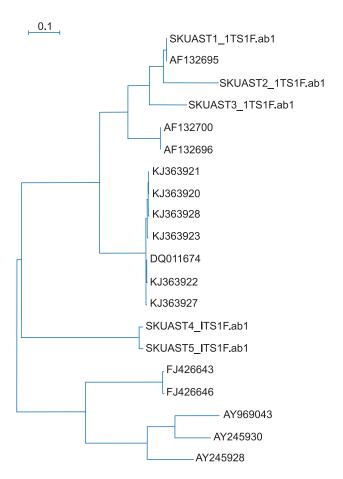


Fig. 1. Genetic relationships of haplotypes, which represent *Echinococcus granulosus* sample from Kashmir valley (SKUAST 1–5), to reference sequences selected from previous studies. The relationships were inferred based on phylogenetic analysis of ITS1 sequence.

phylogenetic tree was reconstructed using the maximum likelihood method implemented in the PhyML program (v3.1/3.0 aL RT) (Fig. 1).

In conclusion, the study inferred that G1 strain in sheep in Kashmir valley is a potential zoonotic parasite and its control both in definitive and intermediate host would in a long way help to curb the disease.

SUMMARY

The aim of the present study was to investigate *Echinococcus granulosus* genotypes in the Kashmir valley. A total of 2,100 sheep and human patients were screened for the presence of hydatidosis. Total 87 isolates were collected, 85 from sheep and 2 isolates from human beings. The rDNA-ITS1 fragment were amplified with BD1 / 4S primers. In addition, fragments of genes coding for ITS1 were sequenced. The length of amplified fragment for all isolated samples with sheep origin was 1,000 bp and with human origin was between 1,000 bp and 1,100 bp. The products on digestion with restriction enzymes *Rsa 1*, *Alu 1*, *Msp 1* yielded identical fragments, 300 and 700 bp in sheep and 325 and 700 bp in humans. Intact 1000 bp fragment was observed with *Taq* 1. The molecular findings

reveal that sheep strain (G1) is the predominant genotype in sheep and humans in Kashmir valley.

ACKNOWLEDGEMENTS

Authors thank the Director of Research, Sher-e-Kashmir University of Agricultural Sciences and Technology, Kashmir for providing financial support to conduct this study. We also thank Mr Ali Mohammad, Mr. Mohammad shafi and staff for their help, cooperation and support extended during the period of study.

REFERENCES

- Beigh A B, Darzi M M, Bashir S, Kashani B, Shah A and Shah S A. 2017. Pathological and histochemical studies of the effects of cystic echinococcosis in sheep. *Comparative Clinical Pathology* **22**: 1–7.
- Bhattacharya D, Bera A K, Bera B C, Maity A and Das S K. 2007. Genotypic characterisation of Indian cattle, buffalo and sheep isolates of *Echinococcus granulosus*. *Veterinary Parasitology* **143**: 371–74.
- Bhattacharya D, Bera A K, Bera B C, Pan D and Das S K. 2008. Molecular appraisal of Indian animal isolates of *Echinococcus granulosus*. *Indian Journal of Medical Research* **127**: 383–87.
- Bowles J and McManus D P. 1993a. Molecular variation in *Echinococcus. Acta Tropica* **53**: 291–305.
- Bowles J and Mc-Manus D P. 1993b. NADH dehydrogenase 1 gene sequence compared for species and strains of the genus *Echinococcus*. *International Journal for Parasitology* **23**: 969–72.
- Bowles J and Mc-Manus D P. 1993c. Rapid discrimination of *Echinococcus* species and strains using a PCR-based method. *Molecular Biochemistry and Parasitology* **57**: 231–39.
- Bowles J, Blair D and Mc-Manus D P. 1995. A molecular phylogeny of the genus *E. granulosus. Parasitology* **110**: 317–28.
- Carmena D and Cardona G A. 2014. Echinococcosis in wild carnivorous species: Epidemiology, genotypic diversity, and implications for veterinary public health. *Veterinary Parasitology* **202**: 69–94.
- Dousti M, Abdi J, Bakhtiyari S, Mohebali M, Mirhendi S H and Rokni M B. 2013. Genotyping of hydatid cyst isolated from human and domestic animals in Ilam province, Western Iran using PCR-RFLP. *Iranian Journal of Parasitology* **8**(1): 47–52.
- Gholami S H, Sosari M, Fakhar M, Sharif M, Daryani A, Hashemi M B and Vahadi M. 2012. Molecular characterization of *Echinococcus granulosus* from hydatid cysts isolated from human and animals in golestan province, north of Iran. *Iranian Journal of Parasitology* 7: 8–16.
- Hanifian H, Diba K, Tappeh K H, Mohammadzadeh H and Mahmoudlou R. 2013. Identification of *Echinococcus* granulosus strains in isolated hydatid Cyst specimens from animals by PCR-RFLP method in West Azerbaijan - Iran.

- Iranian Journal of Parasitology 8(3): 376–81.
- Huttner M, Siefert L, Mackenstedt U and Romig T. 2009. A survey of *Echinococcus* species in wild carnivores and livestock in East Africa. *International Journal for Parasitology* **39**: 1269–76.
- McManus D. 2010. *Echinococcosis* with particular reference to Southeast Asia. *Advances in Parasitology* **72**: 267–303.
- McManus D P, Gray D J, Zhang W and Yang Y. 2012. Diagnosis, treatment, and management of echinococcosis. *British Medical Journal* **344**: 39–44.
- Pednekar R P, Gatne M L, Thompson R C A and Traub R J. 2009. Molecular and morphological characterization of *Echinococcus* from food producing animals in India. *Veterinary Parasitology* **165**: 58–65.
- Sambrook J, Fritsch E F and Maniatis T. 1989. *Molecular cloning: A laboratory manual* (2nd edn.). Cold Spring Harbor Laboratory Press, New York.
- Singh B B, Sharma J K, Ghatak S, Sharma R and Bal M S. 2012. Molecular epidemiology of *Echinococcosis* from food producing animals in north India. *Veterinary Parasitology* 186: 503–06.
- Van Herwerden L, Gasser R B and Blair D. 2000. ITS-1 ribosomal DNA sequence variants are maintained in different species and strains of *Echinococcus*. *International Journal for Parasitology* 30: 157–69.
- Arbabi M, Pirestani M, Delavari M, Hooshyar H, Abdoli A and Sarvi S. 2017. Molecular and morphological characterizations of *Echinococcus granulosus* from human and animal isolates in Kashan, Markazi Province, Iran. *Iranian Journal of Parasitology*. 12: 177–87.
- Hassan H F, Fadhil M H and Fadhil Z H. 2016. Molecular characterization of *Echinococcus granulosus* isolated from human and domestic animals in kirkuk, Iraq. *Animal Research International* 13: 2544 – 47.
- Carlos M and Tamara O. 2016. Molecular epidemiology of cystic Echinococcosis: Genotypic characterization in humans and different livestock. *International Journal of Morphology* **34**: 1472–81.
- Sharma M, Fomda B A, Mazta S, Sehgal R, Singh B B and Malla N. 2013. Genetic diversity and population genetic structure analysis of *Echinococcus granulosus sensu strict* complex based on Mitochondrial DNA signature. *PLoS ONE* 8: 1–8.
- Snabel V, Kuzmina T, Cavallero S, Amelio S D, Georgescu S O, Szenasi Z, Cielecka D, Saamatin R, Yemets A and Kucsera I. 2016. A molecular survey of *Echinococcus granulosus sensu* lato in central-eastern Europe. *Open Life Science* 11: 524–32.
- Oskouei M M, Mehrabani N G, Miahipour A and Fallah E. 2016. Molecular characterization and sequence analysis of *Echinococcus granulosus* from sheep isolates in East Azerbaijan province, northwest of Iran. *Journal of Parasitic Diseases* 40: 785–90.
- Hammada S J, Cavallerob S, Milardib G L, Gabriellib S, Ameliob S D and Al-Nasiria F S. 2018. Molecular genotyping of *Echinococcus granulosus* in the North of Iraq. *Veterinary Parasitology* **249**: 82–87.