## Mycotic endometritis in repeat breeder cows and buffaloes of Himachal Pradesh

PINKI SAINI<sup>1</sup>, MADHUMEET SINGH<sup>2</sup>, PRAVESH KUMAR<sup>3</sup> and B M SHARMA<sup>4</sup>

CSK Himachal Pradesh Agricultural University, Palampur, Himachal Pradesh 176 062 India

Received: 3 June 2018; Accepted: 25 October 2018

Key words: Antifungals, Buffaloes, Cows, Himachal Pradesh, Mycotic endometritis

Among all the reproductive problems, endometritis is one of the most dramatic diseases that causes interference with the functioning of the reproductive tract of cows and hence decreases the livestock profitability (Adnane et al. 2017). Several infectious agents like bacteria, virus, protozoan and fungus are known to have direct impact on reproductive health of cows (Yoo 2010). Fungus is an opportunistic pathogen which can establish itself in disturbed environment of vagina and uterus (Stout 2008). It can cause infection under unusual circumstances such as prolonged intra-uterine antibiotics treatment, local treatment of vagina and cervix, traumatic implantation, pneumovagina (Garoussi et al. 2007). Several workers had reported different species of fungi from reproductive tract of the repeat breeder endometritic cows (Sharma and Singh 2012, Ahmed and Bhattacharyya 2015) and buffaloes (Pal 2002). The purpose of this report was to document the fungal isolations identified from uterus of endometritic cows and buffaloes of Himachal Pradesh.

Repeat breeding is defined as condition when cow/ buffalo is cycling normally, with no clinical abnormalities but has failed to conceive after two or more successive inseminations. Animals selected for this study were endometritic repeat breeder cows and buffaloes of different agro-climatic zones (Dry-temperate, Wet-temperate, Subtropical and Sub-humid) of Himachal Pradesh. The animals selected for this study were maintained under variable managemental conditions in different zones. Uterine discharge samples were collected from 327 cows and 54 buffaloes from different districts of Himachal Pradesh. Collected uterine discharge samples were inoculated on Sabouraud's Dextrose agar plates by spot inoculation technique and the inoculation point was marked outside the plates to rule out the growth of the contaminants. Inoculated plates were incubated at 25-30°C in an upright position. Plates were checked each day for presence of any growth. Fungal isolates were identified by studying gross

Present address: <sup>1</sup>Veterinary Officer (sainipinky142 @gmail.com), Veterinary Hospital Kataula, District Mandi, Himachal Pradesh. <sup>2</sup>Professor and Head (madhumeetsingh2004 @gmail.com), <sup>3</sup>Assistant Professor (pk9919@gmail.com), Department of Veterinary Gynaecology and Obstetrics; <sup>4</sup>Professor (drbrijsharma@gmail.com), Department of Plant Pathology, College of Veterinary and Animal Sciences.

morphological characters and microscopic characters after staining with Lactophenol Cotton Blue stain (Larone 1993). Gross morphological characters included growth rate, colony topography, colony texture, colony pigmentation and microscopic characters included hyphae, spores, vesicle, yeast like cell and any other identifying special feature. Yeast isolates were tested for their sensitivity to various antifungals by disc diffusion method for yeast (according to CLSI procedure) using Muller Hinton agar supplemented with 2% glucose and methylene blue (Himedia Laboratories Pvt. Ltd.) against 8 antifungal diffusion discs (Hi-media) namely itraconazole (10 mcg/disc), clotrimazole (10 mcg/ disc), ketoconazole (10 mcg/disc), amphotericin B (100 units/disc), nystatin (50 mcg/disc), variconazole (1 mcg/ disc), fluconazole (10 mcg/disc) and miconazole (10 mcg/ disc). Different fungal isolates recovered from endometritic cows and buffaloes have been shown in Table 1.

Different geographical location, climate and meteorological factors like temperature, wind speed, relative humidity and rainfall are responsible for level of fungal spores and their dispersion in the atmosphere (Tomas 2003,

Table 1. Fungal isolates obtained

Isolate	N (%)	
Cows		
Rhizopus spp.	6 (17.65)	
Aspergillus niger	4 (11.76)	
Blastomyces spp.	3 (8.82)	
Mucor spp.	3 (8.82)	
Aspergillus spp.	3 (8.82)	
Rhodotorula spp.	3 (8.82)	
Penicillium spp.	3 (8.82)	
Candida spp.	2 (5.88)	
Penicillium vermiculatum	1 (2.94)	
Aspergillus versicolor	1 (2.94)	
Murogenella spp.	1 (2.94)	
Carvularia spp.	1 (2.94)	
Nigrospora spp.	1 (2.94)	
Alternaria spp.	1 (2.94)	
Pithomyces chartrum	1 (2.94)	
Buffaloes		
Rhodotorula spp.	1 (33.33)	
Mucor spp.	1 (33.33)	
Fusarium spp.	1 (33.33)	

Table 2. Incidence of mycotic endometritis in cows and buffaloes in different agro-climatic zones of Himachal Pradesh

Agro-climatic zone	Cows			Buffaloes		
	Total samples	Positive for fungal growth	Incidence (%)	Total samples	Positive for fungal growth	Incidence (%)
Dry temperate (2200 m AMSL)	68	10	14.70	_	_	_
Wet temperate (1801-2200 m AMSL)	93	10	10.75	_	_	_
Sub-humid (651-1800 m AMSL)	59	5	8.47	15	1	6.66
Sub-tropical (350-650 m AMSL)	107	9	8.41	39	2	5.12
Overall	327	34	10.39	54	3	5.55

AMSL, above mean sea level.

Okten et al. 2005). Sharma and Singh (2012) corroborated our findings as they also reported 15.50 and 3.70% incidence of mycotic endometritis in cows and buffaloes, respectively. Ahmed and Bhattacharyya (2015) reported a comparative high prevalence (17.98%) of pathogenic fungi in repeat breeder cattle and buffaloes. Fungi are common in indoor and outdoor environment. Higher incidence (14.70%) of mycotic endometritis in cows was found in dry temperate zone (Table 2). This may be due to indoor favourable environment of cow sheds for fungal growth, as the cows in these areas are mostly confined in their sheds for almost throughout the year. The negative balance of energy, vitamin and minerals may lead to immunosuppression (Galvao 2013), which could be another reason for high incidence of mycotic endometritis in cows of these districts, as they are just maintained on available fodder crops only. The comparatively low incidence (8.47 and 8.41%) was found in sub-humid and sub-tropical zone (Table 2) than dry temperate zone which may be attributed to immunocompetence of the animals. Incidence of mycotic endometritis in buffaloes was reported from sub-tropical and sub-humid zones which may be due to favourable temperature and humidity conditions for fungal growth.

In this study, yeast isolates were found 100, 75, 75 and 37.5% sensitive to itraconazole, miconazole, clotrimazole and ketoconazole. Nearly similar high sensitivity of antifungals had been recorded in mares where itraconazole, miconazole, clotrimazole and ketoconazole were 97, 74, 94 and 100% sensitive, respectively (Beltaire *et al.* 2012). However, a comparatively low susceptibility of itraconazole and miconazole with high sensitivity against clotrimazole and ketoconazole had also been reported (Ferris 2017). Others antifungals namely fluconazole, variconazole, nystatin and amphotericin-B were 100% resistant. There is no known specific reason for their resistance, but it may be due to intrinsic/ natural resistance of these yeast isolates owing to less use of antifungals in field conditions.

This study was aimed to explore the fungal etiology of endometritis in repeat breeder cows and buffaloes of different agro-climatic zones of Himachal Pradesh. Uterine discharge samples were collected from 327 endometritic cows and 54 buffaloes and were subjected to fungal isolation by culturing on Sabouraud's Dextrose agar. Overall incidence of mycotic endometritis was 10.39 and 5.55% in cows and buffaloes, respectively. The mycotic isolates from cows included

Rhizopus spp., Aspergillus niger, Blastomyces spp., Mucor spp., Aspergillus spp., Rhodotorula spp., Penicillium spp., Candida spp., Penicillium vermiculatum, Aspergillus versicolor, Murogenella spp., Carvularia spp., Nigrospora spp., Alternaria spp. and Pithomyces chartrum. Fusarium spp., Mucor spp. and Rhodotorula spp. were isolated from buffaloes. Yeast isolates were subjected to in vitro antifungal sensitivity testing and itraconazole antifungal was found to be most sensitive.

## REFERENCES

Adnane M, Kaidi R, Hanzen C and England G C W. 2017. Risk factors of clinical and subclinical endometritis in cattle: a review. *Turkish Journal of Veterinary and Animal Sciences* 41: 1–11.

Ahmed K and Bhattacharyya D K. 2015. Isolation of pathogenic fungi associated with repeat breeder bovine. *Intas Polivet* **16**(2): 459.

Beltaire K A, Cheong S H and Silva M A C. 2012. Retrospective study on equine uterine fungal isolates and antifungal susceptibility patterns (1999–2011). *Equine Veterinary Journal* 44: 84–87.

Ferris R A. 2017. Therapeutics for infectious endometritis: A clinical perspective. *Revista Brasileira de Reproducao Animal*, *Belo Horizonte* **41**(1): 75–179.

Galvao K N. 2013. Uterine diseases in dairy cows: understanding the causes and seeking solutions. *Animal Reproduction* 10(3): 228–38.

Garoussi M T, Khosrave A R and Havareshti P. 2007. Mycoflora of cervicovaginal fluids in dairy cows with or without reproductive disorders. *Mycopathologia* **164**: 97–100.

Larone D H. 1993. *Medically Important Fungi—A Guide to Identification*. 2<sup>nd</sup> edn. American Society for Microbiology, Washington, DC.

Okten S, Asan A, Tungan Y and Ture M. 2005. Airborne fungal concentrations in East Patch of Edrine City (Turkey) in autumn using two sampling methods. *Trakya University Journal of Science* **6**(1): 97–106.

Pal M. 2002. Endometritis in a water buffalo due to *Candida albicans*. *Buffalo Bulletin* **21**(1): 10–11.

Sharma S and Singh M. 2012. Mycotic endometritis in cows and its therapeutic management. *Intas Polivet* **13**(1): 29–30.

Stout T A E. 2008. Fungal endometritis in the mare. *Pferdeheilkunde* **24**(1): 83–87.

Tomas J B. 2003. Mesa redonda: estado actual de la alergia a hongos. *Alergologiae Inmunologia Clinica* **18**(3): 106–21.

Yoo H S. 2010. Infectious causes of reproductive disorders in cattle. *Journal of Reproduction and Development* **56**: S53–56.