

## Anti-stress properties of *Citrus limon* in chickens

PAWAN K VERMA<sup>1</sup>, YASHWANT SINGH<sup>2</sup> and JITENDER KUMAR<sup>3</sup>

Sher-e-Kashmir University of Agriculture and Technology-Jammu, R. S. Pura, Jammu and Kashmir 181 102 India

Received: 3 August 2009; Accepted: 2 May 2010

**Key words:** Antioxidant status, Chickens, *Citrus limon*, Lipid peroxidation

Stress, a major concern for poultry industry especially in hot regions of the world reduces feed intake, body weight gain, carcass yield, increase morbidity and mortality and susceptible to changes in behavioural and physiological response in broilers (McGeehin and Mirabelli 2001, Altan *et al.* 2003). Heat stress increases oxygen consumption resulting into increased production of free radicals/reactive oxygen species (ROS) possibly by disruption of the electron transport assemblies of the membrane (Valko *et al.* 2007). Increased production of free radicals/ROS may be responsible for molecular changes in DNA, proteins, lipids and other biological molecules (Bruskov *et al.* 2002). Lemon (*Citrus limon* L.) is a good source of vitamins and important macro- and micro-minerals, carbohydrates, proteins, lipids and beta-carotene (Paul and Shaha 2004). Poultry has an ability to synthesize vitamin C, but this ability is inadequate under stress conditions (Padayatty *et al.* 2003). Vitamin C supplementation increases the feed conversion ratio and immune status in birds (Takahashi *et al.* 1991). It also alleviates the negative side effects of stress (Schwedhelm *et al.* 2003). Decrease in plasma ascorbate levels also reduces the plasma concentration of vitamin E (Padayatty *et al.* 2003). Therefore present study was planned to elucidate the anti-stress effects of lemon water supplementation in Vanaraja chicks during summer.

**Experimental protocol and animals:** Day-old chicks (120) of Vanaraja were procured from the Department of Animal Husbandry, J&K. After 2 weeks of brooding, chicks were randomly divided into 2 groups of 40 chicks with 4 replicates of 10 chicks each. Group 1 chicks were kept as control and offered normal diet whereas, group 2 chicks were offered

lemon juice 2 ml/litre (0.2% v/v) in drinking water continuously from 2 to 8 weeks. The study was conducted during summer (May and June 2008), and during entire study period environmental temperature and humidity were recorded. Temperature humidity index (THI) was calculated as per Mc Dowell (1972).

**Biochemical assays:** Blood was collected directly from the heart using 18 gauge needle in clean sterile heparinised tubes from 2 chicks of each replicates and 8 chicks from each group after 2 weeks of supplementing lemon water (4 weeks of age). At 8 weeks, blood was collected from the tarsal vein. Whole blood was used for the estimation of blood glutathione (GSH) and haemoglobin. Erythrocyte lysate (1%) was used for the catalase, superoxide dismutase (SOD), glutathione peroxidase (GSH-Px) whereas, 33% lysate was used for the determination of lipid peroxidation (LPO) activity as per Ohkawa *et al.* (1979). The activity of SOD and catalase was measured according to Marklund and Marklund (1974) and Aebi *et al.* (1987) respectively. The GSH-Px and GSH activity in erythrocyte lysate were assayed by the methods of Hafeman *et al.* (1974) and Beutler *et al.* (1975) respectively. Total protein, albumin, total cholesterol, creatinine, total bilirubin and plasma calcium concentration were estimated by standard kits using chemistry analyzer. Globulin was estimated by subtracting albumin from the total

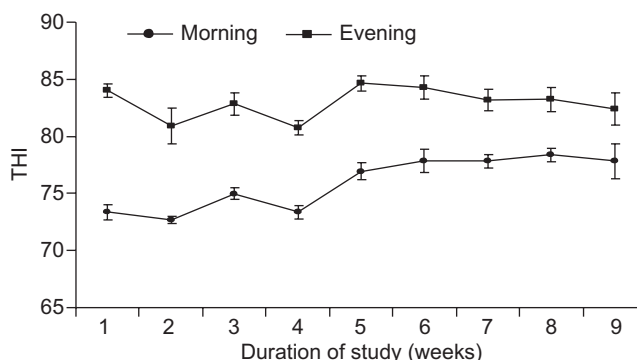


Fig. 1. Alterations in the temperature humidity index (THI) of morning and evening during entire periods of the study period.

Present address: <sup>1</sup>Assistant Professor, Division of Pharmacology and Toxicology, Faculty of Veterinary Sciences and animal Husbandry (e mail: drpawankv@yahoo.co.in).

<sup>2</sup>Associate Professor, Department of Animal Breeding and Genetics GADVASU, Ludhiana 141 004 (e mail: ysinghvet@gmail.com).

<sup>3</sup>Associate Professor, Division of Veterinary Physiology, DHUVASU, Mathura 281122 (e mail: jkvermadr@rediffmail.com).

Table 1. Effect of lemon juice supplementation in drinking water on different stress parameters at different age groups in Vanaraja chickens

| Stress parameters (units)                                                                    | Four weeks of chickens     |                             | Eight weeks of chickens    |                             |
|----------------------------------------------------------------------------------------------|----------------------------|-----------------------------|----------------------------|-----------------------------|
|                                                                                              | Control Group-I            | Lemon supplemented Group-II | Control Group-III          | Lemon supplemented Group-IV |
| Catalase ( $\mu\text{mole of H}_2\text{O}_2$ decomposed $\text{min}^{-1} \text{mgHb}^{-1}$ ) | 42.15 <sup>B</sup> ±6.64   | 33.61 <sup>B</sup> ±3.61    | 24.08 <sup>B</sup> ±4.34   | 35.10 <sup>B</sup> ±3.19    |
| SOD ( $\text{mgHb}^{-1}$ )                                                                   | 0.068 <sup>aB</sup> ±0.004 | 0.100 <sup>aB</sup> ±0.020  | 0.165 <sup>bA</sup> ±0.005 | 0.200 <sup>bB</sup> ±0.013  |
| GSH-Px ( $\text{mgHb}^{-1}$ )                                                                | 3.42 <sup>aA</sup> ±0.48   | 5.17 <sup>aA</sup> ±0.16    | 11.92 <sup>bB</sup> ±0.42  | 6.05 <sup>bB</sup> ±0.16    |
| GSH (n moles $\text{ml}^{-1}$ )                                                              | 47.34 <sup>aB</sup> ±3.29  | 62.344 <sup>aA</sup> ±8.13  | 112.85 <sup>bB</sup> ±1.79 | 110.71 <sup>bB</sup> ±6.11  |
| LPO (n moles MDA produced $\text{g Hb}^{-1} \text{h}^{-1}$ )                                 | 3.26 <sup>a</sup> ±0.26    | 2.14 <sup>b</sup> ±0.34     | 3.97 <sup>b</sup> ±0.59    | 3.07 <sup>b</sup> ±0.15     |
| Hb (g %)                                                                                     | 9.78 <sup>a</sup> ±0.37    | 10.05 <sup>a</sup> ±0.37    | 10.72 <sup>B</sup> ±0.35   | 11.40 <sup>B</sup> ±0.25    |
| Total protein (g %)                                                                          | 6.56 <sup>a</sup> ±0.21    | 6.99 <sup>a</sup> ±0.18     | 7.64 <sup>b</sup> ±0.19    | 8.40 <sup>b</sup> ±0.24     |
| Albumin (g %)                                                                                | 1.32 <sup>a</sup> ±0.098   | 1.27 <sup>a</sup> ±0.089    | 2.08 <sup>b</sup> ±0.052   | 2.12 <sup>b</sup> ±0.096    |
| Globulin (g %)                                                                               | 5.67 <sup>a</sup> ±0.139   | 5.72 <sup>a</sup> ±0.117    | 5.82 <sup>a</sup> ±0.096   | 5.71 <sup>a</sup> ±0.135    |
| A/G ratio                                                                                    | 0.23 <sup>a</sup> ±0.015   | 0.22 <sup>a</sup> ±0.014    | 0.37 <sup>b</sup> ±0.014   | 0.36 <sup>b</sup> ±0.006    |
| Creatinine (mg/dl)                                                                           | 1.18 <sup>aA</sup> ±0.029  | 0.89 <sup>aA</sup> ±0.137   | 1.35 <sup>B</sup> ±0.060   | 1.12 <sup>A</sup> ±0.247    |
| Total bilirubin (g/litre)                                                                    | 0.29 <sup>a</sup> ±0.02    | 0.26 <sup>c</sup> ±0.04     | 0.56 <sup>bB</sup> ±0.06   | 0.40 <sup>A</sup> ±0.02     |
| Total cholesterol (mg %)                                                                     | 235.85 <sup>a</sup> ±9.18  | 218.92 <sup>a</sup> ±7.29   | 328.36 <sup>b</sup> ±32.00 | 308.8 <sup>b</sup> ±26.69   |
| Plasma $\text{Ca}^{+2}$ conc. (g/litre)                                                      | 6.99 <sup>a</sup> ±0.22    | 6.79 <sup>a</sup> ±1.86     | 12.45 <sup>b</sup> ±1.87   | 16.00 <sup>b</sup> ±1.25    |

Values are expressed as mean±SE of 8 birds; Capital alphabets: different superscript differs significantly ( $P<0.05$ ) between groups with in a time period (4 weeks); small alphabets: different superscript differs significantly ( $P<0.05$ ) between time periods (48 weeks of age within a group).

protein. The data were expressed as mean±SE. Statistical analyses were done by one-way ANOVA followed by Dunnet's test with  $P \leq 0.05$  as a limit of significance.

Various stress parameters estimated during study at pre-determined time intervals are presented in the Table 1 whereas changes in the THI of morning and evening during entire study is presented in Fig. 1. During stress, excess energy generation in mitochondria served as the primary source of excess superoxide production in tissues (Andreyev *et al.* 2005). In present study, the SOD activity increased significantly ( $P<0.05$ ) in lemon water chickens at 8 weeks of age. Increased erythrocyte SOD activity in birds in lemon water supplementation indicates increased ability of the tissues to handle excessive free radicals generated during oxidative stress (Andreyev *et al.* 2005). Catalase activity decreased significantly in lemon water supplemented group as compared to control. Reduced activity suggested that the supplementation helps in scavenging the excess  $\text{H}_2\text{O}_2$  production during stress, which is responsible for several degenerative diseases (Sen *et al.* 2006). The activity of GSH-Px was significantly high ( $P<0.05$ ) in lemon water group at 4 weeks of age and increase was more in control as compared to treated group at 8 weeks. This may be due to increased ability of cell to remove/scavenge excess inorganic and organic peroxides thus help in protecting cell membrane (Sen *et al.* 2006). In present study, GSH was significantly high ( $P<0.05$ ) in lemon water supplemented group as compared to control. The increased GSH level with age indicates either increase of GSH level synthesis or decrease utilization of GSH (Singh *et al.* 2001). Excess free radicals attack the thiol

group of cysteine residue of proteins and polyunsaturated fatty acid (PUFA) chain of phospholipids of biological membrane and increase in free radical-induced LPO deteriorates the activity of biological membranes (Sen *et al.* 2006). Significant reduction in LPO in group 2 as compared to Group I at 4 weeks of age, suggests either reduced free radicals production or increased scavenging activity in birds due to supplementation of lemon water.

Non-significant increase in total protein, albumin and haemoglobin was observed in group 2 birds as compared to their respective control. However, increase was significant as age increases (McDowell 1989). Similarly Sahin *et al.* (2002) and Gursu *et al.* (2004) reported significant increase in total protein and albumin in vitamin C supplemented Japanese quails reared in heat stress. Nonsignificant decrease in globulin and A : G ratio was observed in group 3 birds as compared with control birds at 8 weeks and significant increase in A : G ratio was observed in both the groups as the age progresses. Creatinine concentration increased significantly in group 4 birds as age progress whereas increase was nonsignificant in lemon water offered birds. Similar findings were also reported by Rao *et al.* (2005) in rats. Total bilirubin concentration decreased significantly ( $P<0.05$ ) at 4 weeks and nonsignificantly at 8 weeks in lemon offered group as compared to control. Total cholesterol decreased non-significantly during the study in treated birds, but increased significantly as the age progresses in both the groups. Similar results were also reported in Japanese quails (Gursu *et al.* 2004) and broilers (Takahashi *et al.* 1991).

## SUMMARY

Significant reduction in lipid peroxidation, catalase and increased activity of SOD, GSH-Px and GSH was observed in chicks of lemon water offered non-significantly birds. Total protein, albumin and haemoglobin were increased whereas concentration of globulin was not changed in lemon water offered chickens when compared with control in both. Alterations in different parameters suggests that supplementation of lemon water (0.2% v/v) reduces stress in chickens during summer either by reducing free radicals production or increasing the scavenging ability.

## REFERENCES

- Aebi H. 1983. *Catalase, Methods Enzymology*. pp. 276–86. (Ed.) Bergmeyer H U. Academic Press, New York.
- Altan O, Pabuccuoglu A A, Altan S, Konyalioglu H and Bayraktar. 2003. Effect of heat stress on oxidative stress, lipid peroxidation and some stress parameters in broilers. *British Poultry Science* **44**: 545–50.
- Andreyev A Y, Kushnareva Y E and Starkov A A. 2005. Mitochondrial metabolism of reactive oxygen species. *Biochemistry* **70**: 200–14.
- Beutler E. 1975. Red cell metabolism. Grune Strottan. *A Manual of Biochemical Methods*. pp. 67–69, New York.
- Bruskov V I, Malakhova L V, Masalimov Z K and Chernikov A V. 2002. Heat-induced formation of reactive oxygen species and 8-oxoguanine, a biomarker of damage to DNA. *Nucleic Acids Research* **30**: 1354–63.
- Gursu F M, Onderci M, Gulcu F and Sahin K. 2004. Effects of vitamin C and folic acid supplementation on serum paraoxonase activity and metabolites induced by heat stress *in vivo*. *Nutrition Research* **24** (2): 157–64.
- Hafeman D G, Sunde R A and Hoekstra W G. 1974. Effect of dietary selenium on erythrocyte and liver glutathione peroxidase in the rats. *Journal of Nutrition* **104**: 580–87.
- Marklund S and Marklund M. 1974. Involvement of superoxide anion radical in autoxidation of pyrogallol and a convenient assay of superoxide dismutase. *European Journal of Biochemistry* **47**: 469–74.
- Mc Dowell L R. 1989. Vitamin A and E. Vitamins in animal nutrition. *Comparative Aspects to Human Nutrition*. pp. 10–52, 93–131. (Ed.) Mc Dowell L R. Academic Press, London.
- Mc Dowell R E. 1972. *Improvement of Livestock Production in Warm Climate*. pp 51–53. W H Freeman & co. San Frisisco.
- McGeehin M A and Mirabelli M. 2001. The potential impacts of climate variability and change on temperature-related morbidity and mortality in the United States. *Environmental Health Prospect* **109**: 185–89.
- Ohkawa H, Ohishi N and Yagi K. 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Annual Biochemistry* **95**: 351–58.
- Padayatty S, Katz A, Wang Y, Eck P, Kwon O, Lee J, Chen S, Corpe C, Dutta A, Dutta S and Levine M. 2003. Vitamin C as an antioxidant: evaluation of its role in disease prevention. *Journal of American Cell Nutrition* **22** (1): 18–35.
- Paul D K and Shaha R K. 2004. Nutrients, vitamins and minerals content in common citrus fruits in the northern region of Bangladesh. *Pakistan Journal of Biological Sciences* **7** (2): 238–42.
- Rao T P, Sakaguchi N, Juneja L R, Wada E and Yokozawa T. 2005. Amla (*Emblica officinalis* Gaertn.) extracts reduce oxidative stress in streptozotocin-induced diabetic rats. *Journal of Medicinal Food* **8** (3): 362–68.
- Sahin K, Sahin N and Yalayoglu S. 2002. Effects of vitamin C and vitamin E on lipid peroxidation, blood serum metabolites and mineral concentrations of laying hens reared at high ambient temperature. *Biological Trace Element Research* **85**: 35–45.
- Schwedhelm E, Maas R, Troost R and Boger R. 2003. Clinical pharmacokinetics of antioxidants and their impact on systemic oxidative stress. *Clinical Pharmacokinetics* **42**: 437–59.
- Sen T, Sen N, Tripathy G, Chatterjee U and Chakraborty S. 2006. Lipid peroxidation associated cardiolipin loss and membrane depolarization in rat brain mitochondria. *Neurochemistry International* **49**: 20–27.
- Singh S N, Vats P, Kumria M M, Ranganathan S, Shyam R, Aroram M P, Jain C L and Sridharan K. 2001. Effect of high altitude (7620 m) exposure on glutathione and related metabolism in rats. *European Journal of Physiology* **84**: 133–37.
- Takahashi K, Akiba Y and Horiguchi M. 1991. Effects of supplemental ascorbic acid on performance, organ weight and plasma cholesterol concentration in broilers treated with propylthiouracil. *British Poultry Science* **32** (3): 545–54.
- Trojanowski J Q. 2003. Rotenone neurotoxicity: a new window on environmental causes of Parkinson's disease and related brain amyloidoses. *Experimental Neurology* **179**: 6–8.
- Valko M, Leibfritz D, Moncol J, Cronin M, Mazur M and Telser J. 2007. Free radicals and antioxidants in normal physiological functions and human disease. *International Journal of Biochemistry Cell Biology* **39** (1): 44–84.