

Histoenzymic localization of hydrolases in goat kidney

R P SAIGAL¹, S K NAGPAL² and B S NANDA³

Punjab Agricultural University, Ludhiana, Punjab 141 004

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The present experiment was undertaken to obtain histoenzymic data in the goat kidney.

Fresh tissues collected from 8 goats were subjected to cryostat sectioning. Sections, 8 µm thick were incubated for the demonstration of alkaline phosphatases, acid phosphatases and glucose-6-phosphatases (Barka and Anderson 1963), 5-nucleotidase and adenosine triphosphatase (Wachstein and Meisel 1956) and nonspecific esterases (Gomori 1952).

The observations showing relative intensities of various enzymes in different segments are summarized in Table 1.

other segments being weak to absent (Fig. 1).

Acid phosphatase (ACPase) activity was strong to very strong in PCT (supra- and perinuclear), very weak in glomerular epithelium and weak in rest of the tubular epithelium of the goat kidney (Fig. 2).

5-Nucleotidase (5-NTase) was moderate to strong in PCT and weak in Bowman's capsule and distal convoluted tubules (DCT) (Fig. 3).

Adenosine triphosphatase (ATPase) recorded strong to very strong reactions in PCT, weak to moderate in DCT and weak in

Table 1. Relative intensities of histoenzymic activities in various segments of nephron in goat

Enzymes	Glomerular epithelium	Proximal convoluted tubule	Henle's loop	Distal convoluted tubule	Collecting tubules
AKPase	±	++++	+	±	-
ACPase	++	++++	++	++	++
5-NTase	++	+++	-	++	-
ATPase	++	++++	++	++/+++	++
G-6-Pase	-	+++/+	-	+++	-
Ease	-	+++/+	+	+	+

-, Absent; ±, doubtful; +, very weak; ++, weak; +++, moderate; +++++, strong; ++++++, very strong.

Alkaline phosphatase (AKPase) reactivity in goat was localized as very strong along the brush borders of the epithelium of proximal convoluted tubules (PCT), the reaction in

rest of the goat nephron (Fig. 4).

Glucose-6-phosphatase (G-6-Pase) was more or less uniformly moderate in PCT and DCT (Fig. 5).

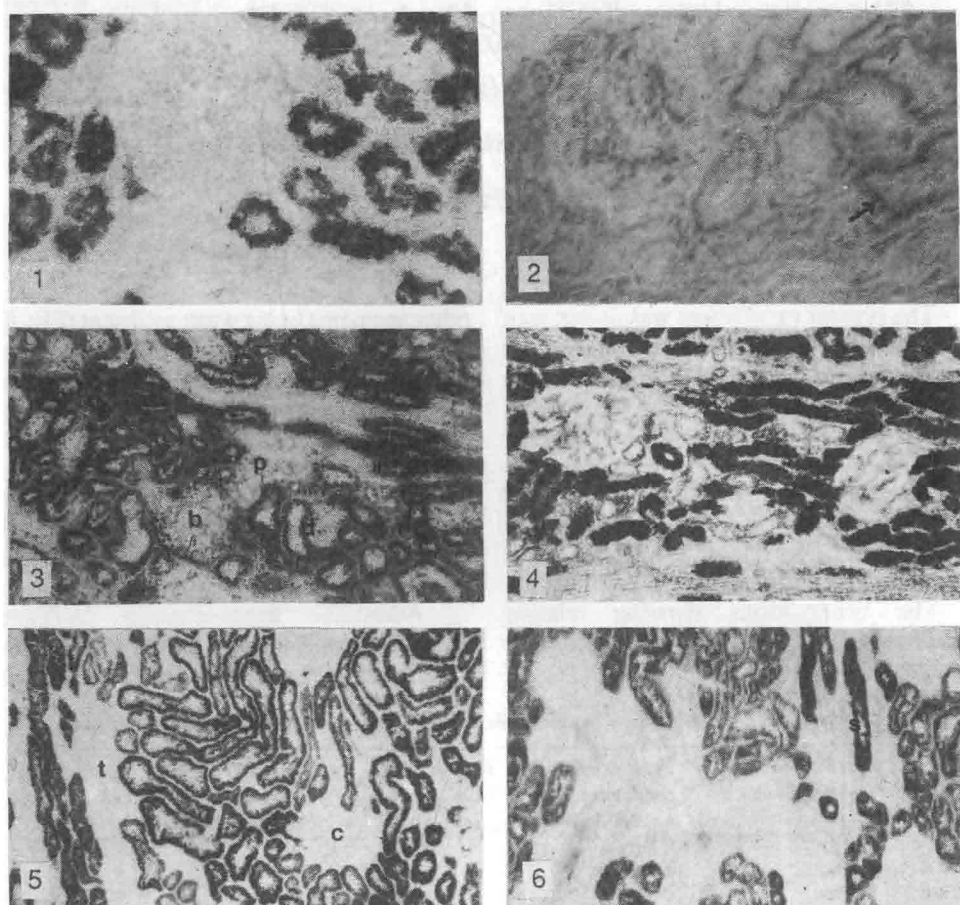
Nonspecific esterases (Ease) reaction product was moderate to strong in the PCT, being relatively stronger in straight tubules and very weak to absent in other segments (Fig. 6).

Thus, in goat PCT all the enzymes showed

Present address: ¹Professor, Department of Anatomy and Histology, College of Veterinary Science.

²Associate Professor, Department of Anatomy and Histology, College of Veterinary Science, CCS Haryana Agricultural University, Hisar 125 004.

³C-3, 3025, Vasant Kunj, New Delhi 110037.



Figs 1-6. Sections of goat kidney showing: 1. AKPase activity in the proximal convoluted tubules. $\times 460$; 2. ACPase activity prominent in proximal convoluted tubules (arrow) $\times 460$; 3. 5-NTase activity with varying intensities in decreasing order in proximal tubules (p), distal tubules (d) and Bowman's capsule (b). $\times 230$; 4. ATPase strongest in proximal tubules. $\times 230$; 5. G-6-Pase strongest in proximal tubules, and not demonstrated in renal corpuscles (c) and thin segments in areas at 't'. $\times 230$; 6. Ease activity strongest in straight segment of proximal tubule (s). $\times 230$.

maximum intensities. In DTC the AKPase was doubtful, Ease was very weak and all other enzymes varied from weak to moderate. The activities were not demonstrated in Henle's loop for 5-NTase and G-6-Pase, in collecting tubules for AKPase, 5-NTase and G-6-Pase, and in glomerular epithelium for G-6-Pase, and Ease; otherwise these 3 segments had very weak to weak reactivities.

The different hydrolases studied in kidney of the goat have more or less similar distribution in different segments of the nephron as in

sheep, cattle and pig (Robinson and Gopinath 1974) and buffalo (Nanda *et al.* 1983). However, certain species variations are concluded. The AKPase was very strong also in thin descending loops in buffalo. The 5-NTase in DCT was localized only in goat. The G-6-Pase was localized in PCT as well as DCT in goat and buffalo, but was limited to PCT in other animals. The Ease was not found in any segment other than PCT and collecting tubules in cattle, sheep and pig, whereas in goat and buffalo it was located with varying intensities

in almost all the segments of the nephron, and was relatively stronger in buffalo than in goat.

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REFERENCES

Barka T and Anderson P.J. 1963. *Histochemistry: Theory, Practice and Bibliography*. pp.233-46. Hoeber

- Medical Division, New York.
Gomori G. 1952. *Microscopic Histochemistry*. pp.206-208. University Press, Chicago.
Nanda B S, Saigal R P and Nagpal S K.1983. Histochemical localization of hydrolytic and oxidative enzymes in the kidney of buffalo. *Indian Journal of Animal Sciences* 53: 1087-93.
Robinson M and Gopinath C. 1974. A comparative histochemical study of some oxidative and hydrolytic enzymes in the kidneys of domestic animals, laboratory animals and the fowl. *Research in Veterinary Science* 16: 335-69.
Wachstein M and Meisel M. 1956. Histochemistry of substrate specific phosphatases at a physiological pH. *Journal of Histochemistry and Cytochemistry* 4: 424-25.