Occurrence of multiple combinations of G and P types of group A bovine and human rotaviruses in Uttarakhand and Nagaland states, India

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Rotaviruses (RVs) are important enteric pathogens. India alone accounts for 22% of 453,000 global RV associated deaths among children <5 years annually (Tate et al. 2012). The RV associated mortality in animals has not been comprehensively testified in India. So far, 27 G- and 37 P-types of RVA have been reported (http://rotac.regatools.be/classificationinfo.html). Although RV infects particular species, heterologous infections may occur. Rotaviruses are continuously evolving due to lack of proofreading activity in RNA polymerase and segmented nature of genome which leads to frequent reassortment (Malik et al. 2013). Sequencing of whole genome of RVs has shed light into complex events of interspecies transmission and reassortment among human and bovine RVs. Owing to high genetic diversity among RVs, present study was undertaken to know circulation status of different genotypic combinations of RVA among bovine and human population in two states of India (Uttarakhand and Nagaland) during 2010–2012.

Diarrhoeic faecal samples (150) from cattle calves (0–3 months) were collected from dairy farms in Pantnagar (Uttarakhand) during December-January in 2010–11 and 2011–12. Diarrhoeic stool samples from human infants (0–5 years), were obtained from Dr Sushila Tiwari Memorial Hospital, Haldwani, Uttarakhand (n=28) during December-January 2011–2012 and Civil Hospital Kohima, Nagaland (23) during June-July 2011. A 10% faecal suspension (w/v) was prepared in phosphate buffer saline.

cDNA synthesis and amplification of full length VP7 gene (1062 bp) and partial length VP4 gene (864 bp) was performed as described earlier (Malik et al. 2012). Bovine RVA genotyping was performed using G3, G6, G8, G10, P[1], P[5] and P[11] typing primers (Iturriza-Gomara et al. 2004). Human RVA genotyping was performed using G1, G2, G3, G4, P8, and P9 genotype specific primers (Iturriza-Gomara et al. 2004).

A typical 4: 2: 3: 2 migration pattern characteristic of RVA was observed in 17.33% bovine and 15.68% human samples. Prevalence of bovine RVA (26/150) is higher than reported in this region. Basera et al. (2010) reported 10.15% bovine RVA and Malik et al. (2014a) detected only 8 samples positive for bovine RVA out of 87 in Hilly areas of Uttarakhand. This may be attributed to period of sampling as peak winter months were chosen when incidence is maximum. All bovine RVA were long electropherotypes having two patterns. In type A segments 1, 2, 3 and 4 migrated separately and in Type B segment 2 and 3 co-migrated (Fig. 1a). 7 and 8 moved as a single segment in all. Basera et al. (2010) reported all 13 bovine RVA as long electropherotypes having 1–4 migrating separately, 2 and 3 co-migrating and 7–9 as single segment. Malik et al. (2012) detected all 17 bovine isolates as long electropherotypes with 7, 8 and 9 segments co-migrating.

In human RVA, 14.28% (4/28) were from Haldwani, Uttarakhand and 17.39% (4/23) were from Kohima, Nagaland. Both long and short electropherotypes were detected (Fig. 1b). No distinct seasonal variation was observed as samples were taken in winter and dry months in Haldwani and summer and humid months in Kohima. The prevalence was less as compared to a study in another North Eastern State, Manipur where Mukherjee et al. (2010) detected ~ 50% human RVA. They found no distinct seasonal variation but a high frequency in children of 3–23 months age group. Dash et al. (2012) reported 46/220 samples positive for human RVA in diarrhoeic children from Uttarakhand and Uttar Pradesh and detected both long and short electropherotypes.

In ELISA, 22% (33/150) bovine samples were confirmed positive for bovine RVA.
positive while 31/150 (20.66%) bovine samples and 8/51 (15.68%) human samples were positive by VP6 gene specific RT-PCR. This indicates ELISA and VP6 gene positive while 31/150 (20.66%) bovine samples and 8/51 (15.68%) human samples were positive by VP6 gene specific RT-PCR. This indicates ELISA and VP6 gene specific RT-PCR being the more sensitive assays than RNA-PAGE.

The first round RT-PCR from bovines yielded 1062 bp and 864 bp amplicons for VP7 and VP4 genes, respectively. The second round PCR produced amplicons of 373, 381 and 397 bps for G3, G6 and G10 genotypes, respectively (Fig. 2). Mixed G3G10 and G3G6G10 types were also observed. P typing produced amplicons 334 bp and 453 bp corresponding to P[11] and P[1], respectively (Fig. 3). P[5] was not detected in any isolate. Among typed bovine samples, G3 and G10 were most predominant G types (23.5%). Mixed G types (G3G10, G3G6G10) were identified as 52.93%. Gulati et al. (1999) detected G10 (83%) as most predominant bovine G type in India. In a recent study on BRV from North India, Manuja et al. (2008) showed 73.3% as G10 type and 26.7% as G6 genotype. In earlier reports, Basera et al. (2010) detected G3 as the most predominant G-type in different regions of Uttarakhand. Malik et al. (2012) showed that 52.9% (9/17) bovine RVAs were G3 and 47% were mixed G types as 35.3% G3G8 (6/17) and 11.8% G3G10 (2/17) in the temperate region of Western Himalaya. In present study, P[1] and P[11] were most predominant P types (23.07%) and 38.46% exhibited dual type (P[1]P[11]). This study is in conformity with earlier report by Malik et al. (2012), where P types were mostly P[11] (94.1%) while 5.9% were the dual type (P[1]P[11]) in temperate region of Western Himalaya.

In human samples, 1062–bp amplicons for VP7 and 877-bp product for VP4 were obtained. G typing produced amplicons of 748 and 812 bps for G1 and G3, respectively (Fig. 4). None of samples was positive for G2 and G4. Mixed G1G3 genotype was also observed. P typing produced amplicons 362 and 224 bps, corresponding to P[4] and P[8], respectively (Fig. 5), P[9] was not detected in any isolate. Among typed human samples, 28.57% were G1, 14.28% were G3 and 57.14 % were mixed (G1G3) genotypes. P[4] was 14.28% and P[8] was 42.85%. In Uttarakhand, G1 and G3 were 14.28%, G1G3 was 57.14%, P[4] was 14.28% and P[8] was 28.56%. In Nagaland, G1 and P[8] were 14.28%. Presence of G1 as most predominant type is in conformity with Mukherjee et al. (2010) who reported G1 as most predominant type (51.63%), followed by G2 (23.36%) and G12 (8.19%) in Manipur. Their P genotype analysis showed P[8] as predominant genotype (39%) followed by P[4] (30%) and P[6] (17.6%). No such data is available on human RVA types in Uttarakhand for comparison. Though this appears to be first report on detection of human RVA from the region, molecular evidences of mixed viral infection with human RVA and Picobirnavirus in an infant hospitalized with acute gastroenteritis in Uttarakhand have been reported (Malik et al. 2014b).


In present study, only 17 bovine and 7 human samples were typed and rest remained non typeable. Similar were findings of bovine RV G typing where Basera et al. (2010) failed to amplify viral gene despite being known to contain sufficient rotaviral particles by RNA-PAGE and sandwich ELISA. In a study conducted by Hussain et al. (1996), neither P9 nor P10 genotype was detected using type specific primers.

Overall, results confirm a changing pattern of circulation of group A RVs in India. The detection of bovine G3, G10, P[1], P[11] types and human G1, G3, P[4], P[8] provides significant information on increasing trend of these genotypes. G3 was reported as most predominant bovine
type in Uttarakhand but based on this study G10 has emerged as a major type circulating in this region. The study on human G and P types has reinforced previous described types and also provided insight into types circulating in two hill states Uttarakhand and Nagaland where reports are scanty. There are 45 different combinations of G and P types that have been recognized (Mukherjee et al. 2011a). The present study, has given newer combinations and emphasized the presence of more number of G types circulating together. In human isolates from Manipur, India, rotavirus strains were found to possess porcine or bovine characteristics (Mukherjee et al. 2011b) indicating possible zoonotic transmission. Therefore, further work is needed to completely characterize the isolates in the present study to get the phylogenetic data.

**SUMMARY**


**REFERENCES**


Hussain M. 1996. ‘Study of variation in rotavirus gene segments 4 and 9 by Polymerase Chain Reaction (PCR) and restriction enzyme analysis’. Ph.D. Thesis submitted to AIIMS, Delhi, India.


