



Comparative lethality of *Salmonella* Enteritidis and *Salmonella* Typhimurium in broiler chickens

MUHAMMAD YOUNUS¹, RAHEELA AKHTAR² and SHAUKAT ALI³

University of Veterinary and Animal Sciences, Lahore, Punjab 54000 Pakistan

Received: 8 August 2014; Accepted: 24 December 2014

Key words: Broiler chickens, Lethal dose, *Salmonella* Enteritidis, *Salmonella* Typhimurium

Lethal dose₅₀ is an accurate estimate of the virulence and pathogenicity of a pathogen. Recent increase in the number of food poisoning outbreaks due to *Salmonella* Enteritidis and *S. Typhimurium* is attributed to increased virulence of this potentially zoonotic enteric pathogen contaminating the poultry meat, eggs and egg products (Utrachkij *et al.* 2011). Both of these pathogens are capable of developing into recovered carriers after disease and are being shed for indefinite period of time. This study was based upon the hypothesis that variations in mortality rates, clinical signs, faecal shedding, and frequency of eggs

contamination in broiler chickens could be associated with differential lethality of *S. Enteritidis* and *S. Typhimurium* therefore it is essential to determine the virulence/lethality of these *Salmonella* species in our locality. As a virulent strain spreads through the body faster, persists for a longer period and is more invasive and difficult to treat than an avirulent strain therefore the information about the increased lethality may help to insight the complex pathogenesis of virulent *Salmonella*.

One-day-old broiler chickens (400) were equally divided into 2 main groups designated as group A (for *S. Enteritidis*)

Table 1. Calculation of dead percentage of broiler chickens by various doses of *S. Enteritidis* and *S. Typhimurium*

Bacterial dilution inoculated	No. of birds		Accumulated No		Proportion	Percentage
	Dead	Live	Dead	Live	Dead/total	Dead
<i>S. Enteritidis</i>						
10 ⁻¹	10	0	31	0	31/31= 1	X100=100
10 ⁻²	10	00	21	0	21/21= 1	X100=100
10 ⁻³	8	2	11	2	11/13= 85	X100=85
10 ⁻⁴	3	7	3	9	3/12= 25	X100=25*
10 ⁻⁵	0	10	0	19	0/19= 0	X100=0
10 ⁻⁶	0	10	0	29	0/29= 0	X100=0
10 ⁻⁷	0	10	0	39	0/39= 0	X100=0
10 ⁻⁸	0	10	0	49	0/49= 0	X100=0
10 ⁻⁹	0	10	0	59	0/59= 0	X100=0
<i>S. Typhimurium</i>						
10 ⁻¹	10	0	25	0	25/25= 1	X100=100
10 ⁻²	10	0	15	0	15/15= 1	X100=100
10 ⁻³	5	5	5	5	5/10= 5	X100=50*
10 ⁻⁴	0	10	0	15	0/15= 0	X100=0
10 ⁻⁵	0	10	0	25	0/25= 0	X100=0
10 ⁻⁶	0	10	0	35	0/35= 0	X100=0
10 ⁻⁷	0	10	0	45	0/45= 0	X100=0
10 ⁻⁸	0	10	0	55	0/55= 0	X100=0
10 ⁻⁹	0	10	0	65	0/65= 0	X100=0

*P < 0.05.

Present address: ¹Principal (younusrana@hotmail.com), College of Veterinary Sciences, Jhang. ²Assitant Professor (raheela.akhtar@uvas.edu.pk), Department of Pathology. ³Veterinary Officer (iwasriwapda@yahoo.com), Veterinary Research Institute, Lahore, Pakistan.

and B (*S. Typhimurium*). Each group was further divided into 10 subgroups comprising 2 replicates. At seventh day of age, the birds in the first 9 subgroups of both group A and B were infected orally with various dilutions of *S. Enteritidis* and *S. Typhimurium* (1 mL/bird of 10⁻¹ to 10⁻⁹

Table 2. Calculation of probits of killing to determine LD₅₀ for *S. Enteritidis* and *S. typhimurium*

Group	Dose	Log dose	% Dead		Corrected%		Probits	
			Group A	Group B	Group A	Group B	Group A	Group B
1	10 ⁻¹	-1	100	100	97.5	97.5	6.96	6.96
2	10 ⁻²	-2	100	100	97.5	97.5	6.96	6.96
3	10 ⁻³	-3	85	50	85	50	6.04	5.00
4	10 ⁻⁴	-4	25	0	25	2.5	4.33	3.04
5	10 ⁻⁵	-5	0	0	2.5	2.5	3.04	3.04
6	10 ⁻⁶	-6	0	0	2.5	2.5	3.04	3.04
7	10 ⁻⁷	-7	0	0	2.5	2.5	3.04	3.04
8	10 ⁻⁸	-8	0	0	2.5	2.5	3.04	3.04
9	10 ⁻⁹	-9	0	0	2.5	2.5	3.04	3.04

Group A, *S. Enteritidis*; group B, *S. Typhimurium*; *P < 0.05.

dilutions, respectively) while the 10th or control group in group A and B was inoculated with normal saline and served as an uninfected control. The inoculated birds were observed for a period of 1 week for any morbidity or mortality (David *et al.* 1998). This data was then used to determine the LD₅₀ of *S. Enteritidis* and *S. Typhimurium* by the method of Randhawa (2009).

The lethal dose (LD₅₀) of *S. Enteritidis* and *S. Typhimurium* determined was 10^{3.58}/mL and 10³/mL respectively (Tables 1, 2).

These results indicated differential killing ability of these 2 *Salmonella* species in broiler chickens under same environmental and nutritional conditions. *S. Typhimurium* was more lethal as compared to *S. Enteritidis* as it could infect the birds even at low dose rate. These findings are in close agreement with previous study (Diez-Garcia *et al.* 2012), which described the differential ability of *Salmonella* species to cause disease in poultry birds.

Our results are also supported by Boldrin-de-Paiva *et al.* (2011) who concluded greatly variable ability of *Salmonella* species to kill day-old chickens. They also described that age of bird and route of infection can play an important role in determining the virulence of *Salmonella* and found that 14 strains belonging to 11 food poisoning serotypes other than *S. Typhimurium* were practically non-lethal when given orally but were lethal by the intramuscular route. Moreover, the bacterial species, strain, phage type, age of bird and inoculum size might also affect the outcome of an infection.

In addition some previous studies (Barrow *et al.* 1987, Gast and Benson 1995) explained that the virulence of *S. Typhimurium* in day-old broiler chickens was dependent upon breeds and found that some breeds were more susceptible than others. However, according to them there was no difference between oral and parenteral administration as far as virulence was concerned.

Our findings also correlated with those of Allen *et al.* (2001) who also stated the severity of clinical illness and enhanced virulence in *S. Typhimurium* DT104. The difference in lethality of 2 *S. Enteritidis* and *S. Typhimurium* may be associated with differences in lipopolysaccharides (Rehman *et al.* 1997) or virulence related plasmid (Jose *et*

al. 1997). There is further need to investigate the difference in genes coding for the virulence in these 2 *Salmonella* species.

SUMMARY

The present study determined the comparative lethality of *Salmonella* Enteritidis and *S. Typhimurium* by calculating the lethal dose (LD₅₀) of these 2 *Salmonella* species in broiler chickens. The objective was to investigate that which *Salmonella* can affect broiler birds at low dose rate and faster than other. The serial 10-fold dilutions of each bacterium were inoculated in 1 week-old broiler birds. The present study revealed that the calculated LD₅₀ of *S. Typhimurium* (10³/mL) was less than *S. Enteritidis* (10^{3.58}/mL). Therefore it can be assumed that *S. Typhimurium* is more virulent than *S. Enteritidis*.

ACKNOWLEDGEMENT

We acknowledge Higher Education Commission (HEC) of Pakistan for funding this research project.

REFERENCES

- Allen C A, Fedorka-Cray P J, Vazquez-Torres A, Suyemoto M, Altier C, Ryder L R, Fang F C and Libby S J. 2001. *In vitro* and *in vivo* assessment of *Salmonella enterica* serovar *Typhimurium* DT104 virulence. *Infection and Immunity* **69**: 4673–77.
- Barrow P A, Huggins M B, Lovell M A and Simpson J M. 1987. Observations on the pathogenesis of experimental *Salmonella* Typhimurium infection in broiler birds. *Research in Veterinary Sciences* **42**: 194–99.
- Boldrin-de-Paiva, Filho J R A C P, Junior A B and Lemos M V F. 2011. Requirement for cobalamin by *Salmonella enterica* serovars *typhimurium*, *pullorum*, *gallinarum* and Enteritidis during infection in broiler birds. *Brazilian Journal of Microbiology* **42**: 1409–19.
- David E S, Glisson R J, Jackwood M W, Pearson J E and Reed W M. 1998. *A Laboratory Manual for the Isolation and Identification of Avian Pathogens*. 4th edn. pp. 261–65. Rose Printing, Tallahassee, Florida.
- Diez-Garcia M, Capta R and Allonso-Calleja C. 2012. Influence of serotype on growth kinetics and the ability to form biofilms of *Salmonella* isolates from poultry. *Food Microbiology* **31**:

- 173–80.
- Gast R K and Benson S T.1995. The comparative virulence for chicks of *Salmonella* Enteritidis phage type 4 isolates and isolates of phage types commonly found in poultry in the United States. *Avian Diseases* **39**: 567–74.
- Jose M, Pena R, Alvarez I, Ibanez M, and Rotger R.1997. Homologous regions of the *Salmonella* Enteritidis virulence plasmid and the chromosome of *Salmonella typhi* encode thiol disulphide oxidoreductases belonging to the DsbA thioredoxin family. *Microbiology* **143**: 1405–13.
- Randhawa M A. 2009. Calculation of LD50 values from the method of Miller and Tainter, 1944. *Journal of Ayub Medical College Abbottabad* **21**: 184–85.
- Rehman M M, Guard-Petter J and Carlson R W.1997. A virulent isolate of *Salmonella* Enteritidis produces a *Salmonella typhi*-like lipopolysaccharide. *Journal of Bacteriology* **179**: 2126–31.
- Utrarachkij F, Pornraungwong S, Siripanichgon K, Nakajima C, Suzuki Y and Suthienkul O. 2011. Possible horizontal transmission of *Salmonella* via reusable egg trays in Thailand. *International Journal of Food Microbiology* **154**: 73–78.