Effects of GandhPaalashi (*Hedychium spicatum*) on the expression of hepatic genes associated with biotransformation, antioxidant and immune systems in WLH cockerels fed indoxacarb

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ABSTRACT

The objective of the present study was to evaluate the efficacy of *Hedychium spicatum* (HSRP) root powder, to ameliorate changes in gene expression in WLH cockerels fed indoxacarb. After 16 weeks of treatment, all birds from different groups were sacrificed humanly and collected the liver pieces in RNA later for expression study by RT-PCR to evaluate changes in the expression of genes involved in antioxidant function [catalase (CAT), superoxide dismutase (SOD)], glutathione peroxidase (GPx), glutathione S-transferase (GST)], biotransformation [cytochrome P450 1A1 (CYP1A1)], and the immune system [interleukins 6 (IL-6)]. Changes in gene expression were determined using the quantitative real-time PCR technique. There were significant upregulation of IL-6, CYP1A1 and GPx genes in indoxacarb treated groups. There was significant down regulation of CAT and SOD genes in indoxacarb treated groups in comparison to control. Simultaneous treatments with HSRP produce ameliorative effect and restore the gene activities at par with control. The current study demonstrates protective effects of HSRP on changes in expression of antioxidant, biotransformation, and immune system genes in cockerels fed indoxacarb.

Key words: Antioxidant, Biotransformation, Cockerels, GandhPaalashi, *Hedychium spicatum*, Hepatic genes, Indoxacarb, Immune system
standard management practices. Experimental cockerels were randomly divided into seven groups of 7 birds each and were kept for 15 day for adaptation. Feed and drinking (tap) water were given ad lib. during the study.

Present study was conducted after approval by the Institutional Ethical Committee (IAEC) vide approval number IAEC/VPT/CVAc/193 dated. 27.02.2015.

Indoxacarb (Avaunt 15.8% EC) used in the study was purchased from local market. Chronic toxicities were induced in WLH cockerels by indoxacarb @ 25 mg/kg body weight. The dose of indoxacarb was selected on the basis of LD₅₀ in cockerels (250 mg/kg). The experimental plan is detailed in Table 1.

Expression of genes by RT-PCR

Collection of liver tissue: After 16 weeks of trial, each birds was sacrificed and immediately a small portion (about 2–3 g) of liver tissue was collected and kept in an RNA later® (Thermo Fisher Scientific). After collection of sample in 2 ml ependorff tube and to assure that the tissue cuts were properly dipped in RNA later and kept –20°C for further use.

RNA isolation and cDNA synthesis: RNA was isolated from liver tissues of birds by Trizol method. The liver tissues from each bird were grinded/ cut into small pieces and 700 µl Trizol reagent was added (Thermo Fisher Scientific) and homogenized (Omni, International, USA). The homogenates were incubated for 5 min. Chloroform (200 µl) was added and mixed by vortexing. The tubes were incubated for 5–10 min at room temperature. Then the samples were centrifuged at 1,200 rpm for 15 min at 4°C. The different phases were separated and then, aqueous phases were removed by angling the tube at 4°C and pipetting these solutions into the new tubes. Isopropanol (500 µl) was added in the aqueous phase and mixed by inverting the tube in 5–10 times. Tubes were incubated for 10 min at room temperature and centrifuged the samples at 10,000 rpm for 5 min at 4°C. The supernatant were removed carefully and only saving the RNA pellets. The RNA pellets were washed with 1 ml of 75% ethanol by mixing it and further centrifuged the samples at 10,000 rpm for 15 min at 4°C. The supernatant were removed. The RNA pellets were air dried for 10 min. The RNA pellets were resuspended in 25 µl RNSase free water and incubated in water bath at 55–60°C for 10 min. The purity of the RNA was assessed by Nanodrop measuring the absorbance of RNA solution at 260 nm and 280 nm. The RNA samples showing the OD₂₆₀:OD₂₈₀ value in between 1.9–2.2 were considered as good quality and used for further analysis. The Reverse transcription of total RNA was carried out using Quanti Tect Reverse Transcription Kit for cDNA synthesis according to the manufacturer’s instructions. The cDNA product was stored at –20°C. Real time RT-PCR was performed using ABI Prism 7500 real time machine (Applied Biosystem). GAPDH was used as an endogenous control. The primer sequences of genes used in the study are given in Table 2.

All the samples were run in triplicates. The amplification was carried out in 20 ml volume reaction mixture containing 10 ml of 2× master mix (Qiagen SYBR Green qPCR Master Mix), 1 ml (10 pmol) each of gene specific forward and reverse primer, 2 ml cDNA template and 6 ml nuclease free water. Negative control was included for the RT-qPCR assay. In negative control (NTC), cDNA was not added. The thermal profile used was as follows: 50°C, 2 min; 95°C for 15 min then 40 cycles of 95°C for 15 sec, AT for 30 sec, and 60°C for 1 min with fluorescence recording at the end of each cycle, followed by denaturation of products from

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of birds</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I (Control)</td>
<td>7</td>
<td>Standard ration ad lib.</td>
</tr>
<tr>
<td>Group II</td>
<td>7</td>
<td>Indoxacarb (250 ppm) in feed</td>
</tr>
<tr>
<td>Group III</td>
<td>7</td>
<td>Silymarin (250 ppm) in feed</td>
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<tr>
<td>Group IV</td>
<td>7</td>
<td>Indoxacarb (250 ppm) + Silymarin (250 ppm) in feed</td>
</tr>
<tr>
<td>Group V</td>
<td>7</td>
<td><em>Hedychium spicatum</em> rhizome powder-(HSRP) @ 4000 ppm in feed</td>
</tr>
<tr>
<td>Group VI</td>
<td>7</td>
<td>HSRP @ 2000 ppm + indoxacarb @ 250 ppm in feed</td>
</tr>
<tr>
<td>Group VII</td>
<td>7</td>
<td>HSRP @ 4000 ppm + indoxacarb @ 250 ppm in feed</td>
</tr>
</tbody>
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<thead>
<tr>
<th>Name</th>
<th>Symbol</th>
<th>Primer Product size</th>
</tr>
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<tbody>
<tr>
<td>Catalase</td>
<td>CAT</td>
<td>GGGGAGCTGTTTACTGCAA-F 139 bp</td>
</tr>
<tr>
<td>Cytochrome P450A1</td>
<td>CYP1A1</td>
<td>TTTCCATTGGCTATGGCATT-R 125 bp</td>
</tr>
<tr>
<td>Glutathione peroxidase</td>
<td>GPx</td>
<td>CACTTTCTGCCGCTCCTG-F 140 bp</td>
</tr>
<tr>
<td>Superoxide dismutase</td>
<td>SOD</td>
<td>GTGCCCTTCCTCACGCTCCAG-R 122 bp</td>
</tr>
<tr>
<td>Glyceraldehyde-3 phosphate dehydrogenase</td>
<td>GAPDH</td>
<td>TTGTAAACATCAGGGGCCAA-F 128 bp</td>
</tr>
<tr>
<td>Interleukin-6</td>
<td>IL-6</td>
<td>CAACATCAAA TGGGCAGATG-R 128 bp</td>
</tr>
</tbody>
</table>

Table 2. Real time based analysis of transcript abundance
65°C to 95°C with fluorescence recording throughout the step.

Statistical analysis: Expression level of genes were analyzed in six ways—control versus indoxacarb @ 250 ppm; control versus silymarin @ 250 ppm; control versus indoxacarb @ 250 ppm + silymarin @ 250 ppm; control versus HSRP @ 4000 ppm; control versus HSRP @ 2000 ppm + indoxacarb @ 250 ppm and control versus HSRP @ 2000 ppm + indoxacarb @ 250 ppm. The relative expression of each sample was calculated using the $2^{-\Delta\Delta CT}$ method with control group as calibrator (Schmittgen and Livak 2008). One way ANOVA was done in JMP9 (SAS Institute Inc, Cary, USA) and differences between groups were considered significant at P<0.05.

RESULTS AND DISCUSSION

The expression of hepatic genes involved in antioxidant function [catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx)] is depicted in Figs 1, 2, and 3 respectively. The expression of biotransformation [cytochrome P450 1A1] and the immune system [interleukin 6 (IL-6)] genes is listed in Fig. 4 and 5 respectively.

Antioxidant gene expression: Antioxidant enzymes catalase (CAT) present within the peroxisomes and cytosolic GPx are involved in the conversion of hydrogen peroxide (a powerful and potentially harmful oxidizing agent) into water and molecular oxygen (Liska 1998). Hydrogen peroxide is unstable and forms hydroxyl free radicals; its break-down on the inner mitochondrial matrix and within the mitochondrial matrix targets. The decomposition of hydrogen peroxide outside the mitochondria may cause damage to the outer mitochondrial membrane itself which suggests that the breakdown of hydrogen peroxide to the hydroxyl radical is highly detrimental.

Compared with controls, the expression of the CAT gene was significantly (P<0.05) down regulated in indoxacarb treated group I and also down regulated in group III treated with silymarin + indoxacarb (Fig. 1). Down regulation in indoxacarb + HSRP high dose group VI was down regulated but it was four fold less than the group I treated with indoxacarb. This, finding suggested that the toxicity of indoxacarb suppressed the gene expression and lowered level of suppression in group III and VI which might be due to the ameliorating effect of silymarin and HSRP by the presence of antioxidants available for neutralizing the ROS/RNS generated inside body. In group II and IV treated with silymarin and HSRP, the expression was nonsignificantly up regulated in comparison to control. The present study on CAT in RBC and tissues is also in agreement with the CAT gene expression in liver tissue as CAT enzyme activity was also decreased in indoxacarb treated group (Choudhary and Singh 2017). This result was in agreement with findings of aflatoxin toxicity in broiler chick causing down regulation of CAT gene expression (Yarru et al. 2009).

Superoxide dismutase catalyses the conversion of superoxide anions to hydrogen peroxides and is one of the primary enzymatic defence system against ROS. The hepatic gene expression of SOD in comparison to control was significantly (P<0.05) down regulated in group I, III, V and VI (Fig. 2). The indoxacarb treated group I showed suppression of SOD genes which was 4/3 fold more than the group III and VI. Group II and IV showed up regulation of SOD gene in comparison to control. This finding suggest that the SOD gene was suppressed by indoxacarb and caused down regulation due to the toxicity produced by the indoxacarb but simultaneous feeding of silymarin in group III and HSRP in group V and VI ameliorated the toxicity produced by the indoxacarb and led to up regulation in V and VI groups in comparison to group I down regulation.

Glutathione peroxidase gene expression was significantly up regulated in indoxacarb treated group in comparison to control (Fig. 3). This finding also corroborated with finding of Choudhary and Singh (2017). In group II and group IV, GPx gene was down regulated that might be due to antioxidant action of silymarin and HSRP which combat the ROS/ RNS generated by the indoxacarb. In group III, V and VI, the GPx gene was up regulated in comparison to control but less than the Group I treated with indoxacarb. This might be due to amelioration by simultaneous feeding of silymarin and HSRP in dose dependent manner which directly or indirectly influenced the GPx gene expression.

Expression of biotransformation genes (CYP1A1): The main hepatic transformation processes include xenobiotics biotransformation through phase I metabolism and further by the conjugation of the resulting metabolites to convert them into more water soluble conjugates which are rapidly readily removed from the body. The microsomal cytochrome P450 (CYP)- dependent monoxygenase system in the liver plays an essential role in phase I metabolism (Akahori et al. 2009). The CYP enzymes are associated with several biological interaction involving
hydroxylation, epoxidation, oxygenation, dehydrogenation, nitrogen dealkylation and oxidative deamination (Bruton et al. 2011). Cytochrome P450 mediated reactions can also generate ROS. The CYP enzymes are involved in hepatic metabolism of indoxacarb in poultry and other enzymes like AST, ALT, GGT and ALP were elevated in indoxacarb treated poultry and feeding of *H. spicatum* root powder ameliorated the enzymic activities towards normal (Choudhary and Singh 2017).

The hepatic CYP1A1 gene was up regulated in indoxacarb treated group in comparison to control (Fig. 4). Overexpression of these CYP isoforms has been shown to induce chronic oxidative stress by generating more ROS/RNS, leading to hepatocellular injury (Kumar and Kuttan 2006). In group II and IV treated with silymarin and HSRP alone, the CYP1A1 gene was down regulated which might be due to suppression of the gene but in group III, V and VI treated with indoxacarb+silymarin and HSRP+indoxacarb with low and high doses, CYP1A1 was up regulated in comparison to control but less than the indoxacarb treated group, which indicated the ameliorative effect of silymarin and HSRP in dose dependent manner. The overexpression of the CYP1A1 gene was also reported by Kumar and Kuttan (2006) following aflatoxin toxicity in broiler chick.

Expression of cytokine gene (IL-6): Interleukins form a group of cytokines that are important components of the immune system. They play vital role and are now well recognised in inflammation and a pathological condition in systemic inflammatory states. Any imbalance in cytokine production and dysregulation in circulating cytokines could result into various pathological disorders (Tayaland Kalra 2007).

The IL-6 is a proinflammatory cytokine. The IL-6 gene was significantly (P<0.05) upregulated in indoxacarb treated group in comparison to control. This might be due to the excessive generation of free radicals which damages the cells leading to release of IL-6 and over expression of IL-6 gene (Fig. 5). IL-6 gene was down regulated in group II and IV treated with silymarin and HSRP. In group III, V and VI, the IL-6 genes was significantly (P<0.05) upregulated in comparison to the control but was less than the indoxacarb treated groups. Reduction in up regulation of IL-6 in group V and VI might be due to scavenging the free radicals by HSRP in dose dependent manner by its anti-inflammatory properties (Kunnumakkara et al. 2008).

Indoxacarb damages the spleen showed moderate depletion of lymphoid cell from white pulp, vacuolation of the tunica media/tunica muscularis of the many large and small blood vessels of arterial side; hemosiderosis were also seen noticed.
but simultaneous feeding of *H. spicatum* rhizome powder the lesion become mild to moderate (Choudhary et al. 2017).

Thus it is concluded from the present investigation that the *Hedychium spicatum* produces amelioration on long term intoxication with indoxacarb in WLH cockerel by the up regulation and down regulation of different hepatic genes involved in antioxidant function [catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx)], biotransformation [cytochrome P450 1A1] and the immune system [interleukin 6 (IL-6)] in cockerels fed indoxacarb.

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REFERENCES


Fig. 5. Log2 fold changes in gene expression of IL-6 gene following administration of HSRP in feed for 16 weeks in indoxacarb treated WLH cockerels.