



## Note

# DNA barcoding of selected species of pufferfishes (Order: Tetraodontiformes) of Puducherry coastal waters along south-east coast of India

K. KALESHKUMAR, R. RAJARAM, S. VINOTHKUMAR, V. RAMALINGAM AND KHANGEMBAM BRAJAMANI MEETEI\*

School of Marine Sciences, Bharathidasan University, Tiruchirappalli - 620 024, Tamil Nadu, India

\*Krishi Vigyan Kendra, Bishnupur District, Utlou P. O., Nambol, Manipur - 795 134, India

e-mail: drrajaram69@rediffmail.com

## ABSTRACT

Samples of three species of pufferfishes were collected from Puducherry in the south-east coast of India. Collected fishes were identified as *Chilomycterus reticulatus*, *Arothron hispidus* and *Lagocephalus guentheri* by mitochondrial CO1 gene sequencing. Neighbor-Joining method was used for phylogenetic analysis which confirmed that the analysed species had a dichotomous relationship with its ancestor species. Genetic variation was studied by sequence similarity using the homologous sequences retrieved from BLAST server. The maximum length of open reading frames (ORF) for the three pufferfish species viz., *C. reticulatus*, *A. hispidus* and *L. guentheri* were 171, 345 and 465 respectively. Results from a simple gene prediction algorithm revealed that *C. reticulatus*, *A. hispidus* and *L. guentheri* have 7 start codons each. The pairwise genetic distance calculated among the three species ranged from 0.002 to 0.176% for *C. reticulatus*, 0.002 to 0.169% for *A. hispidus* and 0.002 to 0.208% for *L. guentheri*. Genetic distance analysis of mitochondrial DNA sequences is demonstrated as unique to their related species of all pufferfishes. Genetic distance is also used in the comparison of genetic similarity between different species and also in different subspecies of pufferfishes belonging to the order Tetraodontiformes.

Keywords: Genetic distance analysis, Mitochondrial CO1 sequencing, Phylogenetic analysis, Pufferfishes, Tetraodontiformes

*In silico* studies can identify the genetic diversity and enable fast prediction of gene expression based on sequence comparison within species. Genetic materials are the major source to reveal intra-specific genetic diversity and phenotypic plasticity from intra-specific differences for all kinds of organisms (Fabrice *et al.*, 2010). DNA - based identification techniques have been proved to be powerful analytical tools (Ko *et al.*, 2013). Morphological information such as body shape, pigmentation and meristic count are inadequate for genus or species level study in fish (Zhang *et al.*, 2004; Comi *et al.*, 2005; Teletchea, 2009). Hence, molecular level information becomes important for ecological monitoring, environmental impact assessment, formulating fisheries compensation plans, resource management, smuggling prevention and establishing marine protected areas (Moura *et al.*, 2008; Valdez-Moreno *et al.*, 2010). Presently, DNA barcoding is the standardised and universal method to identify species and in uncovering biological diversity (Hebert *et al.*, 2003; 2004).

Mitochondrial DNA (mt DNA) has proven to be a useful molecular marker for evolutionary studies in genetic diversity, because of its predominantly maternal inheritance, relatively rapid base substitution rate, lack of

recombination and easy isolation (Avisé *et al.*, 1987). In particular, the mitochondrial COI gene, can be amplified from most living beings using universal primers and has been proposed as a reference marker for cross - amplification over a wide range of species, genera and families (Hebert *et al.*, 2003). The evolution rate of mt DNA and its amino acid sequence is highly conserved to COI gene across the phyla that have been used in various molecular tools in several genetic studies. Finally, it has been proposed as a universal barcode database for all animal species (<http://www.barcodinglife.com>) and has already been used as an identifier for numerous insect taxa (Hogg and Hebert, 2004; Hufbauer *et al.*, 2004). Ajmal Khan *et al.* (2011) barcoded 42 fishes belonging to 32 genera and 23 families while Lakra *et al.* (2011) barcoded 115 marine finfish species belonging to 37 families and 79 genera, from Indian waters.

Pufferfishes are appropriate group to study the process of genome size evolution and in particular, smooth pufferfishes (family Tetraodontidae) are the smallest vertebrates with a haploid genome size of ~400 million bp (Neafsey and Palumbi, 2003). Nevertheless, the genetic relationship of scaffold is still unknown owing to the requirement of genetic map of the fish. The use

of pufferfish gene sequences for comparative study with other related species can provide the answer for genome structure and evolution (Kai *et al.*, 2003). In this study, we have sequenced and reported the genetic diversity based on sequence information, open reading frames (functional region) and also identified the beginning (start codon) and termination region (polyadenylation signals) of functional mitochondrial COI gene for three pufferfish species *viz.*, *Chilomycterus reticulatus*, *Arothron hispidus* and *Lagocephalus guentheri* belonging to the order Tetraodontiformes from Puducherry coastal region along the south-east coast of India.

Pufferfish samples were collected from Puducherry (11°93'N, 79°83'E) and the collected fishes were transported to the laboratory and preserved at -20°C until further analysis. Fishes were identified based on morphological features and tissue samples from the muscle portion of the fishes were dissected and preserved in 95% ethanol for DNA isolation. The mitochondrial DNA was extracted from the tissue following the method of Sambrook *et al.* (2001). DNA amplification was carried out using the specific primer; Forward - FISH F1 (5'- TCAACCAACCACAAAGA CATTGGCAC-3') and Reverse - FISH R1 (5'-TCGACT AATCATAAAGATATCGGCAC-3') (Ward *et al.*, 2005). The PCR conditions used were: denaturation step of 95°C for 1 min, followed by 35 cycles of 30 s at 95°C, 30 s at 40°C, and 90 s at 72°C, followed by 10 min at 72°C. PCR products were then purified using Ultra Clean PCR clean up (Mo-Bio) spin columns, and then submitted to Eurofins Genomics India, Bangalore, for sequencing using ABI3730XL (Applied Biosystems). Sequences were verified and manipulated with Sequencer ver. 4.8 (Gene Codes Corporation). Mitochondrial COI gene from the three fish species *viz.*, *C. reticulatus*, *A. hispidus* and *L. guentheri* were sequenced and submitted to Barcode database (with accession nos. JX495771, KF422720 and KF442241 respectively) and the length of the COI regions were 631, 636 and 615 bp respectively.

DNA sequences were initially aligned using CLUSTAL X (Thompson *et al.*, 1994) and default gap, extension penalties were used followed by manual editing using Seq App 1.9 (Gilbert, 1994). Alignments were adjusted manually following guidance of Odorico and Miller (1997) and Chen *et al.* (2004). A BLAST search was performed in GenBank database for each sequence and the matching homologous sequences were retained for subsequent alignment. Neighbor-Joining method was implemented in MEGA v 5.0 (Kumar *et al.*, 2001). BLASTn and BLASTx searches of the NCBI non-redundant protein database were performed for all processed sequences and hits with

Expect values (E values) of less than  $1 \times 10^{-4}$  were considered to be homologs to the query sequence. For the differentially expressed sequences, BLASTn searches of the non-human, non-mouse EST database were performed for all sequences while homologs with significant E-values from the BLASTn search were then used as a query in a BLASTx search and the open reading frames and start codons were predicted (Ramalingam *et al.*, 2012). Genetic distance calculations implemented in MEGA version were used to estimate sequence divergence (Kumar *et al.*, 2001).

In the present study, the nucleotide sequences of three pufferfish species belonging to the families Diodontidae and Tetraodontidae (Order: Tetraodontiformes) *viz.*, *Chilomycterus reticulatus*, *Arothron hispidus* and *Lagocephalus guentheri* were initially compared with nucleotide database using BLASTn server which indicated that these organisms acquired maximum identity as compared with other genus. *C. reticulatus* was recorded for the first time in Indian waters and also the sequence information happens to be the first submission in barcode database. Details of barcode submissions of *C. reticulatus*, *A. hispidus* and *L. guentheri* in barcode database are presented in Table 1.

Table 1. List of barcoded species available in BOLD

Name	Family	Accession no.	No. of submissions available*
<i>Chilomycterus reticulatus</i>	Diodontidae	JX495771	1
<i>Arothron hispidus</i>	Tetraodontidae	KF422720	30
<i>Lagocephalus guentheri</i>	Tetraodontidae	KF442241	13

\*Number of submission in barcode database

The base of tree forms a dichotomy leading to two clusters *i.e.*, clade I and clade II (Fig. 1). Clade I consists of two sub clades that carries both *A. hispidus* and *L. guentheri* (Family: Tetraodontidae) while clade II consists of *C. reticulatus* (Family: Diodontidae). The branch length leading to *A. hispidus* clade was similar to the branch length leading to the related species of *A. hispidus*. Similarly, *A. hispidus* and *L. guentheri* clade was highly similar with their respective related species which suggests that *A. hispidus* and *L. guentheri* have unique heterogeneity of mt DNA. The different strains of the species *L. guentheri* are closely related (Santini *et al.*, 2013). Holgroft (2005) reported that the mitochondrial sequences of Balistidae and Monacanthidae are sister groups related to Tetraodontidae and Diodontidae that robustly maintain the monophyletic relation.

A blast search revealed that the segment of control gene of mt COI, has 99% sequence similarity in *C. reticulatus*, *A. hispidus* and *L. guentheri*. The nucleotide and protein sequences of *C. reticulatus*, *A. hispidus* and

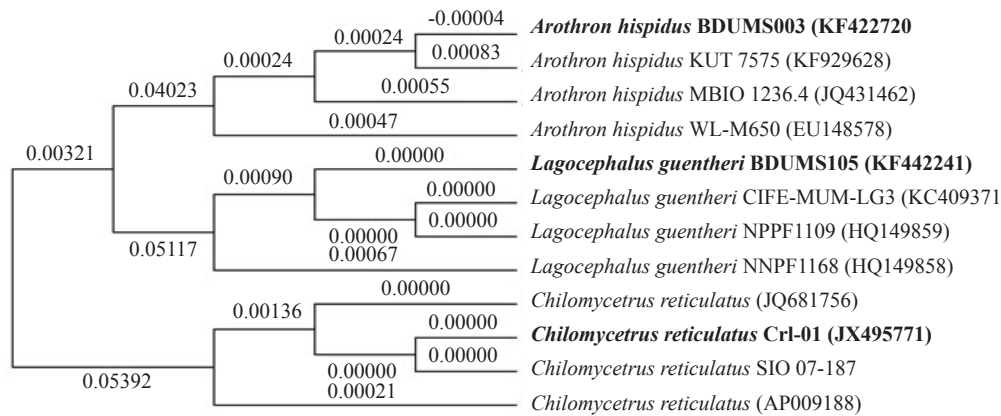


Fig. 1. Phylogenetic analysis of *C. reticulatus* and *L. guentheri* by Neighbor joining method using MEGA v 5.0

*L. guentheri* were primarily compared and analysed with relevant databases using BLASTn and BLASTx as shown in Table 2, 3 and 4. Maximum identity was observed in different strains of *C. reticulatus* nucleotide sequences (Table 2). Maximum identity of *A. hispidus* was with the nucleotide sequence of *A. hispidus* WL-M650 and *A. hispidus* WL-M651 (Table 3). Maximum identity of *L. guentheri* was observed with the nucleotide sequence of *L. guentheri* CIFE-MUM-LG3 and *L. guentheri* Smith 268.19 (Table 4).

The mt COI sequence of *C. reticulatus* and *A. hispidus* contains four open reading frames (ORFs) and *L. guentheri* has three ORFs. In *C. reticulatus*, the long ORF was obtained between 466-636 bp and 165-335 bp length while in *A. hispidus* the long ORF was between 3 and 347 bp and *L. guentheri* has long ORF between 79 and 543 bp length. The long ORF present in the pufferfishes can encode a functional protein in the cytochrome oxidase I gene. Yamaguchi and Brenner (1997) reported that the F1A $\alpha$  has an intronless 1.3 kb

Table 2. Annotated maximum sequence identity of nucleotide and protein for *Chilomycterus reticulatus* with related similar species using Blast n and Blast x server

Species	Blast N			Blast X		
	Accession no.	Max. score	Max identity	Accession no.	Max score	Max identity
<i>C. reticulatus</i>	JQ681756.1	1125	99%	AGZ84817.1	289	99%
<i>C. reticulatus</i>	AP009188.1	1120	99%	ACG50200.1	290	99%
<i>C. reticulatus</i> SIO 07-187	HQ010089.1	1016	99%	AEK04469.1	290	99%
<i>C. schoepfi</i>	JQ681758.1	704	87%	AFJ92749.1	290	99%
<i>C. geometricus</i>	JQ681755.1	693	87%	AFJ92700.1	290	99%

Table 3. Annotated maximum sequence identity of nucleotide and protein for *Arothron hispidus* with related similar species using Blast n and Blast x server

Species	Blast N			Blast X		
	Accession no.	Max score	Max identity	Accession no.	Max score	Max identity
<i>A. hispidus</i>	FJ434547.1	1171	100%	ACG50200.1	414	99%
<i>A. hispidus</i>	JQ681763.1	1157	100%	ACG50207.1	413	99%
<i>A. hispidus</i> WL-M651	EU148579.1	1133	100%	AEV56399.1	414	100%
<i>A. hispidus</i> NBE 0677	JQ349778.1	1129	100%	ABO48435.1	413	100%
<i>A. hispidus</i> WL-M650	EU148578.1	1127	99%	ACG50203.1	413	100%

Table 4. Annotated maximum sequence identity of nucleotide and protein for *Lagocephalus guentheri* with related similar species using Blast n and Blast x server

Species	Blast N			Blast X		
	Accession no.	Max score	Max identity	Accession no.	Max score	Max identity
<i>L. guentheri</i> BDUDMS 105	KF442241.1	1136	100%	AGZ92978.1	400	100%
<i>L. guentheri</i> CIFE-MUM-LG3	KC409371.1	1136	100%	AGC25469.1	400	100%
<i>L. guentheri</i> NPPF1109	HQ149859.1	1131	99%	ACM88301.1	399	99%
<i>L. guentheri</i> NPPF 1168	GQ149858.1	1120	99%	AEK02887.1	398	99%
<i>L. guentheri</i> Smith 268.19#3	JF493722.1	1114	99%	ACR44799.1	398	99%

open reading frame encoding a 423 aa protein. F1A $\beta$  has an intronless 1.4 kb ORF encoding a 416 aa protein.

*C. reticulatus* contains two poly A signals at the nucleotide positions of 421 and 447 while *A. hispidus* has three poly A signals at positions of 201, 425 and 561 whereas *L. guentheri* contains three poly A signal at 211, 433 and 565. The accurate prediction of polyadenylation sites play an important role in understanding transcriptome diversity and regulatory mechanisms of gene expression by defining the termination of genes (Wu *et al.*, 2012). The pairwise genetic distance computed among the three species ranged from 0.002 to 0.176% for *C. reticulatus*, 0.002 to 0.169% for *A. hispidus* and 0.002 to 0.208% for *L. guentheri* (Table 5, 6 and 7). The lowest genetic distance indicates closely related and highest genetic distance indicates highly diverged status.

Table 5. Genetic distance of *C. reticulatus* sequence from related species

Species	1	2	3	4	5	6	7	8
<i>Diodon holocanthus</i> NSMK:P1-000250								
<i>Diodon holocanthus</i> MFC233	<b>0.002</b>							
<i>Diodon holocanthus</i> BIOUG:HLC-15165	<b>0.004</b>	<b>0.005</b>						
<i>Diodon hystrix</i> MBI0319	<b>0.107</b>	<b>0.109</b>	<b>0.104</b>					
<i>Cylichthys orbicularis</i> ADC09_269.2	<b>0.138</b>	<b>0.141</b>	<b>0.136</b>	<b>0.152</b>				
<i>Chilomycterus reticulatus</i> SIO 07-187	<b>0.112</b>	<b>0.115</b>	<b>0.110</b>	<b>0.141</b>	<b>0.150</b>			
<i>Chilomycterus reticulatus</i> BDUMS001	0.115	0.117	0.112	0.143	0.152	<b>0.002</b>		
<i>Chilomycterus spinosus</i> LBP-35009	<b>0.124</b>	<b>0.126</b>	<b>0.126</b>	<b>0.176</b>	<b>0.158</b>	<b>0.117</b>	<b>0.120</b>	

Table 6. Genetic distance of *A. hispidus* sequence from related species

Species	1	2	3	4	5	6	7	8
<i>Arothron immaculatus</i> NBFGR:1102b								
<i>Arothron nigropunctatus</i> BLOUG:HLC-12417	0.075							
<i>Arothron stellatus</i> CIFE-MUM-AS2	0.077	0.079						
<i>Arothron hispidus</i> BDUMS003	0.072	0.083	0.052					
<i>Arothron hispidus</i> CIFE-MUM-AH1	0.081	0.089	0.053	0.012				
<i>Arothron hispidus</i> MBIO1236.4	0.074	0.085	0.053	<b>0.002</b>	0.013			
<i>Canthigaster solandri</i> MBIO949.4	0.169	0.165	0.154	0.144	0.154	0.142		
<i>Canthigaster janthinoptera</i> ECOMAR:NBE1142	0.167	0.165	0.154	0.144	0.154	0.142	0.032	

Table 7. Genetic distance of *L. guentheri* sequence from related species

Species	1	2	3	4	5	6	7	8	9
<i>Sphoeroides karkudineus</i> LBP-41589									
<i>Sphoeroides naphelus</i> SMSA7320	0.133								
<i>Lagocephalus wheeleri</i> SIFU1	0.174	0.151							
<i>Lagocephalus spadiceus</i> FSCS302-00	0.177	0.147	0.012						
<i>Lagocephalus guentheri</i> BDUMS006	0.183	0.170	0.077	0.075					
<i>Lagocephalus guentheri</i> NPPF1168	0.181	0.168	0.075	0.073	<b>0.002</b>				
<i>Dactyloptera orientalis</i> WL-M573	0.174	0.166	0.071	0.069	0.008	0.007			
<i>Lagocephalus larigatus</i> HRCP:46919	0.183	0.165	0.108	0.106	0.103	0.102	0.096		
<i>Myripristis adusta</i> MBIO1182-4	0.185	0.208	0.184	0.182	0.184	0.182	0.177	0.214	

The lowest genetic distance is highlighted and the farthest genetic distance is marked in bold

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Genetic distance analysis of pufferfishes showed close identity with its related species and subspecies. Fish genomes are closely related with human genomes that have potential to identify genes and regulatory regions in humans. Further, pufferfishes especially of the family Tetraodontidae is an important group for studying the phenotypic variation.

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