



Role of *Lactobacillus plantarum*, dextrose and starch on the storage stability and quality attributes of fermented chicken sausages

GEETA PIPALIYA AND AJIT S. YADAV*

ICAR- Central Avian Research Institute, Izatnagar-243122, Uttar Pradesh, India.

(Received on September 25, 2023; accepted for publication on January 03, 2024)

ABSTRACT

Pipaliya, G. and Yadav, A.S. 2024. Role of *Lactobacillus plantarum*, dextrose and starch on the storage stability and quality attributes of fermented chicken sausages. *Indian Journal of Poultry Science*, 59(1): 69-74.

This study aimed to investigate the influence of starter culture, *Lactobacillus plantarum* and substrate, specifically dextrose and starch, on the storage stability of fermented chicken sausages under both refrigerated ($4\pm1^{\circ}\text{C}$) and frozen ($-20\pm1^{\circ}\text{C}$) conditions. In this study, two groups of sausages were analyzed: T_1 group fermented with *L. plantarum* (10^8cfu/g) and T_2 group fermented with *L. plantarum* (10^8cfu/g) in combination with 1% each of dextrose and starch. The sausages from both groups were evaluated at regular intervals: 0, 3, 5, 7, 9 and 12 days for refrigerated storage and 0, 15, 30, 45 and 60 days for frozen storage. Throughout the storage period, T_2 group exhibited higher antioxidant potential compared to the T_1 group, whether stored in refrigerated or frozen conditions. Based on a comprehensive assessment of physicochemical, antioxidant, microbiological and sensory parameters, both groups of fermented chicken sausages remained safe for consumption up to 12 days under refrigerated storage and up to 60 days under frozen storage. This research study demonstrated that, the incorporation of dextrose and starch, together with *L. plantarum* (10^8cfu/g), enhances the antioxidant properties and overall value of fermented chicken sausages, extending their shelf life under refrigerated and frozen storage conditions.

Keywords: *Lactobacillus plantarum*, Dextrose, Starch, Storage stability

INTRODUCTION

Fermented sausage, with its distinct flavor and global popularity, stands as an appreciated meat product amongst consumers (Liu *et al.*, 2021). Traditionally enjoyed at room temperature, its convenience and palatable taste have made it a staple in many diets worldwide. However, the journey from processing to storage presents a numerous of safety challenges, together with the fear of oxidative rancidity, the presence of pathogenic bacteria, and an surplus of biological amines (Luan *et al.*, 2021). These issues underscore the need for inventive approaches to enhance the storage stability and overall quality of fermented chicken sausages.

In the 21st century, as nutrition trends gradually drift towards the pursuit of optimal nutrition, researchers and professionals have turned their consideration to reformulating and incorporating functional ingredients to side with evolving consumer perceptions of food healthiness (Kaur and Sharma, 2019). This growing demand for original and health-conscious food products has ignited a force to create novel fermented meat products with better nutritional profiles and health benefits (Agüero *et al.*, 2020). Therefore, food technologists are now embarking on efforts to develop meat and meat-based products that not only satisfy culinary cravings but also contribute to overall well-being by replacing or increasing specific components (Fernández-López *et al.*, 2021). In this framework, the roles of *Lactobacillus plantarum*, dextrose, and starch in the storage stability and quality attributes of fermented chicken sausages grasp

significant potential offering a glance into the future of meat processing and its efficiency to provide the growing dietary requirements of consumers. In this study, our primary purpose was to explore the impact of a starter culture, *L. plantarum* and specific substrates, namely dextrose and starch, on the storage stability of fermented chicken sausages, subject to both refrigerated and frozen conditions. This study serves as a crucial step toward understanding the role of these components in maintaining the quality and stability of fermented chicken sausages, contributing valuable insights for both consumers and the food industry.

MATERIALS AND METHODS

The starter culture, namely *L. plantarum* (NCDC No. 020) was procured from ICAR-National Dairy Research Institute, Karnal, in a freeze-dried, vacuum-sealed form. The initial stock culture was revived following the manufacturer's instructions, with sub-culturing at fortnight intervals to maintain the mother culture. *L. plantarum* was then sub-cultured in duplicate in *Lactobacillus* MRS Broth, harvested and adjusted to a concentration of 10^{10} bacterial cells/ml using spectrophotometry. For minced meat preparation, White Leghorn spent hens were sourced from ICAR-Central Avian Research Institute, Izatnagar, and processed through fasting, bleeding, scalding, defeathering, and evisceration before being transported to the laboratory for deboning.

Hot dressed carcasses were deboned without delay and the minced meat was then subjected to a 24-hour

*Corresponding author Email: asyadav19@rediffmail.com

fermentation process with *L. plantarum* (10^8 cfu/g) for T_1 group and *L. plantarum* (10^8 cfu/g) and additional substrates (i.e. 1% each of dextrose and starch) for T_2 group. The fermentation was done in a controlled fermentation chamber set at an optimized temperature of 18-20°C with a relative humidity of 90±5%. The resulting fermented minced chicken meat was used for emulsion formulation in both groups.

The emulsion for chicken sausages was prepared by mixing fermented minced lean meat with ingredients such as common salt, sodium nitrite, sodium nitrate, natural fat, condiment mix, spice mix, and refined wheat flour. Care was taken to keep the emulsion temperature below 10°C with the use of ice cubes. After preparation, the emulsion was refrigerated at 4±1°C. Using sausage filler, the emulsion was transferred into casings, linked for uniform sizing and then cooked in boiling water until getting an internal temperature of 72°C. After cooking, the sausages were cooled to room temperature. Samples were initially collected from both groups and underwent comprehensive analysis on day 0, including assessments of antioxidant activity (ABTS, SASA and DPPH radical scavenging), TBA, pH, free fatty acids (FFA), microbiological and sensory evaluation. Subsequently, these samples were stored under refrigerated and frozen conditions and re-evaluated at specific intervals: 3, 5, 7, 9 and 12 days for refrigerated storage and 15, 30, 45 and 60 days for frozen storage.

In the estimation of antioxidant parameters, we followed well-known methodologies, utilizing the approaches of Shirwaikar *et al.* (2006) for ABTS, Kumar and Chattopadhyay (2007) for SASA and Kato *et al.* (1988) for DPPH radical scavenging activity. Thiobarbituric acid (TBA) analysis was conducted according to the method outlined by Tarladgis *et al.* (1960). Physicochemical parameter such as pH was measured in alliance with the methodology described by Keller *et al.* (1974). The quantification of free fatty acids (FFA) was carried out as described by Koniecko (1979). Microbiological parameters were assessed in strict accordance with the standard methods detailed by Speck (1992). Moreover, sensory evaluation was performed using an 8-point hedonic scale to gauge the overall sensory attributes of the samples. The data obtained in this study were analyzed using SPSS software (Version 21; IBM SPSS, 2012). One-way analysis of variance (ANOVA) and T-test were conducted for all the parameters. Statistical significance was determined at the $P < 0.05$ level and the means were separated using the Duncan multiple range test (Duncan, 1955).

RESULTS AND DISCUSSION

Antioxidant parameters

During a 12-day refrigeration and 60-day frozen storage period, the T_2 group consistently exhibited higher

antioxidant activity compared to T_1 , assessed through the ABTS⁺ assay, DPPH, and superoxide anion scavenging activity (Table 1 and 2). A study conducted by Li Shengyu *et al.* (2012) also found impressive hydroxyl radical and DPPH scavenging activities with *L. plantarum* C88. Another study conducted by Kachouri *et al.* (2015) highlighted the capacity of *Lactobacillus* to produce high-value antioxidants from phenolic compounds in olives, while Li Zhongxi *et al.* (2018) demonstrated improved antioxidant capacity when fermenting apple juice with *L. plantarum* ATCC14917. With the significance of oligosaccharide metabolism for the ecological fitness of *Lactobacillus* in different food-related and intestinal environments (Walter, 2008; Tannock *et al.*, 2012), the synergistic role of dextrose and starch, which may be likely owing to the creation of a favorable environment for *Lactobacillus plantarum*, would have contributed to enhanced fermentation ability and antioxidant properties, as evidenced by the increased inhibition activity of *L. plantarum* against ABTS, DPPH, and SASA radicals (Geeta and Yadav, 2017).

Physicochemical parameters

In our study, both groups exhibited a consistent decline in pH during refrigerated and frozen storage, with the lowest pH recorded at the end of each storage phase (Table 3 and 4). This pH reduction is comparable to the findings by Christiansen *et al.* (1975) on summer-style sausage storage and is attributed to lactic acid bacteria multiplying and converting residual carbohydrates into lactic acid. In our study, the T_2 group, enriched with extra sugar substrates during fermentation with *L. plantarum*, exhibited a more significant pH decline throughout storage, likely due to increased lactic acid production with the added sugar substrates (Antara *et al.*, 2004; Ahmad and Shrivastav, 2007).

Throughout the successive refrigerated and frozen storage, there was an increase in the TBA (Thiobarbituric Acid) value observed for both groups (Table 3 and 4). However, this increase was less pronounced during the two-month frozen storage period compared to the twelve-day refrigerated storage period. Furthermore, the TBA values for the T_2 group were a little lower than those for the T_1 group, both during refrigerated storage and frozen storage. The increase in TBA values during refrigerated and frozen storage can be accredited to the oxidation of polyunsaturated fatty acids within the phospholipids fraction, in line with the findings of Igene *et al.* (1980) and Brewer *et al.* (1982). It is significant that the TBA values for both groups remained within acceptable limits (1-2 mg malonaldehyde/kg of meat) during the entire storage period, as specified by Witte *et al.* (1970). Though, previous studies suggest that the functional attributes deteriorate with the extension of the frozen storage time (Smiecinska *et al.*, 2015; Ablikim *et al.*, 2016; Wei *et al.*, 2017). Moreover, the maximum Free

Table 1: Effect of refrigerated storage ($4\pm 1^{\circ}\text{C}$) on antioxidant activity of fermented chicken sausages

Group	Storage Days						Total
	Day 0	Day 3	Day 5	Day 7	Day 9	Day 12	
ABTS (%)							
T ₁	46.09±0.12 ^{aq}	45.52±0.26 ^{aQ}	44.48±0.17 ^{bQ}	42.59±0.02 ^{cQ}	39.65±0.18 ^{dQ}	37.94±0.30 ^{eQ}	42.71±0.73
T ₂	74.25±0.02 ^{ap}	71.99±0.01 ^{bp}	66.61±0.25 ^{cp}	56.96±0.07 ^{dp}	50.34±0.03 ^{ep}	48.98±0.10 ^{fp}	61.52±2.42
SASA (%)							
T ₁	64.59±0.22 ^{aQ}	60.41±0.19 ^{bQ}	50.97±0.04 ^{cQ}	40.85±0.01 ^{dQ}	38.41±0.04 ^{eQ}	36.62±0.06 ^{fQ}	48.64±2.63
T ₂	86.84±0.07 ^{ap}	82.03±0.02 ^{bp}	71.73±0.23 ^{cp}	63.07±1.42 ^{dp}	50.45±0.08 ^{ep}	43.78±0.16 ^{fp}	66.32±3.79
DPPH (%)							
T ₁	26.54±0.20 ^{aQ}	25.13±0.63 ^{bQ}	20.47±0.00 ^{cQ}	18.91±0.02 ^{dQ}	15.90±0.05 ^{eQ}	11.44±0.14 ^{fQ}	19.73±1.25
T ₂	51.77±0.01 ^{ap}	49.90±0.05 ^{bp}	48.45±0.03 ^{cp}	46.23±0.05 ^{dp}	44.19±0.02 ^{ep}	40.16±0.07 ^{fp}	46.78±0.93

Mean $\pm$ S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean $\pm$ S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Table 2: Effect of frozen storage ($-20\pm 1^{\circ}\text{C}$) on antioxidant activity of fermented chicken sausages

Storage Days						
Group	Day 0	Day 15	Day 30	Day 45	Day 60	Total
ABTS (%)						
T ₁	46.09±0.12 ^{aQ}	42.78±0.24 ^{bQ}	39.44±0.23 ^{cQ}	37.52±0.18 ^{dQ}	35.68±0.10 ^{eQ}	40.30±0.99
T ₂	74.25±0.02 ^{aP}	73.62±0.12 ^{bP}	71.24±0.10 ^{cP}	70.46±0.18 ^{dP}	69.43±0.15 ^{eP}	71.80±0.49
SASA (%)						
T ₁	64.59±0.22 ^{aQ}	63.51±0.18 ^{bQ}	61.33±0.16 ^{cQ}	59.36±0.15 ^{dQ}	57.44±0.22 ^{eQ}	61.24±0.70
T ₂	86.84±0.07 ^{aP}	84.44±0.22 ^{bP}	82.37±0.15 ^{cP}	79.32±0.07 ^{dP}	76.60±0.14 ^{eP}	81.91±0.97
DPPH (%)						
T ₁	26.54±0.20 ^{aQ}	25.46±0.23 ^{bQ}	23.58±0.17 ^{cQ}	20.60±0.22 ^{dQ}	15.65±0.22 ^{eQ}	22.37±1.04
T ₂	51.77±0.01 ^{aP}	48.58±0.18 ^{bP}	47.43±0.13 ^{cP}	45.68±0.05 ^{dP}	42.93±0.03 ^{eP}	47.28±0.78

Mean $\pm$ S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean $\pm$ S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Table 3: Effect of refrigerated storage ($4\pm 1^{\circ}\text{C}$) on physico-chemical parameters of fermented chicken sausages

Group	Storage Days						Total
	Day 0	Day 3	Day 5	Day 7	Day 9	Day 12	
pH							
T ₁	6.02±0.04 ^{aP}	5.94±0.07 ^{abP}	5.82±0.02 ^{bcP}	5.76±0.02 ^{cdP}	5.67±0.03 ^{dP}	5.53±0.03 ^{eP}	5.79±0.04
T ₂	5.49±0.01 ^{aQ}	5.37±0.03 ^{bQ}	5.28±0.03 ^{bcQ}	5.18±0.01 ^{cdQ}	5.14±0.04 ^{dQ}	5.08±0.05 ^{dQ}	5.25±0.03
TBA							
T ₁	0.76±0.000 ^{eP}	0.98±0.11 ^{dP}	1.34±0.11 ^{cP}	1.62±0.07 ^{bP}	1.83±0.04 ^{abP}	1.93±0.00 ^{aP}	1.38±0.10
T ₂	0.74±0.004 ^{cQ}	0.80±0.02 ^{cQ}	0.90±0.05 ^{cQ}	1.29±0.12 ^{bQ}	1.68±0.06 ^{aQ}	1.83±0.01 ^{aQ}	1.17±0.10
FFA							
T ₁	0.053±0.003 ^c	0.073±0.012 ^c	0.110±0.015 ^b	0.123±0.014 ^{ab}	0.136±0.008 ^{ab}	0.150±0.005 ^a	0.107±0.009
T ₂	0.040±0.005 ^c	0.053±0.008 ^{bc}	0.070±0.011 ^{bc}	0.080±0.010 ^b	0.123±0.008 ^a	0.133±0.013 ^a	0.083±0.009

Mean $\pm$ S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean $\pm$ S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Fatty Acid (FFA) values for both groups were recorded on the final day of both refrigerated and frozen storage. Though, both groups exhibited non-significant

differences in their FFA values on the same day of their respective storage periods (Table 3 and 4).

Table 4: Effect of frozen storage ($-20\pm1^{\circ}\text{C}$) on physico-chemical parameters of fermented chicken sausages

Groups	Storage Days					
	Day 0	Day 15	Day 30	Day 45	Day 60	Total Mean $\pm$ SE
pH						
T ₁	6.02 $\pm$ 0.04 ^{aP}	5.91 $\pm$ 0.01 ^{bP}	5.80 $\pm$ 0.02 ^{cP}	5.73 $\pm$ 0.02 ^{cP}	5.63 $\pm$ 0.02 ^{dP}	5.82 $\pm$ 0.03
T ₂	5.49 $\pm$ 0.01 ^{aQ}	5.42 $\pm$ 0.02 ^{abQ}	5.37 $\pm$ 0.02 ^{bcQ}	5.30 $\pm$ 0.01 ^{cdQ}	5.24 $\pm$ 0.02 ^{dQ}	5.36 $\pm$ 0.02
TBA						
T ₁	0.76 $\pm$ 0.000 ^{eP}	0.78 $\pm$ 0.001 ^{dP}	0.79 $\pm$ 0.001 ^{cP}	0.80 $\pm$ 0.001 ^{bP}	0.82 $\pm$ 0.000 ^{aP}	0.79 $\pm$ 0.004
T ₂	0.74 $\pm$ 0.004 ^{eQ}	0.75 $\pm$ 0.002 ^{dQ}	0.76 $\pm$ 0.000 ^{cQ}	0.78 $\pm$ 0.002 ^{bQ}	0.79 $\pm$ 0.001 ^{aQ}	0.76 $\pm$ 0.005
FFA						
T ₁	0.053 $\pm$ 0.003 ^c	0.060 $\pm$ 0.005 ^c	0.086 $\pm$ 0.008 ^{bc}	0.116 $\pm$ 0.020 ^{ab}	0.126 $\pm$ 0.012 ^a	0.088 $\pm$ 0.008
T ₂	0.040 $\pm$ 0.005 ^c	0.046 $\pm$ 0.012 ^c	0.073 $\pm$ 0.008 ^{bc}	0.103 $\pm$ 0.016 ^{ab}	0.116 $\pm$ 0.008 ^a	0.076 $\pm$ 0.009

Mean $\pm$ S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean $\pm$ S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Microbiological parameters

The total plate count increased with successive storage days under refrigerated as well as frozen storage for both the groups; however, difference in TPC of successive days was less under frozen storage conditions (Table 5). Under refrigerated and frozen storage conditions, the TPC count was comparatively low and was highly acceptable for both groups as *L. plantarum* is capable of producing bacteriocin and was found to be effective against both gram positive and gram negative bacteria as suggested by Kaktcham *et al.* (2012) and Das *et al.* (2014). Total coliform count, *Salmonella* count and *Staphylococcus* count were not detected at any storage interval of refrigerated and frozen storage. Another study conducted by Ahmad and Shrivastav (2007), LAB present in the sausages inhibits the proliferation of other harmful and spoilage microorganisms. The absence of these organisms during storage period might also be accredited to exclusion of these pathogens during heat processing, hygienic handling and safe packaging of fermented chicken sausages.

Sensory parameters

Both groups were evaluated for sensory parameters including general appearance, flavor, texture, juiciness, sourness and overall acceptability during refrigerated and frozen storage at various time intervals (Table 6 and 7). During refrigerated storage, sensory scores slowly declined over time. Notably, the T₂ group consistently outperformed the T₁ group in all sensory attributes during this period. In contrast, sensory scores remained relatively stable for both groups during frozen storage, maintaining levels similar to day 0. This suggests that frozen storage had no significant impact on sensory attributes, even after 60 days, while refrigerated storage resulted in a slight decline in sensory attributes on the 9th day, still within acceptable limits as refrigeration is a recognized preservation method (Jay, 1996). In contrast, the study conducted by Wereńska and Okruszek (2023) depicted that freezing meat can lessen water holding capacity during thawing and lead to higher meat juice losses during heat treatment, potentially affecting sensory quality. As per the findings of Lagerstedt *et al.* (2008) chilled meat

Table 5: Effect of refrigerated ($4\pm1^{\circ}\text{C}$) and frozen storage ($-20\pm1^{\circ}\text{C}$) on microbiological parameters of fermented chicken sausages

Total Plate Count							
Refrigerated storage days							
Group	Day 0	Day 3	Day 5	Day 7	Day 9	Day 12	Total
T ₁	1.16 $\pm$ 0.01 ^{fP}	1.39 $\pm$ 0.02 ^{eP}	1.70 $\pm$ 0.03 ^{dP}	1.86 $\pm$ 0.05 ^{cP}	2.17 $\pm$ 0.06 ^{bP}	2.34 $\pm$ 0.04 ^{aP}	1.77 $\pm$ 0.18
T ₂	1.13 $\pm$ 0.01 ^{fP}	1.27 $\pm$ 0.01 ^{eQ}	1.42 $\pm$ 0.02 ^{dQ}	1.57 $\pm$ 0.03 ^{cQ}	1.74 $\pm$ 0.02 ^{bQ}	1.93 $\pm$ 0.02 ^{aQ}	1.51 $\pm$ 0.12
Frozen storage days							
Group	Day 0	Day 15	Day 30	Day 45	Day 60	Total	
T ₂	1.16 $\pm$ 0.01 ^{dP}	1.23 $\pm$ 0.02 ^{cP}	1.25 $\pm$ 0.01 ^{bcP}	1.29 $\pm$ 0.01 ^{bP}	1.36 $\pm$ 0.02 ^{aP}	1.25 $\pm$ 0.03	
T ₂	1.13 $\pm$ 0.01 ^{dP}	1.18 $\pm$ 0.03 ^{cdP}	1.22 $\pm$ 0.02 ^{bcP}	1.26 $\pm$ 0.01 ^{bP}	1.34 $\pm$ 0.01 ^{aP}	1.22 $\pm$ 0.03	

Mean $\pm$ S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean $\pm$ S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Table 6: Effect of refrigerated storage ($4\pm1^{\circ}\text{C}$) on sensory quality of fermented chicken sausages

Parameter	Group	Day 0	Day 3	Day 5	Day 7	Day 9	Total
General	T ₁	5.80±0.20 ^{aQ}	5.40±0.24 ^{abQ}	5.00±0.31 ^{bc}	5.00±0.00 ^{bc}	4.40±0.24 ^{cQ}	5.12±0.23
Appearance	T ₂	7.00±0.00 ^{aP}	6.60±0.24 ^{abP}	6.00±0.31 ^{bc}	5.40±0.40 ^c	5.40±0.24 ^{cP}	6.08±0.32
Flavour	T ₁	6.00±0.00 ^a	5.60±0.24 ^a	5.40±0.24 ^{ab}	4.80±0.20 ^{bcQ}	4.60±0.24 ^c	5.28±0.25
	T ₂	6.60±0.24 ^a	6.40±0.24 ^{ab}	5.80±0.20 ^{bc}	5.60±0.24 ^{cP}	5.40±0.24 ^c	5.96±0.23
Texture	T ₁	6.20±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.60±0.24 ^{ab}	5.40±0.40 ^{ab}	4.80±0.37 ^{bQ}	5.56±0.23
	T ₂	7.00±0.00 ^{aP}	6.60±0.24 ^{abP}	6.40±0.24 ^{bc}	6.00±0.00 ^{cd}	5.80±0.20 ^{dP}	6.36±0.21
Juiciness	T ₁	6.00±0.00 ^{aQ}	5.40±0.40 ^{ab}	5.20±0.37 ^{abQ}	5.00±0.31 ^{ab}	4.60±0.40 ^b	5.24±0.23
	T ₂	7.20±0.20 ^{aP}	6.40±0.24 ^{abQ}	6.20±0.20 ^{bP}	6.00±0.31 ^b	5.60±0.40 ^b	6.28±0.26
Sourness	T ₁	5.80±0.20 ^{aQ}	5.40±0.24 ^{abP}	4.80±0.37 ^{bc}	4.60±0.24 ^{bc}	4.40±0.24 ^c	5.00±0.26
	T ₂	6.80±0.20 ^{aP}	6.40±0.24 ^{ab}	5.80±0.48 ^{abc}	5.40±0.24 ^{bc}	5.20±0.37 ^c	5.92±0.30
Overall	T ₁	5.80±0.20 ^{aQ}	5.20±0.37 ^{abQ}	4.80±0.37 ^{bcQ}	4.40±0.24 ^{bcQ}	4.20±0.20 ^{cQ}	4.88±0.28
Acceptability	T ₂	7.00±0.00 ^{aP}	6.60±0.24 ^{aP}	6.40±0.24 ^{abP}	5.80±0.20 ^{bcP}	5.60±0.24 ^{cP}	6.28±0.25

Mean±S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean±S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

Table 7: Effect of frozen storage ($-20\pm1^{\circ}\text{C}$) on sensory quality of fermented chicken sausages

Parameter	Group	Day 0	Day 15	Day 30	Day 45	Day 60	Total
General	T ₁	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.60±0.24 ^{aQ}	5.60±0.24 ^{aQ}	5.72±0.04
Appearance	T ₂	7.00±0.00 ^{aP}	7.00±0.00 ^{aP}	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.88±0.04
Flavour	T ₁	6.00±0.00 ^a	6.00±0.00 ^a	5.80±0.20 ^a	5.80±0.20 ^a	5.80±0.20 ^a	5.88±0.04
	T ₂	6.60±0.24 ^a	6.60±0.24 ^a	6.40±0.24 ^a	6.40±0.24 ^a	6.20±0.20 ^a	6.44±0.07
Texture	T ₁	6.20±0.20 ^{aQ}	6.20±0.20 ^{aQ}	6.00±0.31 ^a	6.00±0.00 ^{aQ}	5.60±0.24 ^{aQ}	6.00±0.10
	T ₂	7.00±0.00 ^{aP}	7.00±0.00 ^{aP}	6.80±0.20 ^a	6.80±0.20 ^{aP}	6.60±0.24 ^{aP}	6.84±0.07
Juiciness	T ₁	6.00±0.00 ^{aQ}	6.00±0.00 ^{aQ}	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.88±0.04
	T ₂	7.20±0.20 ^{aP}	7.20±0.20 ^{aP}	7.00±0.00 ^{aP}	7.00±0.00 ^{aP}	7.00±0.00 ^{aP}	7.08±0.04
Sourness	T ₁	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.60±0.24 ^{aQ}	5.60±0.24 ^{aQ}	5.60±0.24 ^{aQ}	5.68±0.04
	T ₂	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.60±0.24 ^{aP}	6.60±0.24 ^{aP}	6.72±0.04
Overall	T ₁	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.80±0.20 ^{aQ}	5.60±0.24 ^{aQ}	5.60±0.24 ^{aQ}	5.72±0.04
Acceptability	T ₂	7.00±0.00 ^{aP}	7.00±0.00 ^{aP}	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.80±0.20 ^{aP}	6.88±0.04

Mean±S.E. with different superscripts row wise (small alphabets-a b c d e f) differ significantly at $P<0.05$.

Mean±S.E. with different superscripts column wise (capital alphabet-PQ) differ significantly at $P<0.05$

T₁=Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g), T₂= Sausages prepared with starter culture *Lactobacillus plantarum* (10^8 cfu/g) with additional supplementation of 1% each of Dextrose and Starch

was generally perceived as more tender, juicier and had a more intense meat taste compared to frozen meat.

CONCLUSION

This study revealed that incorporation of probiotic *L. plantarum*, along with dextrose and starch, significantly enhanced the antioxidant properties and overall quality of fermented chicken sausages. These improvements translated to extended shelf lives of up to 12 days under refrigerated conditions and a remarkable 60 days under frozen storage, underscoring the potential for tailored fermentation processes to improve food product stability. These findings have valuable implications for the food industry, offering a pathway to produce safer, longer-lasting meat products.

REFERENCES

- Ablikim, B., Liu, Y., Kerim, A., Shen, P., Abdurrim, P. and Zhou, G.H. 2016. Effects of breed, muscle type, and frozen storage on physico-chemical characteristics of lamb meat and its relationship with tenderness. *CyTA-Journal of Food*. **14**(1): 109-16.
- Agüero, N.D., Frizzo, L.S., Ouwehand, A.C., Aleu, G. and Rosmini, M.R. 2020. Technological characterisation of probiotic lactic acid bacteria as starter cultures for dry fermented sausages. *Foods*, **9**(5): 596.
- Ahmad, S. and Srivastava, P.K. 2007. Quality and shelf life evaluation of fermented sausages of buffalo meat with different levels of heart and fat. *Meat Science*, **75**(4): 603-9.
- Antara, N.S., Sujaya, I.N., Yokota, A., Asano, K. and Tomita, F. 2004. Effects of indigenous starter cultures on the microbial and physicochemical characteristics of Urutan, a Balinese fermented sausage. *Journal of bioscience and bioengineering*, **98**(2): 92-8.

- Brewer, M.S., Ikins, W.G. and Harbers, C.A.Z. 1982. TBA values, sensory characteristics and volatiles in ground pork during long term frozen storage: Effect of packaging. *Journal of Food Science*, **57**: 558-563.
- Christiansen, L.N., Tompkin, R.B., Shaparis, A.B., Johnston, R.W. and Kautter, D.A. 1975. Effect of sodium nitrite and nitrate on Clostridium botulinum growth and toxin production in a summer style sausage. *Journal of Food Science*, **40**(3): 488-90.
- Das, A., Sasidharan, S., Achuthan, T. and Sindhuja, M.E. 2014. Isolation, Characterization and Estimation of antimicrobial activity of novel bacteriocin from *Lactobacillus plantarum*. *International Journal of Current Microbiology and Applied Sciences*, **3**(5): 227-32.
- Duncan, D.B. 1955. Multiple range and multiple F tests. *Biometrics*, **11**(1): 1-42.
- Fernández-López, J., Viuda-Martos, M. and Pérez-Alvarez, J.A. 2021. Quinoa and chia products as ingredients for healthier processed meat products: Technological strategies for their application and effects on the final product. *Current Opinion in Food Science*, **40**: 26-32.
- Geeta and Yadav, A.S. 2017. Antioxidant and antimicrobial profile of chicken sausages prepared after fermentation of minced chicken meat with *Lactobacillus plantarum* and with additional dextrose and starch. *LWT-Food Science and Technology*, **77**: 249-58.
- Igene, J.O., Pearson, A.M., Dugan, Jr. L.R. and Price, J.F. 1980. Role of triglycerides and phospholipids on development of rancidity in model meat systems during frozen storage. *Food Chemistry*, **5**(4): 263-76.
- Jay, J.M. 1996. In: *Modern food microbiology*, 4th edn., Publishers and Distributors, New Delhi. pp 38-39.
- Kachouri, F., Ksontini, H., Kraiem, M., Setti, K., Mechmeche, M. and Hamdi, M. 2015. Involvement of antioxidant activity of *Lactobacillus plantarum* on functional properties of olive phenolic compounds. *Journal of food science and technology*, **52**: 7924-33.
- Kaktcham, P.M., Zambou, N.F., Tchouanguep, F.M., El-Soda, M. and Choudhary, M.I. 2012. Antimicrobial and safety properties of lactobacilli isolated from two Cameroonian traditional fermented foods. *Scientia Pharmaceutica*, **80**(1): 189-204.
- Kato, K., Terao, S., Shimamoto, N. and Hirata, M. 1988. Studies on scavengers of active oxygen species. 1. Synthesis and biological activity of 2-O-alkylascorbic acids. *Journal of Medicinal Chemistry*, **31**(4): 793-8.
- Kaur, R. and Sharma, M. 2019. Cereal polysaccharides as sources of functional ingredient for reformulation of meat products: A review. *Journal of Functional Foods*, **62**: 103527.
- Keller, J.E., Skelley, G.C. and Acton, J. C. 1974. Effect of meat particle size and casing diameter on summer sausage properties during drying. *Journal of Food Protection*, **37**(2): 101-6.
- Konieczko, R. 1979. *Handbook for meat chemists*. (Avery Publishing Group: Wayne, NJ).
- Kumar, A. and Chattopadhyay, S. 2007. DNA damage protecting activity and antioxidant potential of pudina extract. *Food Chemistry*, **100**(4): 1377-84.
- Lagerstedt, Å., Enfält, L., Johansson, L. and Lundström, K. 2008. Effect of freezing on sensory quality, shear force and water loss in beef M. *Longissimus dorsi*. *Meat Science*, **80**(2): 457-61.
- Li, Shengyu., Zhao, Y., Zhang, L., Zhang, X., Huang, L., Li, D., Niu, C., Yang, Z. and Wang, Q. 2012. Antioxidant activity of *Lactobacillus plantarum* strains isolated from traditional Chinese fermented foods. *Food chemistry*, **135**(3): 1914-9.
- Li, Z., Teng, J., Lyu, Y., Hu, X., Zhao, Y. and Wang, M. 2018. Enhanced antioxidant activity for apple juice fermented with *Lactobacillus plantarum* ATCC14917. *Molecule*, **24**(1): 51.
- Liu, Y., Wan, Z., Yohannes, K.W., Yu, Q., Yang, Z., Li, H., Liu, J. and Wang, J. 2021. Functional characteristics of Lactobacillus and yeast single starter cultures in the ripening process of dry fermented sausage. *Frontiers in Microbiology*, **11**: 611260.
- Luan, X., Feng, M. and Sun, J. 2021. Effect of *Lactobacillus plantarum* on antioxidant activity in fermented sausage. *Food Research International*, **144**: 110351.
- Shirwaikar, A., Shirwaikar, A., Rajendran, K. and Punitha, I.S. 2006. *In vitro* antioxidant studies on the benzyl tetra isoquinoline alkaloid berberine. *Biological and Pharmaceutical Bulletin*, **29**(9): 1906-10.
- Emiecińska, K., Hnatyk, N., Daszkiewicz, T., Kubiak, D. and Matusevičiu, P. 2015. The effect of frozen storage on the quality of vacuum-packaged turkey meat. *Veterinarija ir Zootechnika*, **71**(93).
- Speck, M.L. 1992. *Compendium of methods for the microbiological examination of foods* (2nd ed.). Washington, D.C: APHA Technical Committee on Microbiological Methods for Foods.
- Tannock, G.W., Wilson, C.M., Loach, D., Cook, G.M., Eason, J., O'toole, P.W., Holtrop, G. and Lawley, B. 2012. Resource partitioning in relation to cohabitation of Lactobacillus species in the mouse forestomach. *The ISME journal*, **6**(5): 927-38.
- Tarladgis, B.G., Watts, B.M., Younathan, M.T. and Dugan, L. 1960. *Journal of the American Oil Chemists Society*, **37**: 44-48.
- Toldrá, F. and Flores, M. 1998. The role of muscle proteases and lipases in flavor development during the processing of dry-cured ham. *Critical Reviews in Food Science*, **38**(4): 331-352.
- Walter, J. 2008. Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. *Applied and environmental microbiology*, **74**(16): 4985-4996.
- Wei, R., Wang, P., Han, M., Chen, T., Xu, X. and Zhou, G. 2017. Effect of freezing on electrical properties and quality of thawed chicken breast meat. *Asian-Australasian journal of animal sciences*, **30**(4): 569.
- Wereńska, M. and Okruszek, A. 2023. Impact of frozen storage on some functional properties and sensory evaluation of goose meat. *Poultry Science*, **102**(10): 102894.
- Witte, V.C., Krause, G.F. and Bailey, M.E. 1970. A new extraction method for determining 2 thiobarbituric acid values of pork and beef during storage. *Journal of food Science*, **35**(5): 582-585.