Full Length Article

OCCURRENCE OF *Dictyocaulus filaria* INFECTION IN AN ORGANIZED SHEEP FARM OF TAMIL NADU, INDIA

M. Prabhu ^{1*}, C. Sreekumar², N. Prema³, P.C. Sakthivel⁴, R. Anilkumar⁵ and M. Iyue⁶

Sheep Breeding Research Station Tamil Nadu Veterinary and Animal Sciences University Sandynallah, Nilgiris District – 643 237 Tamil Nadu, India

ABSTRACT

The sheep lungworm Dictyocaulus filaria is of world-wide distribution and causes verminous pneumonia which may result in weight loss and death of the infected sheep. The current investigation was carried out in an organized farm in the Nilgiris district of Tamil Nadu, India during the period from 2012 to 2014. Infected sheep showed clinical illness varying from dullness, depression, moderate coughing, dyspnoea and labored breathing with little mortality. On necropsy, both larval and adult stages of lungworm were recovered from trachea, bronchi and bronchioles. The identity of the nematodes was confirmed as D. filaria based on the socks shaped spicules in males and anterior knob in the larva. Of the 55 animals, including 17 lambs examined during the study period, eight animals were confirmed to have died due to verminous pneumonia. Apart from that, in a few lambs, lung abscess was noticed. Though the infection was present throughout the year, the clinical illness was pronounced in dry summer (n=23) and rainy season of south west monsoon (n=19). The disease was managed by minimizing the exposure of lambs to the contaminated pasture and by deworming with two doses of levamisole hydrochloride @ 7.5 mg/kg body weight at 21 day interval to control the lungworm infection in sheep.

Key words: Sheep, Dictyocaulus filaria, Verminous pneumonia, Tamil Nadu, India

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¹ Assistant Professor, Department of Veterinary Microbiology, Veterinary College and Research Institute,

Udumalpet, Tamil Nadu - 642 205, Corresponding author Email: drprabhuvirol@gmail.com

² Professor and Head, Department of Wildlife Science, Madras Veterinary College, Chennai - 600 007

³Assistant Professor and Head,

⁴Assistant Professor, Department of Animal Nutrition, Veterinary College and Research Institute, Namakkal - 637 001

⁵Director of Distance Education, Tamil Nadu Veterinary and Animal Sciences University, Anna Salai, Nandanam, Chennai – 600 035

⁶Professor (Rted)

INTRODUCTION

India has the largest sheep population following China (Hegde, 2019) and the total sheep population is about 74.26 million during 2019 and is about 13.8% in total livestock population of the country. This count has increased by 14.13% over previous five years period. The population of sheep is about 4.5 million in Tamil Nadu state, which ranks fifth in position among Indian states (Department of Animal Husbandry and Dairying, 2019; dadh.nic.in). Sheep plays a major role in the livelihood of small and marginal farming community engaged in sheep husbandry through sale of animals.

About half of the loss in terms of mortality and morbidity in sheep farms are caused by pneumonia and endoparasitism including lungworms (Asaye and Alemneh, 2015; Adem, 2016). Ovine lungworms are one of the major respiratory parasites of sheep that are worldwide in distribution particularly common in temperate countries and in high land of tropical and sub-tropical countries (Tesfaye et al., 2015). Dictyocaulus filaria, Muellerius capillaris and Protostrongylus rufescens are the three important nematodes that cause lungworm infection in sheep (Radostits et al., 2007). Although mixed infections of these round worms may occur, in the majority of outbreaks D. filaria is found to be the predominating species (Urguhart et al., 1996). It is also more pathogenic (Umur et al., 2006) and economically significant in sheep husbandry. The lungworms enter the host by ingestion of infective larvae, have direct life cycle and are located in the respiratory tract of the lungs causing chronic syndrome of coughing, unthriftiness in lambs and kids (Tesfaye *et al.*, 2015). They suppress the immunity of respiratory tract of host and hence lead to bronchitis or pneumonia resulting in loss of production in terms of loss of body condition / weight and death (Gelagay *et al.*, 2004).

Depending on the predisposing or risk factors, the incidence of this lungworm infection varies from place to place. Though there are reports on lungworm infection in sheep from various parts of the world, the published literature is rather rare in the Nilgiris district of Tamil Nadu. The current investigation deals with the occurrence of lungworm infection in an organized sheep farm of the Nilgiris district of Tamil Nadu, a southern state of India.

MATERIALS AND METHODS

Study area

The investigation was carried out during the period of January 2012 to October 2014 in Sheep Breeding Research Station (SBRS) an organized farm situated at Sandynallah which is located at latitude of 11° 26' 15.25" north and longitude of 76° 38' 27.34" east in the Nilgiris District, Tamil Nadu state, India. It is situated in the Nilgiri Biosphere Reserve in undulating hill range at 2090 – 2235 m above MSL. The annual mean temperature and rainfall ranges from 0-24°C and 840 – 3000 mm, respectively.

Study animals and management

The Nilagiri, Sandyno and Dorset x Nilagiri cross breeds are the three breeds maintained at SBRS with the average flock strength of 1382, 1307 and 1390 in the years 2012, 2013 and 2014, respectively. The animals were vaccinated once with Johne's disease vaccine after 10 days of birth and once or twice a year with formalin inactivated, alum precipitated enterotoxaemia vaccine. They were dewormed twice with Fenbendazole and Praziquantel during the first three months of life and then followed by deworming with morantel citrate in the hogget and adult animals once in three months period. The animals were maintained under semi-intensive system and allowed for ~ 8 hours grazing in an open pasture; fed with concentrate containing 18% of crude protein in the sheds with ad libitum water even in pasture. During winter and early summer (January to May), the animals received additional supplement of silage prepared from fodder maize.

Clinical picture and postmortem screening of animals

During the study period, many animals showed the clinical manifestations suggestive of respiratory illness such as nasal discharge, ill thrift, weight loss, coughing/ dyspnoea, labored breathing with a few mortalities. A total of 293 sheep carcasses were examined during the study period. The major sites screened for the presence of lungworms include their respiratory passages *viz.*, trachea, lung, bronchi and bronchioles. Representative samples (n=8) from lung that showed the

marked lung lesions along with worms were sent for histopathological examination at Central University Laboratory, Tamil Nadu Veterinary and Animal Sciences University, Chennai.

RESULTS AND DISCUSSION

serious respiratory ailment in domestic small ruminants caused by the parasites called verminous bronchitis or otherwise referred as hoose or husk is worldwide in distribution and is endemic in temperate regions with high rainfall (Taylor et al., 2007). Dictyocaulosis is known to be potentially increasing, and expensive trouble (Ploeger, 2002) in many areas. The prevalence of this illness varies from place to place depending on the climatic factors and ecological conditions such as rainfall, humidity and temperature of the area. Further, the pasture contamination, number of infective larvae in the grazing field, management system followed and immune status of the animals are additional factors contributing to the severity of this infection (Dar et al., 2012).

In the current investigation, the occurrence of the *D. filaria* infection is recorded from the organized sheep farm for the period of three years from 2012-2014. In the flock, many animals showed the clinical manifestations suggestive of respiratory illness. The characteristic clinical signs exhibited by the affected sheep varied from mild depression, dullness, pyrexia, nasal discharge and moderate coughing to severe dyspnoea and labored breathing with a few mortalities. Some animals showed ill thrift

with reduced weight gain, loss of weight, emaciation with retarded growth. These findings were in concurrence with earlier reports (Elsheikh and Khan, 2011; Chakraborty *et al.*, 2014). In severe cases, heavily infected sheep showed characteristic stance with their heads extended forward and open mouth with drooling of saliva.

The pathogenesis of *D. filaria* also depends on their predilection site and the location within the respiratory tract of the animal. They live mainly in the trachea and bronchi affecting a large volume of lung tissue due to aspirated eggs, larvae and debris and hence are considered to be the most pathogenic lungworm species (Tewodros, 2015). In our study, both larval and adult stages of lungworm were recovered from trachea, bronchi and bronchioles. These lungworms were recovered from 55 animals out of 293 animals necropsied during the study period. The details were shown in tables 1 and 2. Among the positive animals,

eight sheep were confirmed to have died mainly due to verminous pneumonia without any other characteristic lesions or confirmed etiology. Further, the histopathology results also confirmed the presence of sections of lungworms.

Further, in seven lambs, tiny nodular lesions or abscesses were noticed in the caudal lobes of lungs with catarrahal bronchitis which is similar to the previous observation of Chilton et al. (2006). The lungs revealed the areas of congestion and consolidation. When dissected, numerous slender, thread-like, creamy white worms were found numbering from very few to bunches along with copious frothy fluid (Fig. 1 and 2). The identity of the nematodes was confirmed as D. filaria based on the socks (boot)-shaped spicules in males and anterior knob in the larva (Dar et al., 2012) (Fig. 3 and 4). The lavage from the tracheal mucus revealed the presence of larviparous egg of the lungworms (Fig. 5).

Table 1. Particulars of infected lambs with *Dictyocaulus filaria* infection (<1 year)

Year	No. of lambs	No. of lambs examined	D. filaria recovered lambs			%
	housed		Ram lambs	Ewe lambs	Total	positivity
2012	327	51	-	1	1	1.96
2013	374	56	12	3	15	26.78
2014	365	55	3	7	10	18.18
Total		162	15	11	26	16.04

Table 2. Details of adult sheep with Dictyocaulus filaria infection

Year	Adult sheep	No. of adult sheep	D. filaria recovered adult sheep			% positivity
	housed	examined	Ram	Ewe	Total	
2012	686	38	-	4	4	10.52
2013	652	40	2	22	24	60.00
2014	859	53	-	1	1	1.88
Total		131	2	27	29	22.13





Fig. 1 and 2. Tracheal passage and section of lung showing the lungworm (*Dictyocaulus filaria*)



Fig. 3. Boot (socks) shaped spicules in the posterior end of the male lungworm (*Dictyocaulus filaria*)

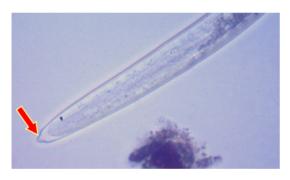


Fig. 4. The anterior knob of larva of Dictyocaulus filaria



Fig. 5. Larvated egg of Dictyocaulus filaria

The main mode of transmission is through ingestion of contaminated pasture and, as the protective immunity develops in adults after exposure, the infection is more common in young animals during their first grazing season. Mainly lambs of 4 - 6 month old are more commonly infected than adults (Tewodros, 2015). Major outbreaks are seen from July to September, when susceptible lambs are on pasture and the parasites have had enough time to reproduce (Taylor et al., 2007). In this sheep farm, the main lambing season falls during February – April, and only 5-10% and 60-75% of the ram lambs and ewe lambs, respectively are retained to replace the breedable stock and the surplus lambs were culled. After about 1½ months of age, the lambs were allowed to suckle the dam twice daily and exposed to grazing. Our report is also in accordance to Taylor et al. (2007) as the major incidence in lambs was recorded during June-September months in lambs. During these months the weaned lambs were solely maintained on pasture and stall feeding of concentrates. In the present investigation, D. filaria worms were recovered from postmortem of 26 lambs with the per cent positivity of 17.1.

In the present study, the per cent positivity for lungworm infection was reported as 22.13 in case of adult sheep. In domestic small ruminants, no significant difference was noticed in the prevalence of lungworm among the different sex of the animals. However, fewer studies reported slightly higher prevalence among ewes than in rams (Ibrahim and Godefa, 2012; Terefe *et al.*, 2013). In this

study also, a higher incidence of *D. filaria* was noticed in adult female (n=27) than in rams (n=2). This might have been due to production and reproduction stress in adult ewes resulting in immunosuppression. Whereas in lambs (< 6 months old) a greater number of male lambs shown positivity than ewe lambs (Table 1). The rationale behind the difference could not be ascertained.

The prevalence of lungworms is different based on geographical region and climatic factor. The distribution of D. filaria depends mainly on pasture contamination by carrier animals. The pasture infectivity in turn is related to rainfall which stimulates the activity of the larvae (Adem, 2016). Moisture is crucial for the survival and development of the larva that is active at moderate temperature of 10-21°C. The larva of lungworm survives best in cool, damp surroundings and under optimum conditions as it can persists for more than a year (Tewodros, 2015). The study area is also located in such climatic conditions (Prabhu et al., 2019). This could have provided the optimum conditions required for the survival of the larvae.

The prevalence rate of 13.4% to 53.6% has been recorded from various parts of Ethiopia by several authors (Alemu *et al.*, 2006; Ibrahim and Godefa, 2012; Adem, 2016). Further, the prevalence is low in spring and summer and rises rapidly in the autumn and winter. Overcrowding in sheds, undersupplied feeding and poor nutrition, concurrent infections usually predispose to lungworm infection (Borji *et al.*, 2012; Tewodros, 2015).

In our study, though the infection was present throughout the year, the clinical illness was pronounced in dry summer (n=23) and heavy rainy seasons (n=19). In the study area summer follows severe winter, during which sheep were maintained in a poor pasture. This could have been the reason for deficient feeding and a greater number of cases in summer. The summer in turn is followed by southwest monsoon which provides ambient temperature for larvae survival. Further, during this period of investigation, the incidence of Oestrus ovis was also recorded in the farm with peak incidence in southwest monsoon (Prabhu et al., 2019). This concurrent infection could have also resulted in stress and might have aggravated the severity of lungworm infection. Hence, the per cent positivity could have been high during summer followed by rainy season of south west monsoon.

The control and prevention of lungworms in sheep mainly involves pasture management to minimize the exposure of lambs, elimination of intermediate host and by treating with anthelmintic drugs. Particularly benzimidazoles, levamisole and ivermectin were reported to be effective (Tewodros, 2015). Further, in places of United Kingdom and Western Europe, an irradiated larval vaccine was also been used. In the current situation, the herd was dewormed with levamisole hydrochloride @ 7.5 mg/ kg body weight and an additional dosing was given at 21 days interval. Thus, it is concluded that preventive measures by way of deworming the animals after proper screening could be taken to avoid the economic loss from mortality due

to lungworm infection and further production loss.

REFERENCES

- Adem, J. (2016). Lungworm infection of small ruminant in Ethiopia. *Advances in Life Science and Technology*, **43:** 12 22.
- Alemu, S., Leykun, E.G., Ayelet, G. and Zeleke, A. (2006). Study on small ruminant lungworms in northeastern Ethiopia. *Veterinary Parasitology*, **142**(3): 330 335.
- Asaye, M. and Alemneh, T. (2015).

 Prevalence of lungworm infection of small ruminants in and Around Bahir Dar City, Amhara Regional State, Ethiopia. *Journal of Veterinary Science and Technology*, **\$12:** 002.
- Borji, H., Azizzadeh, M., Ebrahimi, M. and Asadpour, M. (2012). Study on small ruminant lungworms and associated risk factors in northeastern Iran. *Asian Pacific Journal of Tropical Medicine*, **5**(11): 853 856.
- Chakraborty, S., Kumar, A., Tiwari, R., Rahal, A., Malik, Y., Dhama, K., Pal, A. and Prasad, M. (2014). Advances in diagnosis of respiratory diseases of small ruminants. *Veterinary Medicine International*, 1-16.
- Chilton, N.B., Huby-Chilton, F., Gasser, R.B. and Beveridge, I. (2006). The

- evolutionary origins of nematodes within the order *Strongylida* are related to predilection sites within hosts. *Molecular Phylogenetics and Evolution*, **40**(1): 118 128.
- Dar, L.M., Darzi, M.M., Mir, M.S., Kamil, S.A., Rashid, A., Abdullah, S., Hussain, S.A., Bhat, A.A. and Reshi, P.A. (2012). Prevalence and Pathology of lungworm infection in sheep in Kashmir Valley, India. *Journal of Animal Science Advances*, **2**(8): 678 685.
- Elsheikh, H.M. and Khan, N.A. (2011). Essentials of Veterinary Parasitology. 1st ed. *Caister Academic Press,* Norfolk, UK. 52 - 69 pp.
- Gelagay, A., Laekemariam, Y., Esayas, G., Selam, T. and Kassahun, A. (2004). Epidemiologic and serologic investigation of multifactorial respiratory disease of sheep in the central highland of Ethiopia. *The International Journal of Applied Research in Veterinary Medicine*, 2(4): 274 278.
- Hegde, N.G. (2019). Livestock development for sustainable livelihood of small farmers. Asian *Journal of Research in Animal and Veterinary Sciences*, **3**(2): 1 17.
- Ibrahim, N. and Godefa, Y. (2012). Prevalence of ovine lungworm infection In

- Mekelle Town, North Ethiopia. *The Internet Journal of Veterinary Medicine*, **9**(1): 1 15.
- Ploeger, H.W. (2002). *Dictyocaulus viviparus*: re-emerging or never been away?. *Trends in Parasitology*, **18**(8): 329 332.
- Prabhu, M., Sakthivel, P.C., Prema, N., Anilkumar, R. and Iyue, M. (2019). Occurrence of ovine oestrosis in an organized farm of the temperate terrain region of Nilgiris, Tamil Nadu a southern state of India. *Journal of Entomology and Zoology Studies*, 7(2): 1173 1176.
- Radostits, O.M., Gay, C., Blood, D.C. and Hinchclift, K.W. (2007). Veterinary Medicine: A text book of the diseases of Cattle, Sheep, Goats, Pigs and Horses. 10th ed., London, Harcourt Publishers' Ltd, Pp 1541 1564.
- Taylor, M.A., Coop, R.L. and Wall, R.L. (2007). Veterinary Parasitology. Third edition. Oxford: Blackwell Publishing Ltd.

- Terefe. Y., Tafess, K., Fekadie, G. and Kebede, N. (2013). Prevalence of lungworm infection in small ruminants in North Gondar zone, Amhara National Regional State, Ethiopia. *Journal of Parasitology and Vector Biology*, 5(4): 40 45.
- Tesfaye, S., Hagos, Y., Teklu, A., Gugsa, G. and Gebrekidan, B. (2015). Ovine lungworm infection and associated risk factors in and around Wukro, Eastern Tigray, Ethiopia. *European Journal of Biological Sciences*, 7 (3): 120 124.
- Tewodros, A.E. (2015). A review on: lungworm infection in small ruminants. *World Journal of Pharmaceutical and Life Sciences*, **1**(3): 149 159.
- Umur, S., Koroglu, E., Guclu, F. and Tinar, R. (2006). Nematodes. In: Helminthology (eds. of R. Tinar), Pp 214 449.
- Urquhart, H.M., Armour, J., Duncan, J.L., Dunn, A.M. and Jennings, F.W. (1996). Veterinary Parasitology. 2nd Edition, London, Black well science Ltd: 1996, Pp 301- 309.