#### **Short Communication**

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# Salicylic acid: To assuage salinity stress at the early seedling stage of rice

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Agriculture production is highly vulnerable to biotic and abiotic stresses. Abiotic stress is one of the major factors which overshadow the crop growth and productivity world-wide. Due to global warming, rise in sea levels, surplus irrigation without appropriate drainage in inlands and underlying rocks rich in detrimental salts, the area under salt stress is expanding. Salinity affects the plant growth and yield by interfering the physiological, biochemical, cellular and molecular mechanism in plants (Gupta and Huong, 2014). Rice is the major crop which is adversely affected by saline soil in its seedling and vegetative stage. Salt stress induces cellular accumulation of reactive oxygen species (ROS), which damages the biomolecules of the cell (Amor *et al.*, 2007; Mansour *et al.*, 2005).

Although several mechanical, chemical and biochemical approaches are being pursued to overcome the menace of soil salinity, exogenous application of plant growth regulators are considered most effective. Plant hormones have been shown to play a key role in abiotic stress responses (Kaya *et al.*, 2009). Exogenous application of hormones has been shown to impart abiotic stress

tolerance (Yang and Mei 2004; Azooz, 2009; Kanmani*et al.*, 2017).

Among the endogenous growth regulators, a phenol compound named salicylic acid (SA) plays major role under stress (Sakhabutdinova *et al.*,2003). The application of SA could alleviate the adverse effects of salt stress by the regulation of physiological mechanism in rice (Hayat *et al.*, 2010). SA has a protective function which includes the development of anti-stress programs and acceleration of normalization of growth processes after removal of stress factors (Sakhabutdinova *et al.*, 2003). SA potentially generates a wide range of metabolic responses in plants that affects plant water relations. This molecule has also found to be active in reducing oxidative stress by scavenging the reactive oxygen species (El-Esawi., 2017).

With this above background the primary objective of the study was to assess whether the exogenous application of SA as seed treatment will give salinity tolerance on rice genotypes at early seedling growth stages.

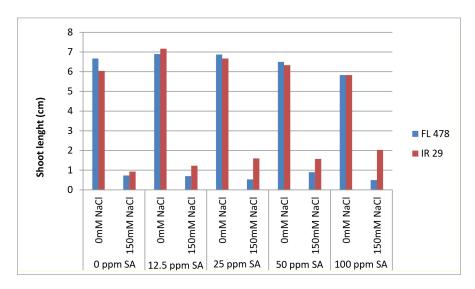
In order to standardize the SA concentration required to overcome the inhibitory effect of salinity stress on early seedling growth, salinity tolerant rice genotype, FL478 and

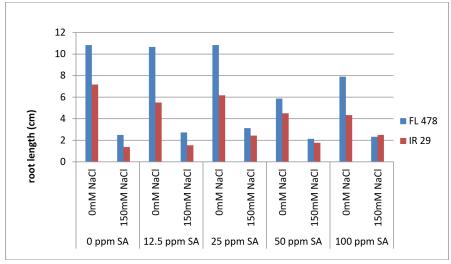


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salinity susceptible rice genotype, IR 29 were selected. The seeds of these genotypes were presoaked overnight in different concentrations of SA such as control (0ppm), 12.5, 25, 50 and 100 ppm. Then the seeds were spread in petriplates containing 150 mM NaCl and a control plate moistened with distilled water was also maintained. After 14 DAS, the growth parameters like shoot length and root length were measured. Under salinity conditions, there was a reduction in both root length and shoot length in two

the genotypes when compared with control (0mM NaCl) (Figure 1). Better root length was noticed in seeds treated with 12.5 ppm and 100 ppm of SA. But better shoot length was noticed in seeds treated with 100 ppm SA. Moreover, in root length the improvement was better in 100 ppm SA treated seeds in IR 29, the salinity susceptible genotype (Figure 1). Therefore 100ppm SA was standardized as an optimum concentration for further experiment to assess the role of salicylic acid in salinity stress tolerance.





 $Figure \ 1: Standardization \ of \ Salicylic \ acid \ concentration \ for \ seed \ treatment \ with \ two \ genotypes \ (FL478-Salt \ tolerant \ and \ IR \ 29-Salt \ susceptible \ genotypes)$ 

To study the effect of salicylic acid in alleviating salinity stress, growth and biochemical parameters of seven genotypes FL 478, ADT 45, ADT 46, ADT 50, CR 1009, KKL(R)1, and IW Ponni were selected and treated with 100 ppm SA overnight and sown under saline condition(150 mM NaCl) and also under control condition. The selected

genotypes were grown in inclined plate method (Punjabi and Basu, 1982) with some modifications, to assess the role of SA in reducing the salinity stress effect on the growth parameters. The root to shoot ratio (length basis), one of the important physiological trait for abiotic stress tolerance (Thomas *et al.*, 2020) of the seedlings were derived from



Salicylic acid treated (100 ppm) and untreated (0 ppm) from control and salinity condition. Under control condition the mean root to shoot ratio has reduced from 1.78 to 1.77 at 100 ppm SA. Whereas, under salinity stress, the mean value has significantly increased from 1.8 to 2.83 when treated with SA at 100 ppm (Table 1). The root to shoot ratio on length basis of FL 478 which is a salt tolerant genotype increases from 2.46 to 2.50 when seeds

were treated with 100 ppm SA under Salinity stress and similar trend was recorded in genotypes ADT 45, ADT 46, ADT 50, KKL (R) 1 and IW PONNI with 100 ppm SA treatment. Root to shoot ratio on length basis increased under salinity condition when seeds are treated with 100 ppm SA than the non treated seeds indicating that, when seeds are treated with SA, the tolerance towards salinity get enhanced.

Table 1. Root to shoot ratio (length basis) of rice genotypes under salinity conditions with and without Salicylic acid treatment in inclined plate method

Genotypes	0mMNaCl		150mMNaCl	
	0 ppm SA RL/SL	100 ppm SA RL/SL	0 ppm SA RL/SL	100 ppm SA RL/SL
ADT 45	1. 88	1. 64	1. 85	2. 23
ADT 46	1. 36	1. 73	0.38	2. 98
ADT 50	2. 32	1. 93	0.43	2. 75
CR 1009	1. 61	1. 89	2.34	1. 87
KKL(R) 1	1. 76	1. 56	2.98	3. 01
IW PONNI	1. 84	2. 11	2. 13	4. 46
Mean	1. 78	1. 77	1. 8	2.83
${ m CD}_{0.05}$				
GENOTYPE	0. 48			
NaCl	0. 26			
SA	0. 68			
G*NaCl	0. 26			
G*SA	0. 68			
NaCl*SA	0. 37			
G*NaCl*SA	0.97			

It indicated that exogenous SA application on rice plants lead to an increase in salt tolerance. Similar results were recorded in rice by Jini and Joseph (2017). Likewise, SA treatment has been reported to enhances the growth of barley (El- Tayeb, 2005), wheat (Tammam et al., 2008), sunflower (Noreen et al., 2009), mungbean (Khan et al., 2012) under salt stress. SA treatment increases the growth of radicle cells by both the division and expansion of meristem cells (Boukraa et al., 2013). The interaction between salinity and SA possibly induces the genes encoding salt resistance and acts on germination by increasing the physiological activity and mobilization of the reserved material necessary for the growth (Szalai et al., 2005).

Different abiotic stresses like heat, drought, salinity, and heavy metal affect the process of photosynthesis directly and indirectly. The direct and indirect effects include a reduced activity of photosynthesis enzymes, decrease in CO2 assimilation rate, degradation of chlorophyll content, etc (Swami et al., 2021). When the biochemical parameters like total chlorophyll and carotenoids were quantified (Aron, 1949), both showed increased trend when seeds were treated with 100ppm SA under salinity and normal condition (Fig. 2). SA treatments, increased pigments content of the plants under control and salinity condition. Ghassemi-Golezani and Lotfi (2015) reported that exogenous application of salicylic acid showed an increase in Chlorophyll a, b and carotenoids content under normal field condition. Similarly, there was an increase in photosynthetic pigments on treatment with SA in corn (Khan et al., 2003) and in soybean (Zhou et al., 1999).



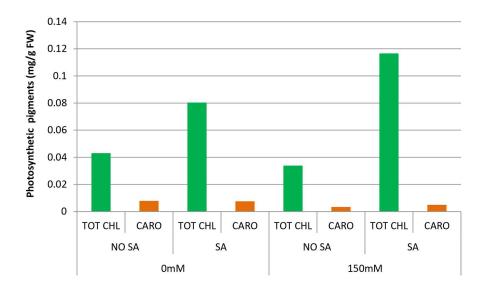


Figure 2: Effects of 100 ppm salicylic acid (SA) on Photosynthetic pigments of rice varieties for salt tolerance during early seedling growth.

# Summary

SA has the ability to overcome the inhibitory effects of salinity at seedling stage of rice. Presoaking in 100ppm salicylic acid showed an increase in root to shoot ratio and photosynthetic pigments, which might have increased the hardiness of seedling to thrive under saline condition.

#### Declarations/ disclosures

## Compliance with ethical standards

NA

### **Author Contribution**

S.Nadaradjan and Ramanadane Tconceptualized and designed the experiments. Nalishma Raghu, Niharika B, Karpagaraj R, Sowmya A, Arulnidhi K performed experiment. Manuscript was drafted by S Nadaradjan and Nalishma Raghu.

# **Conflict of Interest**

Authors declare that they have no conflict of interest.

#### Reference

- Amor N. B, Jiménez A, W Megdiche, M Lundqvist, F Sevilla, C Abdelly.2007. Kinetics of the anti-oxidant response to salinity in the halophyte *Cakilemaritima*. *Journal of Integrative Plant Biology*, 49(7): 982-992.
- Aron D. 1949. Copper enzymes isolated chloroplasts, polyphenoloxidase in *Beta vulgaris*. *Plant Physiology*. 24: 1-15.

- Azooz MM.2009 Salt stress mitigation by seed priming with salicylic acid in two faba bean genotypes differing in salt tolerance. International Journal for Agricultural Biology, 11:343–350.
- 4. Bohra JS, H Dorffling,K Dorffling. 1995. Salinity tolerance of rice (*Oryza sativa* L.) with reference to endogenous and exogenous abscisic acid. Journal of Agronomy and Crop Science, 174:79–8.
- Boukraâ D, K Benabdelli, L Belabid, F Bennabi. 2013.
   Effect of salinity on chickpea seed germination pre-treated with salicylic acid. Scient Journal Biology Science, 2(4): 86-93.
- El-Esawi, MA. 2017. Phytohormones: Signaling Mechanisms and Crosstalk in Plant Development and Stress Responses; BoD-Books on Demand: Norderstedt, Germany, ISBN 978-953-51-3411-4.
- El-Tayeb MA. 2005. Response of barley grains to the interactive effect of salinity and salicylic acid. *Plant Growth Regulators*, 45: 215-224.
- 8. Ghassemi-Golezani K, R Lotfi.2015. The impact of salicylic acid and silicon on chlorophyll a fluorescence in mung bean under salt stress. *Russian journal plant physiology*, **62**(5): 611-616.
- 9. Gupta B and B Huang. 2014. Mechanism of salinity tolerance in plants: physiological, biochemical, and molecular characterization. *International Journal of Genomics*, (1):1-18.



- Hayat Q, S Hayat, M Irfan, A Ahmad. 2010. Effect of exogenous salicylic acid under changing environment: A review Environmental and Experimental Botany, 68(1): 14-25.
- Kanmani, E, V Ravichandran, R Sivakumar, A Senthil, KK Surendarand P Boominathan. 2017.
   Influence of plant growth regulators on physiological traits under salinity stress in constrasting rice varieties (Oryza sativa L.). International Journal of Current Microbiology and Applied Sciences, 6(5): 1654-61.
- 12. Kaya C,AL Tuna, I Yoka .2009. The Role of Plant Hormones in Plants Under Salinity Stress. In: Ashraf M., Ozturk M., Athar H. (eds) Salinity and Water Stress. Tasks for Vegetation Sciences, vol 44. Springer, Dordrecht.
- 13. Khan N, S Syeed, A Masood, R Nazar, N Iqbal. 2010. Application of salicylic acid increases contents of nutrients and antioxidative metabolism in mungbean and alleviates adverse effects of salinity stress. *International Journal of Plant Biology*, 1:1-8.
- 14. Khan W, P Balakrishnanand DL Smith. 2003. Photosynthetic responses of corn and soybean to foliar application of salicylates. *Journal of Plant Physiology*, **160**(5): 485-492.
- 15. Mansour MMF, KH Salama, FZM Ali, AF AbouHadid.2005.Cell and plant responses to NaCl in Zea mays L.cultivars differing in salt tolerance. General and Applied Plant Physiology, 31(1): 29-41.
- Noreen S, M Ashraf, M Hussain, A Jamil.2009. Exogenous application of salicylic acid enhances antioxidative capacity in salt stressed sunflower (*Helianthus annuus* L.) plants. *Pakistan Journal of Botany*, 41(1):473-479.
- 17. Punjabi, Band RN Basu.1982. Testing germination and seedling growth by an inclined glass plate blotter method. *Indian Journal of Plant Physiology*, **25**(3): 289-295.

- Sakhabutdinova AR, DR Fatkhutdinova, MV Bezrukova, FM Shakirova.2003. Salicylic acid prevents the damaging action of stress factors on wheat plants. *Bulgarian Journal of Plant Physiology*, 21:314-319.
- Swami P, R Munjal and K Deswal.2021. Targeting Photosynthesis under Abiotic Stress. Journal of Cereal Research.
- 20. Szalai G, E Páld,T Janda.2005.Effect of salt stress on the endogenous salicylic acid content in maize (*Zea mays* L.) plants. *ActaBiologicaSzegediensis*, **49(1-2)**: 47-48.
- 21. Tammam AA, MFA Alhamd, MM Hemeda. 2008. Study of salt tolerance in wheat (*Triticumaestivum* L.) cultivar. *Australian Journal of Crop Science*, **1**(3):115-125.
- 22. Thomas, T., J Purushothaman, R Janarthanan, N Anusuya, PG Medisetti, J Karthick, . . . & S Thirumeni. 2020. Identification of rice genotypes for seedling stage multiple abiotic stress tolerance. *Plant Physiology Reports*, **25**(4): 697-706.
- 23. Yang, Y, M Qiand C Mei. 2004. Endogenous salicylic acid protects rice plants from oxidative damage caused by aging as well as biotic and abiotic stress. *The Plant Journal*, **40**(6): 909-919.
- 24. Zhou XM, AF MacKenzie, CA Madramootooand DL Smith.1999. Effects of stem-injected plant growth regulator, with or without sucrose, on grain production, biomass and photosynthetic activity of field-grown corn plants. Journal of Agronomy and Crop Science, 183(2):103-110.

