Seed Mycoflora in Kidney Bean (Phaseolus vulgaris L.) in Himachal Pradesh

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ABSTRACT Investigations on comparative occurrence of mycoflora on vegetable and pulse type of beans revealed that vegetable type varieties harboured more mycoflora as compared to pulse type beans. Agar plating of seeds showed the presence of 16 species of pathogenic and storage fungi belonging to ten genera. However, the frequency of fungi was more on unsterilized seeds (4.0-64.0 %) as compared to sterilized seeds (8.0-32.0 %) in both types of beans. Major storage and pathogenic fungi recorded were the species of Alternaria, Aspergillus, Cladosporium, Colletotrichum, Fusarium, Penicillium, Rhizoctonia, Rhizopus, Stemphylium and Trichoderma. Presence of mycoflora on seed had little effect on seed germination; however, higher germination was recorded in surface sterilized seeds of both types of beans. Seed smearing with fungal spore suspension revealed that almost all the storage fungi except Cladosporium sp. had least effect on seed germination. However, culture filtrates of these fungi reduced the germination drastically along with induction of variable symptoms.

Keywords: Seed mycoflora, vegetable and pulse type kidney bean, seed germination, spore suspension, culture filtrate

Kidney bean, popularly known as Rajmash, occupies a premier position amongst the various grain legumes grown in Himachal Pradesh. The pulse type of bean crop is cultivated in mid (900-1300 m asl) and high hills (1800-3000 m asl) of Chamba, Kangra, Kinnaur, Kullu, Mandi, Shimla and Sirmour districts [1] whereas, snap beans (vegetable type) are cultivated mainly in low lying areas of the state. Seed being one of the most important yield contributing factors in both the types, has been reported to be affected by various biotic and abiotic factors during maturation, harvesting and storage [2, 3]. Of these factors, microorganisms cause bio-deterioration of seed during storage and in the field, some of them like Colletotrichum lindemuthianum (Sacc. and Magn.), Sclerotinia sclerotiorum (Lib) de Bary, Sclerotium rolfsii Sacc. and Rhizoctonia solani Kuhn [4, 5] are serious seedborne pathogens of beans. In the present investigations, an effort has been made to study the colonization behavior of seed mycoflora on different types of kidney bean varieties and effect of surface sterilization on seed germination and development of mycoflora.

MATERIALS AND METHODS

The seeds of different types of kidney bean varieties viz. Contender, Hans, Kentucky wonder,

Laxmi, SVM-1 (vegetable type); Baspa, Him-1, Jawala, Kanchan and Triloki (pulse type) were collected from the farmer's stores immediately after harvest and kept in metallic bins for six months at ambient temperature (November-April 2000-2001). After six months storage 100 seeds/sample, both unsterilized and sterilized (with sodium hypochlorite 0.1%) were subjected to Agar plating to record the development of mycoflora. Ten seeds were placed in each plate equidistantly [3] on Malt Extract Agar (MEA) medium supplemented with 100 ppm streptocycline to avoid bacterial contamination. These plates were incubated at 25±1°C in BOD incubator with 12 hours alternating cycles of light and dark and thereafter shifted to deep freezer (-20°C) for 24 hours to prevent seed germination. The plates were then again incubated at 25±1°C for 7 days. Incubated seeds were examined visually and under stereoscopic binocular microscope for the associated mycoflora. Rolled paper towel method [6] was employed to record seed germination percentage of different varieties after six months of storage. In order to study the effect of isolated mycoflora on seed health, two cultivars one each of pulse type (Kanchan) and vegetable type (contender) were treated with spore suspension and culture filtrates of isolated fungi by employing seed smearing method [7]. Culture filtrates obtained from 15 days old culture were used

Table 1. Comparative mycoflora (%) on kidney bean seeds (unsterilised) of vegetable and pulse type cultivars

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Trichoderma sp.				20					32			•	,	
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Penicillium capsulatum		24		h comme	16								80	
E. solani						32			·			1	ad a	
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fusarium spp.						4			95000	·	,			
Colletotrichum lindemuthianum		œ	24	16		16				20	24	,		
Cladosporium sp.			HE TOWN	24	•	80							4 5	*
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.ds Alternaria				44		inst sold in			HEAVE APIL		11.	48	32	
Germination (%)		88.00	85.33	85.33	29.06	29.06	3.28		86.67	20.00	76.76	91.33	95.32	5.48
Cultivars	Vegetable Type	Contender	Hans	Kentucky Wonder	Laxmi	SVM-1	CD (P=0.05)	Pulse Type	Baspa	Him 1	Jawala	Kanchan	Triloki	CD (P=0.05)

Table 2. Comparative mycoflora(%) on kidney bean seeds (sterilised) of vegetable and pulse type cultivars

Cultivars	(%)		E			Solani	Ġ	Jora
	Germination	Alternaria sp.	Colletorichum lindemuthianu	Eusarium spp.	F. oxysporum	Rhizoctoma s	Shemphylium	Total mycoflora
Vegetable Type								
Contender	93.00			48		32	24	3
Hans	92.00					24		
Kentucky Wonder	88.00			20	44	32		3
Laxmi	94.00		12			16		2
SVM-1	93.33	24		20		20	24	4
CD (P=0.05)	4.00							
Pulse Type								0
Baspa	96.66		The Party	E ONE				0
Him I	85.00	-	16		e Tribun	8	State State S	2
Jawala	97.97		12	100	11 -10 11	12	or handle p	2
Kanchan	92.00	32		-		12		2
Triloki	98.65	16	THE PARTY OF THE P		A STORY	12	He Cale	2
CD (P=0.05)	7.09				Transport.	e will erie		

to treat the seeds by giving a dip of six hours and then placed on two layers of moistened germination sheets for growth [8, 9] and data were recorded on seed germination and seedling rot.

RESULTS AND DISCUSSION

Storage studies on occurrence of mycoflora revealed the presence of 16 species of storage and pathogenic fungi belonging to 10 genera on both pulse and vegetable type of beans (Table 1). The vegetable type varieties harboured more number of fungi as compared to pulse type. Maximum number of fungi were recorded on Kentucky Wonder [7] followed by SVM-1 (6) whereas, other 3 varieties viz. Contender, Hans and Laxmi were colonized by 4 species each. Aspergillus, Cephalosporium, Cladosporium, Penicillium and Rhizopus were the dominating storage fungi whereas, Colletotrichum and Rhizoctonia constituted major pathogenic flora. Overall incidence of mycoflora ranged between 4 to 64 per cent. Aspergillus niger and C. lindemuthianum were detected from 4 vegetable type varieties showing 8.0-24.0 and 32.0-48.0 per cent infestation, respectively.

Among pulse type varieties, Him-1 harboured 5 fungi viz. Aspergillus fumigatus, Colletotrichum, Penicillium, Rhizoctonia and Rhizopus sp. followed by Baspa, Kanchan and Triloki. The incidence of storage and field fungi on pulse type beans ranged between 8.0-64.0 and 8.0-48.0 per cent whereas, it was 4.0-48.0 and 16.0-48.0 per cent, respectively, on vegetable type varieties. Rhizopus was prevalent on almost all the varieties showing 8.0-64.0 per cent incidence except Jawala.

Sterilized seeds of these two types of kidney beans yielded comparatively less fungi (Table 2). Cultivar SVM-l was colonized by Alternaria, Fusarium spp., Rhizoctonia and Stemphylium showing 20.0-24.0 per cent incidence. C. lindemuthianum was detected only from variety Laxmi and Rhizoctonia was found to be prevalent on both the beans, though the incidence on pulse type was lower as compared to vegetable type. Cultivar Baspa was free of any fungal infestation. Cultivars like Kentucky Wonder and SVM-1 were the most colonized. Baspa variety of pulse type bean possessed minimum mycoflora. Rhizoctonia was prevalent on both the beans though the incidence was less on pulse type.

Table 3. Effect of isolated mycoflora on germination (%) of kidney bean seeds

Fungi	Spore/1 suspe	Spore/mycelial suspension	Culture filtrate	filtrate	Sterilized distilled water	r	period# (Days)	Effect on germinated seed	nated seed
	Contender	Kanchan	Contender	Kanchan	Contender	Kanchan		Spore/mycelial suspension	Culture filtrate
Alternaria sp.	90.0	95.0 (77.22)	85.0 (67.38)	90.0 (71.59)	95.0 (77.22)	100.0 (89.96)	∞	Discoloration of hypocotyl region	Seed and seedling rot, reddishbrownlesionscovered with creamish fungal mass
Aspergillus fumigatus	75.0 (59.99)	80.0 (63.43)	10.0 (18.37)	7.0 (15.23)	92.0 (73.62)	100.0 (89.96)	6	Extensive necrosis on cotyledons and stem	Seed rot
A. niger	93.0 (75.29)	100.0 (89.96)	10.0 (18.37)	10.0 (18.37)	93.0 (75.29)	100.0 (89.96)	6	Extensive sporulation on cotyledons, Slight necrosis on hypocotyls	Slight necrosis on hypocotyls & root rot
Cladosporium sp.	65.0 (53.71)	70.0 (56.77)	72.0 (58.09)	80.0 (63.53)	94.0 (76.66)	100.0 (89.96)	10	Root rot, Light brown patches on hypocotyls	Root rot, Light brown patches on stem
Colletotrichum lindemuthianum	55.0 (47.85)	(55.53)	42.0 (40.38)	55.0 (47.85)	95.0 (77.22)	100.0 (89.96)	7	Seed and seedling rot	Seed and seedling rot
Fusarium spp.	92.0 (73.62)	98.0 (83.41)	85.0 (67.21)	95.0 (77.22)	96.0 (78.69)	100.0 (89.96)	16	Slight necrosis on hypocotyls	Slight necrosis on hypocotyls
F. oxysporum	42.0 (40.38)	50.0 (44.98)	40.0 (39.21)	42.0 (40.38)	90.0 (71.59)	100.0 (89.96)	S	Hypocotyls having sunken light brown lesions with dark margins	Partial sunken light brown lesions with dark margins
F. solani	50.0 (44.98)	55.0 (47.85)	42.0 (40.38)	50.0 (44.98)	92.0 (73.62)	100.0 (89.96)	10	Inhibition of seed germination and radical necrosis	Inhibition of seed germination
Penicillium	90.0	100.0 (89.96)	7.0 (15.23)	9.0 (17.38)	93.0 (75.29)	100.0 (89.96)	_	Seed rot	Seed rot
P. griseo-fulvum		99.0 (85.34)	10.0 (18.37)	12.0 (20.21)	95.0 (77.22)	100.0 (89.96)	6	No effect	Seed rot
P. simplicissimum		98.0 (83.41)	10.0 (18.37)	8.0 (16.34)	95.0 (77.22)	100.0 (89.96)	12	No effect	Seed rot
P. perpuragenum		100.0 (89.96)	8.0 (16.34)	10.0 (18.37)	95.0 (77.22)	100.0 (89.96)	∞	No effect	Seed rot
Rhizoctonia solani		55.0 (47.85)	40.0 (39.21)	42.0 (40.38)	90.0 (71.59)	100.0 (89.96)	10	Decreased seed germination	Stem necrosis and root rot
Rhizopus stolonifer		98.0 (83.41)	93.0 (75.29)	97.0 (80.42)	92.0 (73.62)	100.0 (89.96)	15	Root necrosis (traces)	Root necrosis (traces)
Stemphylium sp	abo tine	92.0 (73.62)	80.0 (63.43)	90.0 (71.59)	95.0 (77.22)	100.0 (89.96)	10	Discoloration of hypocotyl region	Seed and seedling rot, reddish brown lesions covered with creamish fungal mass
Trichoderma sp.	82.0 (65.01)	93.0 (75.29)	80.0 (63.43)	95.0 (77.22)	90 (71.59)	100.0 (89.96)	~	No effect	No effect
CD(0.05)	4.58	3.22	4.24	5.52	NS	SN	1.66		

Values in parentheses are arcsign transformed values
Incubation period depicts the appearance of fungal species

The occurrence of more mycoflora on vegetable types of beans may be due to soft seeds and tender nature of the plants. More carbohydrate content in vegetable type varieties may be ascribed for more mycoflora in these varieties. Similar observations were recorded on vegetable type beans like Contender, Kentucky Wonder and SVM 1 by Gupta et al [10, 11].

After six months of storage it was observed that seed germination was higher in the sterilized seeds of both types of beans. The germination percentage ranged between 85.33-90.67 % and 88.00-94.00 % in unsterilized and sterilized seeds of vegetable type beans, however, it ranged between 70.00-95.32 % and 85.00-98.65 % among pulse type varieties which was higher than that of recommended standards in both unsterilized and sterilized seeds in two types of beans. Occurrence of mycoflora on bean seeds though had little effect on seed germination but could affect the seed adversely under field condition after sowing.

Seed smearing with fungal spore suspension was found to reduce the germination by 5.0-55.0 per cent after 7-13 days of incubation and with culture filtrates 4.0-48.0 per cent reduction was observed with pulse type variety. However, these values were much lower in vegetable type variety. Treated seeds showed blight, necrosis, seed/seedling rot and several other abnormalities (Table 3). It was also noticed that spore suspension of storage fungi had least effect on seed germination. However, culture filtrates of these fungi reduced the germination drastically along with induction of variable symptoms. Chisholm [12] observed maximum reduction in seed germination of three types of bean by culture filtrates of A. flavus, A. niger and A. terrus. Similar type of seed and seedling abnormalities in number of legume crops due to these fungi have also been reported by many workers [13-17].

REFERENCES

 KINGRA, I.S. & C. SINGH (1986). Agricultural development. In: Agricultural Research and Development in Himachal Pradesh, eds. N.J.N. Nayer and S.C. Verma, pp. 11-24. Government Press, Shimla.

- CHRISTENSEN, C.M. & H.H. KAUFMANN (1955). Loss of viability in storage microflora, Seed Sci. & Technol. 1: 47-562.
- NEERGAARD, P. (1977). Seed Pathology. Vol I. The Macmillon Press, London. 741p.
- DHINGRA, O.D. & L.A. MAFFIA (1978). Acetone as a fungicide carrier in dormant snap bean seeds. Fitopathol. Bras. 3: 267-270.
- DHYANI, A.P., M.C. SATI, & R.D. KHULBE (1989). Phoma medicaginis Malbr. And Roum, a new record of fungi from bean crop of Kumaun Himalaya. Madras Agric. J. 76: 717-718.
- ISTA (1985). International Rules for Seed Testing, Rules 1985. Seed Sci. & Technol. 13:299-355.
- 7. LOKHANDE, S.B., W.D. MORE, & P.A. SHINDE (1986). Fungi associated with common beans. J. Maharashtra Agric. Univ. 11: 275-278.
- REDDY, N.P.E. & K.C.B. CHAUDHARY (1990). Effect of culture filtrate of Fusarium udum on the seeds of pigeon pea. Madras Agric. J. 77: 109-111.
- USHAMALINI, C. RAJAPPAN, R.& K. MANICKAM (1998). Effect of seed-borne fungi on germination, root and shoot length and vigour index of cowpea. Pl. Dis. Res. 13:109-114.
- 10. GUPTA, S.K., DOHROO, N.P. & K.R. SHYAM (1991). Antagonistic studies on seed borne mycoflora of French bean (*Phaseolus vulgaris* L.). *Indian J. of Pl. Pathol* 9: 62-63.
- 11. GUPTA, S.K., MATHEW, K.A. & K.R. SHYAM (2000). Seed transmission of *Rhizoctonia solani* in French bean cultivars. *Pl. Dis. Res.* 15: 83-84.
- 12. CHISHOLM, F.V. & P.L. COATES-BECKFORD (1997). Fungi associated with seeds of three legume species in Jamaica and seed germination at harvest and after storage. *Trop. Agric.* 74: 121-127.
- GIBSON, I.A.S. & P.K.S. CLINTON (1953). Pre-emergence seed bed losses in groundnut at Urambo Tanganyika Territory, Empire. J. Exp. Agri. 21: 226-235.
- CHOHAN, J.S. & V.K. GUPTA (1968). Aflaroot, a new disease of groundnut caused by Aspergillus flavus Link. Indian J. Agric. Sci. 38: 568-570.
- CLINTON, P.K.S. (1960). Seed bed pathogens of groundnut in the Sudan and an attempt at control with an artificial testa. *Empire J. Expt. Agri.* 28: 211-222.
- CHOHAN, J.S. (1971). Seed and soil borne fungi, seed rot and seedling disease of groundnut. *Labdev J. Sci. & Technol.* 9: 10-16.
- RAO, M.B., S. PANKAJA, H.S. PRAKASH & H. SHEKARA SHETTY (1985). Seed borne Aspergilus species of French bean and their significance. Seeds and Farms 11: 49-52.